

Annex B: Horizon scanning reference lists and topic scope

The below reference lists describe all publications identified by the Executive relevant to the horizon scanning topic. A high-level summary of the topic scope is included below each topic subheading.

Unless a paper on the topic was discussed by the SCAAC in 2024, literature searches covered the period between 1st January 2024 – 31st December 2024. For topics brought to 2024 meetings, literature searches were limited to the period since the topic was discussed. Where a new topic has been introduced, literature published over the previous ten years has been included.

The Executive notes that the identified literature is not evaluated for validity.

Alternative methods to derive embryonic and embryonic-like stem cells

Scope: This topic is focused on monitoring the methods used to create, refine and maintain human pluripotent stem cells (induced and embryonic) to ensure that the use of viable embryos in research is justified and able to fulfil the criteria of being 'necessary and desirable'. Publications which describe the establishment of novel human pluripotent stem cell lines, optimisation of methods, or the evaluation of human pluripotent stem cells will be included. Most recently this has included populations of expanded and extended potential stem cells (EPSCs), eight-cell like cells and methods to derive extraembryonic cell lineages. The scope of this topic does not extend to the development of stem-cell based embryo models or in vitro derived gametes (separate topics). Research into differentiation and application of human stem cells are excluded.

A. Ababneh, N., Barham, R., Al-Kurdi, B., Al Hadidi, S., Ali, D., Abdulelah, A. A., Madadha, A., Masri, A., & Awidi, A. (2024). Establishment of a human induced pluripotent stem cell (iPSC) line (JUCTCi018-A) from a patient with Charcot-Marie-Tooth disease type 2EE (CMT2EE) due to a homozygous c.122G > A p.(Arg41Gln) mutation in the MPV17 gene. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103602>

Afshar-Saber, W., Chen, C., Teaney, N. A., Kim, K., Yang, Z., Gasparoli, F. M., Ebrahimi-Fakhari, D., Buttermore, E. D., Pin-Fang Chen, I., Pearl, P. L., & Sahin, M. (2024). Generation and characterization of six human induced pluripotent stem cell lines (hiPSCs) from three individuals with SSADH Deficiency and CRISPR-corrected isogenic controls. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103424>

Ahmad, I., Kamai, A., Zahra, S., Kapoor, H., Kumar Srivastava, A., & Faruq, M. (2024). Generation and characterization of iPSC lines from Friedreich's ataxia patient (FRDA) with GAA.TTC repeat expansion in the Frataxin (FXN) gene's first intron (IGIBi016-A) and a non-FRDA healthy control individual (IGIBi017-A). *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103382>

Al Delbany, D., Ghosh, M. S., Krivec, N., Huyghebaert, A., Regin, M., Duong, M. C., Lei, Y., Sermon, K., Olsen, C., & Spits, C. (2024). De Novo Cancer Mutations Frequently Associate with Recurrent Chromosomal Abnormalities during Long-Term Human Pluripotent Stem Cell Culture. *Cells*, 13(16). <https://doi.org/10.3390/CELLS13161395>

Ali Gideon, S., Sabine, R., Hanna, E., Clarissa, Z., Daniela, H., Wiebke, M., Alexandra, B., Lukas, C., & Katrin, S.-B. (2024). Generation of a heterozygous Calsequestrin 2 F189L iPSC line (UMGi158-B) by CRISPR/Cas9 genome editing to investigate the cardiac pathophysiology of Takotsubo Syndrome and Catecholaminergic Polymorphic Ventricular Tachycardia. *Stem Cell Research*, 81, 103538. <https://doi.org/10.1016/J.SCR.2024.103538>

Alowaysi, M., Baadhaim, M., Al-Shehri, M., Alzahrani, H., Badkok, A., Attas, H., Zakri, S., Alameer, S., Malibari, D., Hosawi, M., Daghestani, M., Al-Ghamdi, K., muharrag, M., Zia, A., Tegne, J., Alfadhel, M., Aboalola, D., & Alsayegh, K. (2024). Derivation of two iPSC lines (KAIMRCi004-A, KAIMRCi004-B) from a Saudi patient with Biotin-Thiamine-responsive Basal Ganglia Disease (BTBGD) carrying homozygous pathogenic missense variant in the SCL19A3 gene. *Human Cell*, 37(5), 1567–1577. <https://doi.org/10.1007/S13577-024-01097-4>

Alsalloum, A., Shefer, K., Bogdanov, P., Mingaleva, N., Kim, A., Feoktistova, S., Mityaeva, O., & Volchkov, P. (2024). Establishment of a human induced pluripotent stem cell line (ABi004-A) carrying a compound

heterozygous mutation in the KCNV2 gene. *Stem Cell Research*, 80.
<https://doi.org/10.1016/J.SCR.2024.103512>

Amprou, A., Yacoub, T. Ben, Letellier, C., Degaetano, V., Méjécase, C., Pormehr, L. A., Condroyer, C., Slembrouck-Brec, A., Wohlschlegel, J., Goureau, O., Zeitz, C., & Audo, I. (2024). Generation of human induced pluripotent stem cell lines from a subject with UBAP1L-associated retinal dystrophy and CRISPR/cas9-corrected isogenic iPSC lines. *Stem Cell Research*, 81, 103558.
<https://doi.org/10.1016/J.SCR.2024.103558>

Atash, A., Cramer, M. J., Mees, B., Doevendans, P. A., Sluijter, J. P. G., & Stillitano, F. (2025). Generation and characterization of novel induced pluripotent stem cell (iPSC) lines derived from three symptomatic carriers of a pathogenic MYH11 variant and two non-carrier relatives. *Stem Cell Research*, 82.
<https://doi.org/10.1016/J.SCR.2024.103630>

Baggiani, M., Santorelli, F. M., Mero, S., Privitera, F., Damiani, D., & Tessa, A. (2024). Generation of a human induced pluripotent stem cell line (FSMi001-A) from fibroblasts of a patient carrying heterozygous mutation in the REEP1 gene. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103472>

Bayarsaikhan, D., Bayarsaikhan, G., Kang, H. A., Lee, S. Bin, Han, S. H., Okano, T., Kim, K., & Lee, B. (2024). A Study on iPSC-Associated Factors in the Generation of Hepatocytes. *Tissue Engineering and Regenerative Medicine*, 21(8). <https://doi.org/10.1007/S13770-024-00674-W>

Bayarsaikhan, D., Yoo, D. H., Lee, J., Im, Y. S., Bayarsaikhan, G., Kang, H. A., Kim, Y. O., & Lee, B. (2024). Generation and characterization of GATA6-specific EGFP expressing human induced pluripotent stem cell line, KSCBi017-A-1, using CRISPR/Cas9. *Stem Cell Research*, 77.
<https://doi.org/10.1016/J.SCR.2024.103426>

Bekhite, M. M., Hübner, S., Kretzschmar, T., Backsch, C., Weise, A., Klein, E., Bogoviku, J., Westphal, J., & Christian Schulze, P. (2024). Generation of human induced pluripotent stem cell lines UKJi001-A and UKJi006-A from patients with heterozygous mutation in the PKP2 gene. *Stem Cell Research*, 81.
<https://doi.org/10.1016/J.SCR.2024.103565>

Bekhite, M. M., Hübner, S., Kretzschmar, T., Backsch, C., Weise, A., Klein, E., Bogoviku, J., Westphal, J., & Schulze, P. C. (2025). Generation of a human induced pluripotent stem cell lines (UKJi003-A) from a patient with Fabry disease and healthy donor (UKJi004-A). *Stem Cell Research*, 82.
<https://doi.org/10.1016/J.SCR.2024.103620>

Bianchini, L., Sieber, L., Hammad, R., Schäfer, R., & Kutscher, L. M. (2024). Generation of two isogenic patient-derived human-induced pluripotent stem cell clones with 6q27 deletion. *Stem Cell Research*, 80.
<https://doi.org/10.1016/J.SCR.2024.103524>

Boullé, M., Leleu, A., Schacre, S., Banal, C., Boucharlat, A., Renault, S., Hollenstein, M., Frosk, P., Yates, F., Lefort, N., & Agou, F. (2024). Generation of IPi002-A/B/C human induced pluripotent stem cell lines from MARCH amniotic fluid cells. *Stem Cell Research*, 81, 103589.
<https://doi.org/10.1016/J.SCR.2024.103589>

Bouwman, L. F., Joosen, M. E. M., Buijsen, R. A. M., van der Graaf, L. M., Pepers, B. A., Voeselek, B. J. B., Brosens, E., van de Warrenburg, B. P. C., & van Roon-Mom, W. M. C. (2024). Generation of human induced pluripotent stem cell lines (LUMCi051-A,B and LUMCi052-A,B,C) of two patients with Spinocerebellar ataxia type 7. *Stem Cell Research*, 78. <https://doi.org/10.1016/J.SCR.2024.103462>

Caballano Infantes, E., Clauzon, L., de la Cerda Haynes, B., & Díaz-Corrales, F. (2025). Generation of the human iPSC line ESi132-A from a patient with retinitis pigmentosa caused by a mutation in the PRPF31 gene. *Stem Cell Research*, 82, 103623. <https://doi.org/10.1016/J.SCR.2024.103623>

Celiker, C., Zelenak, S., Lietava, S., Pachernik, J., Bebarova, M., Zidkova, J., Novotny, T., & Barta, T. (2024). Generation of human induced pluripotent stem cell lines from patients with a RYR2 gene variant c.14201A>G (p.Y4734C): Implications for idiopathic ventricular fibrillation and catecholaminergic polymorphic ventricular tachycardia. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103541>

Chandrasekaran, V., Wellens, S., Bourguignon, A., Djidrovski, I., Fransen, L., Ghosh, S., Mazidi, Z., Murphy, C., Nunes, C., Singh, P., Zana, M., Armstrong, L., Dinnyés, A., Grillari, J., Grillari-Voglauer, R., Leonard, M. O., Verfaillie, C., Wilmes, A., Zurich, M. G., ... Culot, M. (2024). Evaluation of the impact of

- iPSC differentiation protocols on transcriptomic signatures. *Toxicology in Vitro: An International Journal Published in Association with BIBRA*, 98. <https://doi.org/10.1016/J.TIV.2024.105826>
- Chen, G., Bai, R., Huang, P., Liu, X., Ni, J., & Chen, L. (2024). Generation of a homozygous DNAJC19 knockout human embryonic stem cell line by CRISPR/Cas9 system. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103427>
- Chen, J., Dai, T., Li, Q., Xu, T., Zhang, W., Sun, J., & Liu, H. (2024). Generation of FOXJ1-EGFP knock-in reporter human embryonic stem cell line, WAe001-A-2D, using CRISPR/Cas9-based gene targeting. *Stem Cell Research*, 78. <https://doi.org/10.1016/J.SCR.2024.103445>
- Chen, X., Sun, J., Wang, T., Tang, Q., Su, L., Sun, Y., Chen, L., Seo, H., Cheng, T., Wang, J., & Song, B. (2024). Generation of a human iPSC line from a Parkinson's disease patient with a novel CHCHD2 mutation (p.R145Q). *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103419>
- Chen, Y., Ye, X., Zhong, Y., Kang, X., Tang, Y., Zhu, H., Pang, C., Ning, S., Liang, S., Zhang, F., Li, C., Li, J., Gu, C., Cheng, Y., Kuang, Z., Qiu, J., Jin, J., Luo, H., Fu, M., ... Gao, X. (2024). SP6 controls human cytotrophoblast fate decisions and trophoblast stem cell establishment by targeting MSX2 regulatory elements. *Developmental Cell*, 59(12), 1506-1522.e11. <https://doi.org/10.1016/J.DEVCEL.2024.03.025>
- Chen, Z., Yu, X., Ke, M., Li, H., Jiang, Y., Zhang, P., Tan, J., Cao, N., & Yang, H. T. (2024). Human embryonic stem cell-derived cardiovascular progenitor cells stimulate cardiomyocyte cell cycle activity via activating the PI3K/Akt pathway. *Journal of Molecular and Cellular Cardiology*, 197, 5–10. <https://doi.org/10.1016/J.YJMCC.2024.10.002>
- Chiew, M. Y., Wang, E., Lan, K. C., Lin, Y. R., Hsueh, Y. H., Tu, Y. K., Liu, C. F., Chen, P. C., Lu, H. E., & Chen, W. L. (2024). Improving iPSC Differentiation Using a Nanodot Platform. *ACS Applied Materials & Interfaces*, 16(28), 36030–36046. <https://doi.org/10.1021/ACSAMI.4C04451>
- Cho, Y. K., Kim, H. K., Suh, E. J., Kim, H. O., & Kim, S. (2024). Generation of a human induced pluripotent stem cell line (YUCMi020-A) from peripheral blood mononuclear cells derived from a female with the Jr(a-) blood type. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103434>
- Clayton, J. S., Vo, C., Crane, J., Scriba, C. K., Saker, S., Larmonier, T., Malfatti, E., Romero, N. B., Ravenscroft, G., Laing, N. G., & Taylor, R. L. (2024a). Generation of iPSC lines from three Laing distal myopathy patients with a recurrent MYH7 p.Lys1617del variant. *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103491>
- Clayton, J. S., Vo, C., Crane, J., Scriba, C. K., Saker, S., Larmonier, T., Malfatti, E., Romero, N. B., Ravenscroft, G., Laing, N. G., & Taylor, R. L. (2024b). Generation of two iPSC lines from adult central core disease patients with dominant missense variants in the RYR1 gene. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103411>
- Clayton, J. S., Vo, C., Crane, J., Scriba, C. K., Saker, S., Larmonier, T., Malfatti, E., Romero, N. B., Ravenscroft, G., Laing, N. G., & Taylor, R. L. (2024c). Generation of two iPSC lines from patients with inherited central core disease and concurrent malignant hyperthermia caused by dominant missense variants in the RYR1 gene. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103410>
- Devito, L. G., Lim, E. S., O'Toole, S. M., Shepherd, S. T. C., Deng, D., Feng, H., Barber, T., Drake, W. M., Turajlic, S., & Healy, L. (2024). Generation of TWO iPSC lines (CRICKi009-A; CRICKi010-A) from patients with type 1 von Hippel-Lindau (VHL) and histopathologically confirmed renal cell carcinoma (RCC). *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103611>
- Dexheimer, R., Manhas, A., Wu, D., Tripathi, D., Yu Chan, S., Li, J., Yu, R., Sayed, N., Wu, J. C., & Sallam, K. (2024). Generation of two iPSC lines from dilated cardiomyopathy patients with pathogenic variants in the SCN5A gene. *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103498>
- Edwards, S., Hagenau, L., Nowack, B., Rhode, J., Hossain, M. F., Tzvetkova, A., Jensen, L. R., & Kuss, A. W. (2024). Generation of two isogenic iPSC lines from a healthy male donor of European ancestry. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103403>
- Erlindri, I., Sangotra, A., Keller, L., Lieberman, A. P., & Smith, G. D. (2024). A spinal and bulbar muscular atrophy (SBMA) disease-specific human embryonic stem cell (hESC) line, UMICHe002-A/UM197-1. *Stem Cell Research*, 81, 103548. <https://doi.org/10.1016/J.SCR.2024.103548>

- Filippi, K., Riße, I., Judge, L. M., Conklin, B. R., Fleischmann, B. K., & Hesse, M. (2025). Generation and characterization of an isogenic control line by correcting the BAG3 P209L mutation of a human induced pluripotent stem cell (hiPSC) line from a patient with myofibrillar myopathy-6. *Stem Cell Research*, 82. <https://doi.org/10.1016/J.SCR.2024.103626>
- Filippi, K., Wiemann, M., Fleischmann, B. K., & Hesse, M. (2025). Generation of two isogenic control lines by correcting the BAG3 P209L mutation of human induced pluripotent stem cell (hiPSC) lines from patients with myofibrillar myopathy-6. *Stem Cell Research*, 82, 103627. <https://doi.org/10.1016/J.SCR.2024.103627>
- Frederiksen, H. R. S., Skov, S., Tveden-Nyborg, P., Freude, K., & Doehn, U. (2024). Novel traceable CRISPR-Cas9 engineered human embryonic stem cell line (E1C3 + hSEAP + 2xKO + pCD47), has potential to evade immune detection in pigs. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103438>
- Gao, M., Li, X., Lv, Y., Yang, X., Liu, Y., & Gai, Z. (2024). Generation and characterization of an induced pluripotent stem cell (iPSC) line SDQLCHi063-A from peripheral blood mononuclear cells of a patient with Maturity-onset diabetes of the young type 2 carrying GCK exon 1 deletion. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103389>
- Ge, N., Suzuki, K., Sato, I., Noguchi, M., Nakamura, Y., Matsuo-Takasaki, M., Fujishiro, J., & Hayashi, Y. (2024). Generation of human induced pluripotent stem cell lines derived from patients of cystic biliary atresia. *Human Cell*, 38(1). <https://doi.org/10.1007/S13577-024-01147-X>
- Giovenale, A. M. G., Turco, E. M., Mazzoni, M., Ferrone, I., Torres, B., Bernardini, L., Vulcano, E., Ferrari, D., Onesimo, R., D'Arrigo, S., Zampino, G., Pennuto, M., De Luca, A., Vescovi, A. L., & Rosati, J. (2024). Generation of the CSSi020-A (14437) iPSC line from a patient carrying a copy number variation (CNV) in the 17p11.2 chromosome region. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103544>
- Gong, T., Liu, D., Wang, X., Zhou, D., Tang, L., Wang, H., Su, J., & Liang, P. (2024). Establishment of a CIB1 knockout human pluripotent stem cell line via CRISPR/Cas9 genome editing technology. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103580>
- Guo, X., Zhang, M., Xu, Z., Yu, Y., & Shen, Z. (2024). CRISPR/Cas9-mediated generation of AP-1 activity reporter cell line in human embryonic stem cell (WAe007-A-5). *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103557>
- Guo, X., Zhao, K., Zhang, Y., Zhou, T., & Pan, G. (2024). Generation of a PPM1A-deficient human induced pluripotent stem cell line using CRISPR-Cas9 technology. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103420>
- Haake, J., Kaufmann, M., & Steenpass, L. (2024). Generation of three iPSC lines with inducible systems to be used in Angelman syndrome research. *Stem Cell Research*, 78. <https://doi.org/10.1016/J.SCR.2024.103454>
- Haferkamp, U., Telugu, N., Krieg, K., Schaefer, W., Lam, D., Binkle-Ladisch, L., Friese, M. A., Diecke, S., & Pless, O. (2024). Generation of two isogenic human iPSC lines (ZiPi013-B-1, ZiPi013-B-2) carrying a CRISPR/Cas9-mediated deletion of TRPM4. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103609>
- Haidar, M., Schmid, B., Ruiz, A., Ebneith, A., & Cabrera-Socorro, A. (2024). Generation of three isogenic, gene-edited iPSC lines carrying the APOE-Christchurch mutation into the three common APOE variants: APOE2Ch, APOE3Ch and APOE4Ch. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103414>
- Hernández, D., Morgan Schlicht, S., Elli Clarke, J., Daniszewski, M., Karch, C. M., Adams, S., Allegri, R., Araki, A., Barthelemy, N., Bateman, R., Bechara, J., Benzinger, T., Berman, S., Bodge, C., Brandon, S., (Bill) Brooks, W., Brosch, J., Buck, J., Buckles, V., ... Pébay, A. (2024). Generation of a gene-corrected human isogenic iPSC line from an Alzheimer's disease iPSC line carrying the PSEN1 H163R mutation. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103495>
- Hua, C., Sun, W., Zhang, C., Tian, X., Qin, X., Dong, J., & Li, X. (2024). Generation of a human induced pluripotent stem cell line ZZUNEUi030-A from a female patient carrying a heterozygous CALM2 (c.395 A > T) mutation. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103515>

- Jacobs, E. H., Schatzman Raposo, J., Scardamaglia, A., Alkuraya, F. S., Nafissi, S., Houlden, H., Zuchner, S., & Saporta, M. A. (2024). Establishment and characterization of three human pluripotent stem cell lines from Charcot-Marie-Tooth disease Type 4B3 patients bearing mutations in *MTMR5/Sbf1* gene. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103599>
- Jahn, C., Juchem, M., Sonnenschein, K., Gietz, A., Buchegger, T., Lachmann, N., Göhring, G., Behrens, Y. L., Bär, C., Thum, T., & Hoepfner, J. (2024). Generation of human induced pluripotent stem cell line MHHi029-A from a male Fabry disease patient carrying c.959A > T mutation. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103404>
- Jiamvoraphong, N., Lorthongpanich, C., Klaihmon, P., Kheolamai, P., & Issaragrisil, S. (2024). Generation of a human iPSC cell line (MUSli017-A) from a donor with O negative blood type. *Stem Cell Research*, 78. <https://doi.org/10.1016/J.SCR.2024.103466>
- Jiang, L., Tracey, T. J., Gill, M. K., Howe, S. L., Power, D. T., Bharti, V., McCombe, P. A., Henderson, R. D., Steyn, F. J., & Ngo, S. T. (2024). Generation of human induced pluripotent stem cell lines from sporadic, sporadic frontotemporal dementia, familial *SOD1*, and familial *C9orf72* amyotrophic lateral sclerosis (ALS) patients. *Stem Cell Research*, 78. <https://doi.org/10.1016/J.SCR.2024.103447>
- Jiang, M., Long, Y., Liu, Y., Li, R., Xu, J., Chi, T., Sun, M., Li, M., Yu, H., Yu, L., & Han, X. (2024). Establishment of an induced pluripotent stem cell (iPSC) line (JNMUi002-A) from a male patient with schizophrenia. *Stem Cell Research*, 81, 103570. <https://doi.org/10.1016/J.SCR.2024.103570>
- Jiang, S., Dai, T., Li, Q., Xu, T., Zhang, W., Sun, J., & Liu, H. (2024). Generation of ASCL1-mCherry knock-in reporter in human embryonic stem cell line, WAe001-A-2E, using CRISPR/Cas9-based gene targeting. *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103500>
- Jiang, X., Fu, C., Liu, Q., & Gao, J. (2024). Generation of a *KCNQ1* (c.1032 + 2 T > C) mutant human embryonic stem cell line via CRISPR base editing. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103425>
- Jiang, X., Liu, Q., Yang, L., Zhang, X., Gao, J., & Jiang, Y. (2024). Generation of a *MYH6* (c.4034T > C) mutant human embryonic stem cell line via CRISPR base editing. *Stem Cell Research*, 81, 103610. <https://doi.org/10.1016/J.SCR.2024.103610>
- Jin, G., Huang, H., Bao, X., & Palecek, S. P. (2024). Poly(norepinephrine)-Mediated Universal Surface Modification for Patterning Human Pluripotent Stem Cell Culture and Differentiation. *ACS Biomaterials Science & Engineering*, 10(12). <https://doi.org/10.1021/ACSBOMATERIALS.4C01229>
- Joseph, B., Varea, I., Emmerich, K., Manohar-Sindhu, S., Zou, J., Friend, K., Sipwoli, C., Tang, X., Yang, D., de Jesus Rasheed, A. A., Goldbach-Mansky, R., & Boehm, M. (2024). Establishment of a human induced pluripotent stem cell line (NIHTVBi031-A) derived from a COPA syndrome patient with a heterozygous p.Ala239Pro mutation. *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103504>
- Jyoti Saikia, B., Bhardwaj, J., Saini, A., Rajan, R., & B.K, B. (2024). Generation of an induced pluripotent stem cell (iPSC) line (IGiBi026-A) derived from Wilson disease patient harboring compound heterozygous mutations [c.2165dupT (p.R723Efs31) and c.C813A (p.C271*)] in the *ATP7B* gene. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103567>
- Kelters, I. R., Verbueken, D., Beekink, T., Van Laake, L. W., Sluijter, J. P. G., Maas, R. G. C., & Buikema, J. W. (2024). Generation of human induced pluripotent stem cell (hiPSC) lines derived from three patients carrying the pathogenic *CRYAB* (A527G) mutation and one non-carrier family member. *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103497>
- Khiami, M., Ju, Y., Han, L., Klein, J., Han, M. J., Pruett-Miller, S. M., & Wlodarski, M. W. (2024). Generation of CRISPR/Cas9-edited human iPSC lines carrying homozygous and heterozygous *SAMD9* p.I983S mutations. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103532>
- Krivec, N., de Deckersberg, E. C., Lei, Y., Delbany, D. Al, Regin, M., Verhulst, S., van Grunsven, L. A., Sermon, K., & Spits, C. (2024). Gain of 1q confers an *MDM4*-driven growth advantage to undifferentiated and differentiating hESC while altering their differentiation capacity. *Cell Death & Disease*, 15(11), 852. <https://doi.org/10.1038/S41419-024-07236-X>

- Krogulec, E., Dobosz, A. M., Liszewska, E., Majchrowicz, L., & Dobrzyń, A. (2024). Generation of four human induced pluripotent stem cell lines derived from patients with MPAN, subtype of NBIA, carrying the c.204_214del11 mutation in the C19orf12 gene. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103603>
- Leavens, K. F., Osorio-Quintero, C., Yeuteuh, E. A., Perez-Profeta, F. T., Dattoli, A. A., Cardenas-Diaz, F. L., French, D. L., & Gadue, P. (2024). Generation of a fluorescent mNeonGreen insulin reporter line in the H1 (WA01) hESC background. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103559>
- Lee, Y., Park, H.-J., Kim, Y.-O., & Kim, B.-Y. (2024). Generation of a human induced pluripotent stem cell line (KSCBi016-A) from a CPVT patient with an RYR2 mutation. *Stem Cell Research*, 81, 103560. <https://doi.org/10.1016/J.SCR.2024.103560>
- Léger, L., Aalders, J., Heymans, N., Van Acker-Verberckt, K., De Bleekere, L., Coucke, P., Menten, B., Bauce, B., Vitiello, L., Rampazzo, A., Calore, M., & van Hengel, J. (2024). Generation of a human induced pluripotent stem cell line UGENTI002-A from an arrhythmogenic cardiomyopathy patient carrying the c.817C>T DSP heterozygous variant and isogenic control using CRISPR/Cas9 editing. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103537>
- Li, B., Ye, X., Lin, J., Ma, K., He, M., & Fang, Y. (2024). Generation of a human induced pluripotent stem cell line (SHUPLi002-A) from PBMCs of a healthy female donor. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103422>
- Li, R., Tsuboi, H., Ito, H., Takagi, D., Chang, Y.-H., Shimizu, T., Arai, Y., Matsuo-Takasaki, M., Noguchi, M., Nakamura, Y., Ohnuma, K., Takahashi, S., & Hayashi, Y. (2024). Generation of human induced pluripotent stem cell lines derived from two glucose transporter 1 deficiency syndrome patients. *Stem Cell Research*, 81, 103584. <https://doi.org/10.1016/J.SCR.2024.103584>
- Li, S. J., Mei, L., He, C., Cai, X., Wu, H., Wu, X. W., Liu, Y., Feng, Y., & Song, J. (2024). Identification of a family with van der Hoeve's syndrome harboring a novel COL1A1 mutation and generation of patient-derived iPSC lines and CRISPR/Cas9-corrected isogenic iPSCs. *Human Cell*, 37(3), 817–831. <https://doi.org/10.1007/S13577-024-01028-3>
- Liang, Y., Sun, X., Chen, H., Cui, Z., Gu, J., Duan, C., Mao, S., Chen, Y., Li, X., Xiong, S., & Chen, J. (2024). CRISPR/Cas9-mediated generation of a human induced pluripotent stem cell line with PRPF6 c.2699 G > A mutation to model retinitis pigmentosa. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103581>
- Lim, S. W., Lee, K. I., Cui, S., Fang, X., Shin, Y. J., Lee, H., Lee, J. Y., Chung, B. H., & Yang, C. W. (2024). Generation of green fluorescent protein reporter knock-in iPSC line at the 3'UTR region of the KLOTTHO locus. *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103499>
- Liu, D., Shen, J., Yang, Z., Fan, H., Wang, H., Liang, P., & Gong, T. (2024). Generation of a lamin A/C knockout human induced pluripotent stem cell line (ZJULLi007-A) via CRISPR/Cas9. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103579>
- Liu, H. B., Dong, T., Deng, L., Zhou, C., Tang, F., Margolis, R. L., & Li, P. P. (2024). Generation of a human induced pluripotent stem cell line JHUi004-A with heterozygous mutation for spinocerebellar ataxia type 12 using genome editing. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103441>
- Liu, J., Du, J., Ma, X., Jin, Y., Yang, Q., Zhai, Y., Cheng, J., Luan, F., Ma, M., Zhang, Z., Ren, Q., & Cui, H. (2024). Generation of an iPSC line from a 79-year-old female patient diagnosed with sporadic Parkinson's disease. *Stem Cell Research*, 78. <https://doi.org/10.1016/J.SCR.2024.103450>
- Liu, W., Wang, Y., Yang, Y., Wang, Y., Tang, Y., Jiao, Y., Shan, D., Zhan, Z., Zhang, R., Wang, D., Sun, X., Sun, P., Sun, X., Yan, C., & Liu, F. (2024). Establishment of an induced pluripotent stem cell (iPSC) line (INNDUi005-A) from a healthy female Chinese Han. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103386>
- Lu, S., Chen, M., Liu, X., Li, J., Liu, H., & Li, S. (2025). Generation of a BEST1 Pr-EGFP reporter human embryonic stem cell line via CRISPR/Cas9 editing. *Stem Cell Research*, 82. <https://doi.org/10.1016/J.SCR.2024.103625>

- Luo, Y., Zheng, W., Zhong, Y., Liu, H., Yu, J., Qiu, B., Liu, J., & Yang, B. (2024). Generation of a human embryonic stem cell line (SMUDHe010-A-1A) carrying Brainbow cassette in the AAVS1 gene by CRISPR/Cas9-mediated homologous recombination. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103383>
- Lynch, A. T., Douglas, M., Kimber, S. J., & Birket, M. J. (2024). The generation and validation of a dual cardiac HAND1-Tomato NKX2-5-GFP human embryonic stem cell line UMANe002-A-3. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103342>
- Manhas, A., Tripathi, D., Noishiki, C., Wu, D., Liu, L., Sallam, K., Lee, J. T., Fukaya, E., & Sayed, N. (2024). Generation of two iPSC lines from vascular Ehlers-Danlos Syndrome (vEDS) patients carrying a missense mutation in COL3A1 gene. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103485>
- Mengnan, W., Yan, C., Qiong, X., & Man, X. (2024). Generation of a human induced pluripotent stem cell line (FDIBSi001-A) from a patient with ADNP syndrome carrying ADNP mutation (c. 2059 T>C). *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103550>
- Mori, M., Yoshii, S., Noguchi, M., Takagi, D., Shimizu, T., Ito, H., Matsuo-Takasaki, M., Nakamura, Y., Takahashi, S., Hamada, H., Ohnuma, K., Shiohama, T., & Hayashi, Y. (2024). Generation of human induced pluripotent stem cell lines derived from four Rett syndrome patients with MECP2 mutations. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103432>
- Mulero-Russe, A., Mora-Boza, A., Marquez, E. N., Ziegelski, M., Helmrath, M., & García, A. J. (2025a). Synthetic hydrogel substrate for human induced pluripotent stem cell definitive endoderm differentiation. *Biomaterials*, 315. <https://doi.org/10.1016/J.BIOMATERIALS.2024.122920>
- Mulero-Russe, A., Mora-Boza, A., Marquez, E. N., Ziegelski, M., Helmrath, M., & García, A. J. (2025b). Synthetic hydrogel substrate for human induced pluripotent stem cell definitive endoderm differentiation. *Biomaterials*, 315. <https://doi.org/10.1016/J.BIOMATERIALS.2024.122920>
- Mun, D., Kang, J.-Y., Park, M., Yoo, G., Kim, H., Yun, N., Mi Hwang, Y., & Joung, B. (2024). Establishment of a human-induced pluripotent stem cell line from a long QT syndrome type 2 patient harboring a KCNH2 mutation. *Stem Cell Research*, 81, 103592. <https://doi.org/10.1016/J.SCR.2024.103592>
- Mun, D., Yoo, G., Park, M., Kang, J. Y., Yun, N., & Joung, B. (2024). Establishment of two human induced pluripotent stem cell lines from familial long QT syndrome type 1 patients carrying KCNQ1 mutation. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103571>
- Naidoo, J., Hurrell, T., & Scholefield, J. (2024). The generation of human induced pluripotent stem cell lines from individuals of Black African ancestry in South Africa. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103534>
- Oh, J. Y., Yoo, D. H., Im, Y. S., & Kim, Y. O. (2024). Generation and characterization of a human iPSC line expressing EGFP-tagged CDH1, KSCBi002-A-3. *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103510>
- Osuna, M. A. L., Han, L., Connelly, J. P., Miller-Preutt, S., Weiss, M. J., Wlodarski, M. W., & Bhoopalan, S. V. (2024). Generation of iPSC lines and isogenic gene-corrected lines from two individuals with RPS19-mutated Diamond-Blackfan anemia syndrome. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103479>
- Pachernegg, S., Robevska, G., Ferreira, L. G. A., van den Bergen, J. A., Vlahos, K., Howden, S. E., Sinclair, A. H., & Ayers, K. L. (2024). Generation of a homozygous (MCRli031-A-3) WT1 knockout human iPSC line. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103494>
- Pachernegg, S., Robevska, G., G. A. Ferreira, L., van den Bergen, J. A., Vlahos, K., Howden, S. E., Sinclair, A. H., & Ayers, K. L. (2024). Generation of heterozygous (MCRli031-A-1) and homozygous (MCRli031-A-2) SOX9 knockout human iPSC lines. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103484>
- Pachis, S. T., Ramovs, V., Freund, C., Has, C., & Raymond, K. (2024). Generation and genetic repair of two human induced pluripotent stem cell lines from patients with Epidermolysis Bullosa simplex associated with a heterozygous mutation in the translation initiation codon of KLHL24. *Stem Cell Research*, 81, 103551. <https://doi.org/10.1016/J.SCR.2024.103551>

- Pavlova, S. V., Shulgina, A. E., Minina, J. M., Zakian, S. M., & Dementyeva, E. V. (2024). Generation of Isogenic iPSC Lines for Studying the Effect of the p.N515del (c.1543_1545delAAC) Variant on MYBPC3 Function and Hypertrophic Cardiomyopathy Pathogenesis. *International Journal of Molecular Sciences*, 25(23). <https://doi.org/10.3390/IJMS252312900>
- Popović, N. Z., Blanch-Asensio, A., Visser, T., Mummery, C. L., Davis, R. P., & Yiangou, L. (2024). Generation of two induced pluripotent stem cell (iPSC) lines carrying the Brugada Syndrome-associated mutation SCN5A-R282H. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103585>
- Pornratananont, G., Tangprasittipap, A., Hongeng, S., & Anurathapan, U. (2024). Generation of integration-free human induced pluripotent stem cell line MURAI003-A derived from the peripheral blood mononuclear cells of a donor with homozygous Class I and Class II HLAs (A*11:01, B*46:01; C*01:02; DRB1*09:01; DQB1*03:03). *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103514>
- Qin, Y., Godoy-Parejo, C., Skowronska, M., Verma, A., Dejoze, M., & Zwaka, T. P. (2024). Generation of human pluripotent stem cell lines (WAe009-A) with THAP11F80L cobalamin disorder-associated mutation. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103483>
- Rezaeiani, S., Rezaee, M., Shafaghi, M., Karami, M., Hamidi, R., Khodayari, H., Vahdat, S., Pahlavan, S., & Baharvand, H. (2024). Expandable hESC-derived cardiovascular progenitor cells generate functional cardiac lineage cells for microtissue construction. *Stem Cell Research & Therapy*, 15(1). <https://doi.org/10.1186/S13287-024-03919-6>
- Ruan, J., Wu, M., Xiang, J., Hui, X., Yang, L., Lin, R., Xu, W., & Shu, Q. (2024). Generation of a human induced pluripotent stem cell line from a female patient carrying LZTR1 gene mutation. *Stem Cell Research*, 81, 103616. <https://doi.org/10.1016/J.SCR.2024.103616>
- Sabir, M. S., Leoyklang, P., Hackbarth, M. E., Pak, E., Dutra, A., Tait, R., Pollard, L., Adams, D. R., Gahl, W. A., Huizing, M., & Malicdan, M. C. V. (2024). Generation and characterization of two iPSC lines derived from subjects with Free Sialic Acid Storage Disorder (FSASD). *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103600>
- Sareen, N., Srivastava, A., Mittal, I., Shah, A. H., & Dhingra, S. (2024). Establishment of a new human iPSC cell line (UOMI012-A) from a patient with congenital heart defect who has undergone Fontan procedure. *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103509>
- Sarkar, J., Dhepe, S., Shivalkar, A., Kuhikar, R., More, S., Konala, V. B. R., Bhanushali, P., & Khanna, A. (2024). Generation of human-induced pluripotent stem cell line from PBMC of healthy donor using integration-free Sendai virus technology. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103402>
- Selvitella, S., Franco, M. P., Aran, B., Veiga, A., & Kuebler, B. (2024). Generation of the CTRL EiPS J9 mR6F-8 iPSC line derived from healthy human outgrowth blood endothelial cells (BOECs) using mRNA reprogramming methodology. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103606>
- Semenova, P. I., Panova, A. V., Sopova, J. V., Krasnova, O. A., Turilova, V. I., Yakovleva, T. K., Kulikova, K. S., Petrova, D. A., Kiselev, S. L., & Neganova, I. E. (2024). Generation of CRISPR/Cas9 modified human iPSC line with correction of heterozygous mutation in exon 6 of the CaSR gene. *Human Cell*, 38(1). <https://doi.org/10.1007/S13577-024-01135-1>
- Sif Ásgrímsdóttir, E., & Arenas, E. (2024). Generation of a fluorescent hESC reporter line (Kle033-A-1) for the isolation of distinct midbrain progenitor cell types. *Stem Cell Research*, 81, 103523. <https://doi.org/10.1016/J.SCR.2024.103523>
- Simmons, A. D., Baumann, C., Zhang, X., Kamp, T. J., De La Fuente, R., & Palecek, S. P. (2024). Integrated multi-omics analysis identifies features that predict human pluripotent stem cell-derived progenitor differentiation to cardiomyocytes. *Journal of Molecular and Cellular Cardiology*, 196, 52–70. <https://doi.org/10.1016/J.YJMCC.2024.08.007>
- Sleiman, Y., Reisqs, J. B., Bianca Tan, R., Cecchin, F., Chahine, M., & Boutjdir, M. (2024). Generation of an iPSC cell line (VANYHHi001-A) from a patient with cardiac arrhythmias carrying CACNA1D, SCN5A, and DSP variants. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103608>

- Son, Y., Li, P., Ortega, D., Qiu, H., Prachyl, H., Yang, M., & Zhu, W. (2024). Generation of a Human Induced Pluripotent Stem Cell Line Expressing a Magnetic Resonance Imaging Reporter Gene. *Small Methods*, 8(10). <https://doi.org/10.1002/SMTD.202301764>
- Song, M., Chen, S., Zhang, M., Hu, S., Lei, W., & Yu, M. (2024). Generation of a human induced pluripotent stem cell line harboring heteroplasmic m.3243A > G mutation in MT-TL1 gene. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103387>
- Srisantitham, J., Suwanpitak, S., Thongsin, N., & Wattanapanitch, M. (2024). Generation of a homozygous TIGIT gene knockout (TIGIT^{-/-}) human iPSC line (MUSli001-A-3) using CRISPR/Cas9 system. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103601>
- Sun, H., Li, Q., Xu, T., Zhang, W., Sun, J., & Liu, H. (2024). Generation of SFTPC-mCherry knock-in reporter human embryonic stem cell line, WAe001-A-2H, using CRISPR/Cas9-based gene targeting. *Stem Cell Research*, 81, 103597. <https://doi.org/10.1016/J.SCR.2024.103597>
- Sun, X., Liang, Y., Duan, C., Liu, X., Zhou, Y., Mao, S., Cui, Z., Gu, J., Ding, C., Chen, J., & Tang, S. (2024). Generation of a gene-corrected isogenic human iPSC line (CSUASOi006-A-1) from a retinitis pigmentosa patient with heterozygous c.5792C > T mutation in the PRPF8 gene. *Stem Cell Research*, 81, 103572. <https://doi.org/10.1016/J.SCR.2024.103572>
- Takagi, D., Tsukamoto, S., Nakade, K., Shimizu, T., Arai, Y., Matsuo-Takasaki, M., Noguchi, M., Nakamura, Y., Yumoto, N., Kawada, J., Hayata, T., & Hayashi, Y. (2024). Generation of MBP-tdTomato reporter human induced pluripotent stem cell line for live myelin visualization. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103493>
- Tang, P., Keshi, E., Wilken, S., Wutsdorff, L., Mougnekabol, J., Pratschke, J., Sauer, I. M., & Haep, N. (2024). Generation of an induced pluripotent stem cell (iPSC) line (EXSURGi001-A) from a patient homozygous for the p.Ala165Thr mutation in the MTARC1 gene. *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103516>
- Tangprasittipap, A., Innachai, P., Chumchuen, S., Chiangjong, W., Jinawath, N., Sirachainan, N., & Hongeng, S. (2024). Establishment of human induced pluripotent stem cell line MURAI001-A from skin fibroblasts of a patient carrying a c.4404A > G mutation in the TET1 gene. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103474>
- Tanzer, K., Meier, B., Vulinovic, F., Pawlack, H., Klein, C., Seibler, P., & Rakovic, A. (2024). Generation of four human-derived iPSC TorsinA-3xFLAG reporter lines from a DYT-TOR1A patient. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103595>
- Tohari, M., Sanjaya, R., Yuliana Sari, S., Tedjobagaskara, B., Ibnu Faisal, A., Alvin Prawira, M., Oktaviani Dwi Putri, A., Faza, N., Murti, H., & Widyastuti, H. P. (2024). Generation of footprint-free human induced pluripotent stem cell line (SCIKFi001-B) from cGMP grade umbilical cord-derived mesenchymal stem cells (UC-MSCs) using episomal-plasmid based reprogramming approach. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103566>
- Tonin, R., Feo, F., Falliano, S., Giunti, L., Calamai, M., Procopio, E., Mari, F., Scirucchio, V., Conti, V., Fanelli, I., Bambi, F., Guerrini, R., & Morrone, A. (2024). Generation of a human induced pluripotent stem cell line from a patient with GM3 synthase deficiency using self-replicating RNA vector. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103431>
- Tran, H. D., Denman, C. R., Shin, M. K., Jeon, D., Kuhn, B., & Jo, J. (2024). Establishment of TH-EGFP human embryonic stem cell line for specific labeling of dopaminergic neurons. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103613>
- Ura, H., Togi, S., Hatanaka, H., & Niida, Y. (2024). Establishment of a human induced pluripotent stem cell line, KMUGMCi010-A, from a patient with X-linked Ohdo syndrome bearing missense mutation in the MED12 gene. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103388>
- Veleva, D., Ay, M., Ovchinnikov, D. A., Prowse, A. B. J., Menezes, M. J., & Nafisinia, M. (2024a). Generation of fibroblast-derived induced pluripotent stem cell (iPSC) lines from two paediatric patients with phenylketonuria. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103405>

- Veleva, D., Ay, M., Ovchinnikov, D. A., Prowse, A. B. J., Menezes, M. J., & Nafisinia, M. (2024b). Generation of two lymphoblastoid-derived induced pluripotent stem cell (iPSC) lines from patients with phenylketonuria. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103407>
- Villegas, L. D., Chandrasekaran, A., Andersen, S. A. F., Nørremølle, A., Schmid, B., Pouladi, M. A., & Freude, K. (2024). Generation of three isogenic gene-edited Huntington's disease human embryonic stem cell lines with DOX-inducible NGN2 expression cassette in the AAVS1 safe locus. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103408>
- Wang, B., Ren, Q., Cui, X., shan, W., Guo, X., Wang, X., Wang, J., Li, Y., & An, G. (2024). Generation of KCNH2 heterozygous knockout induced pluripotent stem cell (iPSC) line (Long and Short QT Syndrome). *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103400>
- Wang, B., Yang, L., Gao, M., Zhang, H., Ji, A., Liu, G., & Liu, Y. (2024). Establishment of a human induced pluripotent stem cell line (SDQLCHi079-A) from a patient with Johanson-Blizzard syndrome carrying heterozygous mutation in UBR1 gene. *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103505>
- Wang, B., Yang, L., Gao, M., Zhang, H., Liu, Y., & Gai, Z. (2024). Establishment of a human induced pluripotent stem cell line (SDQLCHi059-A) from a patient with congenital disorder of glycosylation carrying heterozygous mutation in MPI gene. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103381>
- Wang, J., Bering, J., Alonzo, M., Ye, S., Texter, K., Garg, V., & Zhao, M. T. (2024). Generation of iPSC line NCHi015-A from a patient with truncus arteriosus carrying heterozygous variants in KMT2D and NOTCH1. *Stem Cell Research*, 78. <https://doi.org/10.1016/J.SCR.2024.103457>
- Wang, P., Sun, W., Gong, J., Han, X., Xu, C., Chen, Y., Yang, Y., Luan, H., Li, S., Li, R., Wen, B., Lv, S., Chen, R., Guo, J., & Wei, C. (2024). Generation of human induced pluripotent stem cell line (XWHNi004-A) from a male with APOE gene mutation. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103398>
- wang, R., & shen, Q. (2024). Generation of RAB4A homozygous knockout induced pluripotent stem cell (iPSC) line. *Stem Cell Research*, 81, 103556. <https://doi.org/10.1016/J.SCR.2024.103556>
- Wang, T., Li, Y., Yang, C., Yuan, H., Na, W., & Yu, S. (2024). Generation of iPSC line (FMCPGHi003-A) from human PBMCs of a patient with Familial hemiplegic migraine type 3. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103465>
- Wang, X., Gao, J., Liu, C., & Sun, J. (2024). Establishment of human embryonic stem cell lines carrying LQT1 mutations by CRISPR base editing. *Stem Cell Research*, 79. <https://doi.org/10.1016/J.SCR.2024.103496>
- Wang, X., Zhuang, H., Tan, Y., Lei, F., Li, C., Zhang, C., Yu, X., & Sang, H. (2024). Generation of a human induced pluripotent stem cell (iPSC, DVSi001-A) with a heterozygous mutation in KRAS (A209T). *Stem Cell Research*, 80. <https://doi.org/10.1016/J.SCR.2024.103528>
- Wang, Y., Liu, W., Yang, Y., Wang, Y., Tang, Y., Zhan, Z., Sun, X., Jiao, Y., Shan, D., Zhang, R., Wang, D., Sun, P., Sun, X., Yan, C., & Liu, F. (2024). Establishment of an induced pluripotent stem cell (iPSC) line (INNSUi004-A) from a patient with Congenital Nemaline Myopathy. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103435>
- Wang, Y., Wei, C., Liu, Y., Lu, X., Wang, W., Song, N., Zhang, W., Xu, J., & Han, F. (2024). Generation of an induced pluripotent stem cell (iPSC) line from a Parkinson's disease patient with a pathogenic LRP10/c.688C > T(p.Arg230Trp) mutation. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103359>
- Weng, S., Liu, X., Wang, D., Wang, J., Ren, Q., Wang, Q., & Shan, W. (2024). Generation of a human induced pluripotent stem cell (iPSC) line from a patient with a CDKL5 gene mutation. *Stem Cell Research*, 81, 103607. <https://doi.org/10.1016/J.SCR.2024.103607>
- Wongkidakarn, S., Yodtup, C., Jantapalaboon, D., Phairoh, P., Suparak, S., Dhepakson, P., Tubsuwan, A., & Bumroongthai, K. (2024). Generation of human induced pluripotent stem cell (DMSCi001-A) line from

hematopoietic stem cells of a healthy female donor. *Stem Cell Research*, 81, 103605.
<https://doi.org/10.1016/J.SCR.2024.103605>

Woo Lim, S., In Lee, K., Cui, S., Fang, X., Jin Shin, Y., Lee, H., Woo Yang, C., Young Lee, J., & Ha Chung, B. (2024). Generation of a human SLC12A3 knock-in human induced pluripotent stem cell line (CMCi014-A-82) using CRISPR-Cas9 system. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103522>

Xin, H., Lv, Y., Wei, X., Song, W., Li, Z., Liu, Y., & Gai, Z. (2024). Establishment of a non-integrated iPSC (SDQLChi068-A) line derived from a patient with autosomal dominant immunodeficiency-14A carrying a heterozygous mutation (c.3061G>A) in PIK3CD gene. *Stem Cell Research*, 77.
<https://doi.org/10.1016/J.SCR.2024.103385>

Xu, H., Zhang, H., Pop, N., Hall, J., Shazlee, I., Wagner-Tsukamoto, M., Chen, Z., Gu, Y., Zhao, C., & Ma, D. (2025). The isoflavone puerarin promotes generation of human iPSC-derived pre-oligodendrocytes and enhances endogenous remyelination in rodent models. *Journal of Neurochemistry*, 169(1).
<https://doi.org/10.1111/JNC.16245>

Xu, Y., Wu, H., Jiang, J., Ye, L., Hao, K., Han, K., Hu, S., Lei, W., & Guo, Z. (2025). Generation and characterization of the LINC01405 knockout human embryonic stem cell line. *Stem Cell Research*, 82.
<https://doi.org/10.1016/J.SCR.2024.103619>

Ye, L., Ni, B., Wu, H., Han, X., Li, H., Liu, J., Hu, S., & Lei, W. (2024). Establishment of a human induced pluripotent stem cell line from a patient with dilated cardiomyopathy. *Stem Cell Research*, 78.
<https://doi.org/10.1016/J.SCR.2024.103467>

Zeng, Y., Tan, X., Xu, S., Wang, K., Li, X., & Jiang, Y. (2024). Generation of an integration-free induced pluripotent stem cell (iPSC) line (SDHI001-A) from a 65-year old adult mitral valve prolapse (MVP) patient. *Stem Cell Research*, 78. <https://doi.org/10.1016/J.SCR.2024.103464>

Zhang, C., Li, J., Sai, Y., Su, H., Jiang, Y., Zhang, L., Jian, L., Zhang, H., Guo, G., Li, E., Li, X., & Sun, L. (2024). Establishment of heterozygous LMOD2 knockout human embryonic stem cell line (ZZUNEU022-A-1) using CRISPR/Cas9 system. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103586>

Zhang, S., Li, J., Pan, W., Ren, Q., Zhou, Y., Chen, M., Qu, J., & Li, S. (2024). Establishment of a CPAMD8-GFP reporter human embryonic stem cell line, IBBDe001-B, using CRISPR/Cas9 editing. *Stem Cell Research*, 81. <https://doi.org/10.1016/J.SCR.2024.103615>

Zhang, T., Zhang, F., Wang, N., Xu, T., Zhu, L., Chen, L., & Liu, H. (2024). Generation of SST-P2A-mCherry reporter human embryonic stem cell line using the CRISPR/Cas9 system (WAe001-A-2C). *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103397>

Zhang, X., Li, Z., Liu, Y., Xin, H., & Gai, Z. (2024). Establishment of a non-integrated iPSC (SDQLChi066-A) line derived from Segawa syndrome patients harboring heterozygous mutations in the TH gene (p.G247S and p.D491H). *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103392>

Zhao, Y., Pan, Z., Hong, Z., Sun, M., Hong, Y., Peng, X., Li, X., Wang, X., & Wang, K. (2024). Protocol for scarless genome editing of human pluripotent stem cell based on orthogonal selective reporters. *STAR Protocols*, 5(2). <https://doi.org/10.1016/J.XPRO.2024.103084>

Zhen, Y., He, Q., Sun, C., Ma, Y., Li, Q., & Wen, L. (2024). Generate an AZFa deleted human embryonic stem cell line. *Stem Cell Research*, 77. <https://doi.org/10.1016/J.SCR.2024.103436>

Artificial intelligence (AI), robotics and automation in fertility treatment

Scope: This topic is focused on the integration of artificial intelligence, robotics or automation at any stage during the fertility journey. This includes the use of robotics for automation in the laboratory (eg automated ICSI, gamete/embryo freezing, preparation of culture dishes) or in the clinical treatment of infertility (eg endometriosis, myomectomy, fibroids, polyps), and AI tools/algorithms for basic science, embryo and gamete selection, and for prediction and improvement of outcomes before and after treatment. Time-lapse imaging is excluded from the search as it is considered under the treatment add-on 'time-lapse imaging and incubation'. Whilst patient support apps are included, AI apps to improve general health and wellbeing, which in turn impact fertility outcomes, are additionally excluded. Literature on regulation, guidelines and ethical considerations is included.

- AlSaad, R., Abd-Alrazaq, A., Choucair, F., Ahmed, A., Aziz, S., & Sheikh, J. (2024). Harnessing Artificial Intelligence to Predict Ovarian Stimulation Outcomes in In Vitro Fertilization: Scoping Review. *Journal of Medical Internet Research*, 26(1). <https://doi.org/10.2196/53396>
- Aykaç, A., Kaya, C., Çelik, Ö., Aydın, M. E., & Sungur, M. (2024). The prediction of semen quality based on lifestyle behaviours by the machine learning based models. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01268-w>
- Bachelot, G., Ly, A., Rivet-Danon, D., Sermondade, N., Frydman, V., Lamazière, A., Hamid, R. H., Levy, R., & Dupont, C. (2024). Artificial intelligence: to a better predictive strategy for testicular sperm extraction outcome in azoospermia. *Annales de Biologie Clinique*, 82(2), 1–11. <https://doi.org/10.1684/abc.2024.1882>
- Braude, I., Haikin Herzberger, E., Semo, M., Soifer, K., Goren Gepstein, N., Wisner, A., & Miller, N. (2024). Machine learning for predicting elective fertility preservation outcomes. *Scientific Reports*, 14(1). <https://doi.org/10.1038/s41598-024-60671-w>
- Canat, G., Duval, A., Gidel-Dissler, N., & Boussommier-Calleja, A. (2024). A novel deep learning approach to identify embryo morphokinetics in multiple time lapse systems. *Scientific Reports*, 14(1), 29016. <https://doi.org/10.1038/s41598-024-80565-1>
- Canon, C., Leibner, L., Fanton, M., Chang, Z., Suraj, V., Lee, J. A., Loewke, K., & Hoffman, D. (2024). Optimizing oocyte yield utilizing a machine learning model for dose and trigger decisions, a multi-center, prospective study. *Scientific Reports*, 14(1). <https://doi.org/10.1038/s41598-024-69165-1>
- Canosa, S., Licheri, N., Bergandi, L., Gennarelli, G., Paschero, C., Beccuti, M., Cimadomo, D., Coticchio, G., Rienzi, L., Benedetto, C., Cordero, F., & Revelli, A. (2024). A novel machine-learning framework based on early embryo morphokinetics identifies a feature signature associated with blastocyst development. *Journal of Ovarian Research*, 17(1). <https://doi.org/10.1186/s13048-024-01376-6>
- Cao, P., Derhaag, J., Coonen, E., Brunner, H., Acharya, G., Salumets, A., & Esteki, M. Z. (2024). Generative artificial intelligence to produce high-fidelity blastocyst-stage embryo images. *Human Reproduction*, 39(6), 1197–1207. <https://doi.org/10.1093/humrep/deae064>
- Cao, S. S., Liu, X. M., Song, B. T., & Hu, Y. Y. (2024). Interpretable machine learning models for predicting clinical pregnancies associated with surgical sperm retrieval from testes of different etiologies: a retrospective study. *BMC Urology*, 24(1). <https://doi.org/10.1186/s12894-024-01537-1>
- Chavez-Badiola, A., Fariás, A. F. S., Mendizabal-Ruiz, G., Silvestri, G., Griffin, D. K., Valencia-Murillo, R., Drakeley, A. J., & Cohen, J. (2024). Use of artificial intelligence embryo selection based on static images to predict first-trimester pregnancy loss. *Reproductive BioMedicine Online*, 49(2). <https://doi.org/10.1016/j.rbmo.2024.103934>
- Conversa, L., Bori, L., Insua, F., Marqueño, S., Cobo, A., & Meseguer, M. (2024). Testing an artificial intelligence algorithm to predict fetal heartbeat of vitrified-warmed blastocysts from a single image: predictive ability in different settings. *Human Reproduction*. <https://doi.org/10.1093/humrep/deae178>
- Cook, D. A. (2024). Creating virtual patients using large language models: scalable, global, and low cost. *Medical Teacher*, 1–3. <https://doi.org/10.1080/0142159X.2024.2376879>
- Correa, N., Cerquides, J., Arcos, J. L. L., Vassena, R., & Popovic, M. (2024). Personalizing the first dose of FSH for IVF/ICSI patients through machine learning: a non-inferiority study protocol for a multi-center randomized controlled trial. *Trials*, 25(1). <https://doi.org/10.1186/s13063-024-07907-2>
- Coticchio, G., Bartolacci, A., Cimadomo, V., Trio, S., Innocenti, F., Borini, A., Vaiarelli, A., Rienzi, L., Ahlström, A., & Cimadomo, D. (2024). Time will tell: time-lapse technology and artificial intelligence to set time cut-offs indicating embryo incompetence. *Human Reproduction (Oxford, England)*, 39(12), 2663–2673. <https://doi.org/10.1093/humrep/deae239>
- Dehghan, S., Rabiei, R., Choobineh, H., Maghooli, K., Nazari, M., & Vahidi-Asl, M. (2024). Comparative study of machine learning approaches integrated with genetic algorithm for IVF success prediction. *PLoS One*, 19(10), e0310829. <https://doi.org/10.1371/journal.pone.0310829>
- Dissler, N., Nogueira, D., Keppi, B., Sanguinet, P., Ozanon, C., Geoffroy-Siraudin, C., Pollet-Villard, X., & Boussommier-Calleja, A. (2024). Artificial intelligence-powered assisted ranking of sibling embryos to

increase first cycle pregnancy rate. *Reproductive BioMedicine Online*, 49(1).
<https://doi.org/10.1016/j.rbmo.2024.103887>

Elofsson, A., Han, L., Bianchi, E., Wright, G. J., & Jovine, L. (2024). Deep learning insights into the architecture of the mammalian egg-sperm fusion synapse. *ELife*, 13. <https://doi.org/10.7554/ELIFE.93131>

Fjeldstad, J., Qi, W., Mercuri, N., Siddique, N., Meriano, J., Krivoi, A., & Nayot, D. (2024). An artificial intelligence tool predicts blastocyst development from static images of fresh mature oocytes. *Reproductive BioMedicine Online*, 48(6). <https://doi.org/10.1016/j.rbmo.2024.103842>

Fjeldstad, J., Qi, W., Siddique, N., Mercuri, N., Nayot, D., & Krivoi, A. (2024). Segmentation of mature human oocytes provides interpretable and improved blastocyst outcome predictions by a machine learning model. *Scientific Reports*, 14(1). <https://doi.org/10.1038/s41598-024-60901-1>

Gallos, P., Matragkas, N., Islam, S. U., Epiphaniou, G., Hansen, S., Harrison, S., van Dijk, B., Haas, M., Pappous, G., Brouwer, S., Torlontano, F., Abbasi, S. F., Pournik, O., Churm, J., Mantas, J., Parra-Calderón, C. L., Petkousis, D., Weber, P., Dzingina, B., ... Arvanitis, T. N. (2024). INSAFEDARE Project: Innovative Applications of Assessment and Assurance of Data and Synthetic Data for Regulatory Decision Support. *Studies in Health Technology and Informatics*, 316, 1193–1197.
<https://doi.org/10.3233/SHTI240624>

Gardella, J., Abrahamsson, D., & Zelikoff, J. (2024). IMPACT OF REAL-LIFE ENVIRONMENTAL EXPOSURES ON REPRODUCTION: A contemporary review of machine learning to predict adverse pregnancy outcomes from pharmaceuticals, including DDIs. *Reproduction (Cambridge, England)*, 168(6).
<https://doi.org/10.1530/REP-24-0183>

Gbagbo, F. Y., Ameyaw, E. K., & Yaya, S. (2024). Artificial intelligence and sexual reproductive health and rights: a technological leap towards achieving sustainable development goal target 3.7. *Reproductive Health*, 21(1). <https://doi.org/10.1186/S12978-024-01924-9>

Goss, D. M., Vasilescu, S. A., Vasilescu, P. A., Cooke, S., Kim, S. H., Sacks, G. P., Gardner, D. K., & Warkiani, M. E. (2024). Evaluation of an artificial intelligence-facilitated sperm detection tool in azoospermic samples for use in ICSI. *Reproductive BioMedicine Online*, 49(1).
<https://doi.org/10.1016/j.rbmo.2024.103910>

Goswami, N., Winston, N., Choi, W., Lai, N. Z. E., Arcanjo, R. B., Chen, X., Sobh, N., Nowak, R. A., Anastasio, M. A., & Popescu, G. (2024). EVATOM: an optical, label-free, machine learning assisted embryo health assessment tool. *Communications Biology*, 7(1). <https://doi.org/10.1038/s42003-024-05960-w>

Gurbuz, T., Gokmen, O., Devranoglu, B., Yurci, A., & Madenli, A. A. (2024). Artificial intelligence in reproductive endocrinology: an in-depth longitudinal analysis of ChatGPTv4's month-by-month interpretation and adherence to clinical guidelines for diminished ovarian reserve. *Endocrine*, 86(3), 1171–1177. <https://doi.org/10.1007/s12020-024-04031-8>

Hall, J. M. M., Nguyen, T. V., Dinsmore, A. W., Perugini, D., Perugini, M., Fukunaga, N., Asada, Y., Schiewe, M., Lim, A. Y. X., Lee, C., Patel, N., Bhadarka, H., Chiang, J., Bose, D. P., Mankee-Sookram, S., Minto-Bain, C., Bilen, E., & Diakiw, S. M. (2024). Use of federated learning to develop an artificial intelligence model predicting usable blastocyst formation from pre-ICSI oocyte images. *Reproductive Biomedicine Online*, 49(6), 104403. <https://doi.org/10.1016/j.rbmo.2024.104403>

Huang, Z., Pinggera, G. M., & Agarwal, A. (2024). Enhancing Male Fertility Through AI-Based Management of Varicoceles. *Current Urology Reports*, 26(1), 18. <https://doi.org/10.1007/s11934-024-01241-5>

Iannone, A., Carfi, A., Mastrogiovanni, F., Zaccaria, R., & Manna, C. (2024). On the role of artificial intelligence in analysing oocytes during in vitro fertilisation procedures. *Artificial Intelligence in Medicine*, 157, 102997. <https://doi.org/10.1016/j.artmed.2024.102997>

Illingworth, P. J., Venetis, C., Gardner, D. K., Nelson, S. M., Berntsen, J., Larman, M. G., Agresta, F., Ahitan, S., Ahlström, A., Cattrall, F., Cooke, S., Demmers, K., Gabrielsen, A., Hindkjær, J., Kelley, R. L., Knight, C., Lee, L., Lahoud, R., Mangat, M., ... Hardarson, T. (2024). Deep learning versus manual morphology-based embryo selection in IVF: a randomized, double-blind noninferiority trial. *Nature Medicine*, 30(11), 3114–3120. <https://doi.org/10.1038/s41591-024-03166-5>

- Jin, L., Si, K., Li, Z., He, H., Wu, L., Ma, B., Ren, X., & Huang, B. (2024). Multiple collapses of blastocysts after full blastocyst formation is an independent risk factor for aneuploidy — a study based on AI and manual validation. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01242-6>
- Kalyani, K., & Deshpande, P. S. (2024). A deep learning model for predicting blastocyst formation from cleavage-stage human embryos using time-lapse images. *Scientific Reports*, 14(1), 28019. <https://doi.org/10.1038/s41598-024-79175-8>
- Kim, H. M., Kang, H., Lee, C., Park, J. H., Chung, M. K., Kim, M., Kim, N. Y., & Lee, H. J. (2024). Evaluation of the Clinical Efficacy and Trust in AI-Assisted Embryo Ranking: Survey-Based Prospective Study. *Journal of Medical Internet Research*, 26. <https://doi.org/10.2196/52637>
- Kolokythas, A., & Dahan, M. H. (2024). Is Artificial Intelligence (AI) currently able to provide evidence-based scientific responses on methods that can improve the outcomes of embryo transfers? No. *JBRA Assisted Reproduction*. <https://doi.org/10.5935/1518-0557.20240050>
- Koplin, J. J., Johnston, M., Webb, A. N. S., Whittaker, A., & Mills, C. (2024). Ethics of artificial intelligence in embryo assessment: mapping the terrain. *Human Reproduction*. <https://doi.org/10.1093/HUMREP/DEAE264>
- Lattin, M. T., Djandji, A. S., Kronfeld, M. T., Samsel, T., Ling, R., Ciskanik, M., Sadowy, S., Forman, E. J., & Williams, Z. (2024). Development and validation of an automated robotic system for preparation of embryo culture dishes. *Fertility and Sterility*, 122(2), 297–303. <https://doi.org/10.1016/j.fertnstert.2024.04.016>
- Lee, C. I., Huang, C. C., Lee, T. H., Chen, H. H., Cheng, E. H., Lin, P. Y., Yu, T. N., Chen, C. I., Chen, C. H., & Lee, M. S. (2024). Associations between the artificial intelligence scoring system and live birth outcomes in preimplantation genetic testing for aneuploidy cycles. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01185-y>
- Letterie, G. (2024). Moonshot. Long shot. Or sure shot. What needs to happen to realize the full potential of AI in the fertility sector? *Human Reproduction*, 39(9), 1863–1868. <https://doi.org/10.1093/humrep/deae144>
- Li, X., Yao, Y., Zhao, D., Chang, X., Li, Y., Lin, H., Wei, H., Wang, H., Mi, Y., Huang, L., Lu, S., Yang, W., & Cai, L. (2024). Clinical outcomes of single blastocyst transfer with machine learning guided noninvasive chromosome screening grading system in infertile patients. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01231-9>
- Li, Y., Zeng, H., & Fu, J. (2024). Preovulatory progesterone levels are the top indicator for ovulation prediction based on machine learning model evaluation: a retrospective study. *Journal of Ovarian Research*, 17(1). <https://doi.org/10.1186/s13048-024-01495-0>
- Liao, Z., Yan, C., Wang, J., Zhang, N., Yang, H., Lin, C., Zhang, H., Wang, W., & Li, W. (2024). A clinical consensus-compliant deep learning approach to quantitatively evaluate human in vitro fertilization early embryonic development with optical microscope images. *Artificial Intelligence in Medicine*, 149. <https://doi.org/10.1016/j.artmed.2024.102773>
- Liu, M., Lee, C. I., Tzeng, C. R., Lai, H. H., Huang, Y., & Chang, T. A. (2024). WISE: whole-scenario embryo identification using self-supervised learning encoder in IVF. *Journal of Assisted Reproduction and Genetics*, 41(4), 967–978. <https://doi.org/10.1007/s10815-024-03080-2>
- Liu, Z., Wang, M., He, S., Wang, X., Liu, X., Xie, X., & Bao, H. (2024). Derivation and validation of the first web-based nomogram to predict the spontaneous pregnancy after reproductive surgery using machine learning models. *Frontiers in Endocrinology*, 15, 1378157. <https://doi.org/10.3389/fendo.2024.1378157>
- Lopez-Campos, G., Gabarron, E., Martin-Sanchez, F., Merolli, M., Petersen, C., & Denecke, K. (2024). Digital Interventions and Their Unexpected Outcomes - Time for Digitalovigilance? *Studies in Health Technology and Informatics*, 310, 479–483. <https://doi.org/10.3233/SHTI231011>
- Luong, T. M. T., & Le, N. Q. K. (2024). Artificial intelligence in time-lapse system: advances, applications, and future perspectives in reproductive medicine. *Journal of Assisted Reproduction and Genetics*, 41(2), 239–252. <https://doi.org/10.1007/s10815-023-02973-y>

- Luong, T.-M.-T., Ho, N.-T., Hwu, Y.-M., Lin, S.-Y., Ho, J. Y.-P., Wang, R.-S., Lee, Y.-X., Tan, S.-J., Lee, Y.-R., Huang, Y.-L., Hsu, Y.-C., Le, N.-Q.-K., & Tzeng, C.-R. (2024). Beyond black-box models: explainable AI for embryo ploidy prediction and patient-centric consultation. *Journal of Assisted Reproduction and Genetics*, 41(9), 2349–2358. <https://doi.org/10.1007/s10815-024-03178-7>
- Luz, A., Hourvitz, A., Moran, E., Itzhak, N., Reuveny, S., Hourvitz, R., Youngster, M., Baum, M., & Maman, E. (2024). Improved clinical pregnancy rates in natural frozen-thawed embryo transfer cycles with machine learning ovulation prediction: insights from a retrospective cohort study. *Scientific Reports*, 14(1), 29451. <https://doi.org/10.1038/s41598-024-80356-8>
- Ma, B. X., Zhao, G. N., Yi, Z. F., Yang, Y. Le, Jin, L., & Huang, B. (2024). Enhancing clinical utility: deep learning-based embryo scoring model for non-invasive aneuploidy prediction. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01230-w>
- Mehrjerd, A., Dehghani, T., Jajroudi, M., Eslami, S., Rezaei, H., & Ghaebi, N. K. (2024). Ensemble machine learning models for sperm quality evaluation concerning success rate of clinical pregnancy in assisted reproductive techniques. *Scientific Reports*, 14(1), 24283. <https://doi.org/10.1038/s41598-024-73326-7>
- Montjean, D., Godin Pagé, M. H., Pacios, C., Calvé, A., Hamiche, G., Benkhalifa, M., & Miron, P. (2024). Automated Single-Sperm Selection Software (SiD) during ICSI: A Prospective Sibling Oocyte Evaluation. *Medical Sciences (Basel, Switzerland)*, 12(2). <https://doi.org/10.3390/medsci12020019>
- Mu, Y., Zhou, X., Li, L., Liu, X., Wen, X., Zhang, L., Yan, B., Zhang, W., Dong, K., Hu, H., Liao, Y., Ye, Z., Deng, A., Wang, Y., Mao, Z., Yang, M., & Xiao, X. (2024). Automatic high-throughput and non-invasive selection of sperm at the biochemical level. *Med*, 5(6), 603-621.e7. <https://doi.org/10.1016/j.medj.2024.03.008>
- Offord, C. (2024). AI reveals how sperm sticks to egg during fertilization. *Science (New York, N.Y.)*, 386(6720), 363–364. <https://doi.org/10.1126/science.adu0710>
- Olawade, D. B., Teke, J., Adeleye, K. K., Weerasinghe, K., Maidoki, M., & Clement David-Olawade, A. (2024). Artificial intelligence in in-vitro fertilization (IVF): A new era of precision and personalization in fertility treatments. *Journal of Gynecology Obstetrics and Human Reproduction*, 54(3). <https://doi.org/10.1016/J.JOGOH.2024.102903>
- Ortiz, J. A., Lledó, B., Morales, R., Mánuez-Grau, A., Cascales, A., Rodríguez-Arnedo, A., Castillo, J. C., Bernabeu, A., & Bernabeu, R. (2024). Factors affecting biochemical pregnancy loss (BPL) in preimplantation genetic testing for aneuploidy (PGT-A) cycles: machine learning-assisted identification. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01271-1>
- Palaniappan, K., Lin, E. Y. T., & Vogel, S. (2024). Global Regulatory Frameworks for the Use of Artificial Intelligence (AI) in the Healthcare Services Sector. *Healthcare (Basel, Switzerland)*, 12(5). <https://doi.org/10.3390/healthcare12050562>
- Panner Selvam, M. K., Moharana, A. K., Baskaran, S., Finelli, R., Hudnall, M. C., & Sikka, S. C. (2024). Current Updates on Involvement of Artificial Intelligence and Machine Learning in Semen Analysis. *Medicina (Lithuania)*, 60(2). <https://doi.org/10.3390/medicina60020279>
- Pantanowitz, L., Hanna, M., Pantanowitz, J., Lennerz, J., Henricks, W. H., Shen, P., Quinn, B., Bennet, S., & Rashidi, H. H. (2024). Regulatory Aspects of Artificial Intelligence and Machine Learning. *Modern Pathology*, 37(12). <https://doi.org/10.1016/j.modpat.2024.100609>
- Papamentzelopoulou, M. S., Prifti, I. N., Mavrogianni, D., Tseva, T., Soyhan, N., Athanasiou, A., Athanasiou, A., Athanasiou, A., Vogiatzi, P., Konomos, G., Loutradis, D., & Sakellariou, M. (2024). Assessment of artificial intelligence model and manual morphokinetic annotation system as embryo grading methods for successful live birth prediction: a retrospective monocentric study. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01198-7>
- Pavlovic, Z. J., Jiang, V. S., & Hariton, E. (2024). Current applications of artificial intelligence in assisted reproductive technologies through the perspective of a patient's journey. *Current Opinion in Obstetrics and Gynecology*, 36(4), 211–217. <https://doi.org/10.1097/GCO.0000000000000951>

- Peng, J., Geng, X., Zhao, Y., Hou, Z., Tian, X., Liu, X., Xiao, Y., & Liu, Y. (2024). Machine learning algorithms in constructing prediction models for assisted reproductive technology (ART) related live birth outcomes. *Scientific Reports*, 14(1). <https://doi.org/10.1038/S41598-024-83781-X>
- Pérez-Padilla, N. A., Garcia-Sanchez, R., Avalos, O., Gálvez, J., Bian, M., Yu, L., Shu, Y., Feng, M., & Yelian, F. D. (2024). Optimizing trigger timing in minimal ovarian stimulation for In Vitro fertilization using machine learning models with random search hyperparameter tuning. *Computers in Biology and Medicine*, 179. <https://doi.org/10.1016/j.combiomed.2024.108856>
- Rajendran, S., Brendel, M., Barnes, J., Zhan, Q., Malmsten, J. E., Zisimopoulos, P., Sigaras, A., Ofori-Atta, K., Meseguer, M., Miller, K. A., Hoffman, D., Rosenwaks, Z., Elemento, O., Zaninovic, N., & Hajirasouliha, I. (2024). Automatic ploidy prediction and quality assessment of human blastocysts using time-lapse imaging. *Nature Communications*, 15(1). <https://doi.org/10.1038/s41467-024-51823-7>
- Sengupta, P., Dutta, S., Jegasothy, R., Slama, P., Cho, C. L., & Roychoudhury, S. (2024). 'Intracytoplasmic sperm injection (ICSI) paradox' and 'andrological ignorance': AI in the era of fourth industrial revolution to navigate the blind spots. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01193-y>
- Setegn, G. M., & Dejene, B. E. (2024). Explainable artificial intelligence models for predicting pregnancy termination among reproductive-aged women in six east African countries: machine learning approach. *BMC Pregnancy and Childbirth*, 24(1). <https://doi.org/10.1186/s12884-024-06773-9>
- Shankar, V., van Blitterswijk, C., Vrij, E., & Giselsbrecht, S. (2024). Automated, High-Throughput Phenotypic Screening and Analysis Platform to Study Pre- and Post-Implantation Morphogenesis in Stem Cell-Derived Embryo-Like Structures. *Advanced Science*, 11(4). <https://doi.org/10.1002/advs.202304987>
- Shkodzik, K. (2024). Innovative Approaches to Digital Health in Ovulation Detection: A Review of Current Methods and Emerging Technologies. *Seminars in Reproductive Medicine*, 42(2), 81–89. <https://doi.org/10.1055/s-0044-1793829>
- Sun, L., Li, J., Zeng, S., Luo, Q., Miao, H., Liang, Y., Cheng, L., Sun, Z., Tai, W. H., Han, Y., Yin, Y., Wu, K., & Zhang, K. (2024). Artificial intelligence system for outcome evaluations of human in vitro fertilization-derived embryos. *Chinese Medical Journal*, 137(16), 1939–1949. <https://doi.org/10.1097/CM9.0000000000003162>
- Svempe, L. (2024). Exploring Impediments Imposed by the Medical Device Regulation EU 2017/745 on Software as a Medical Device. *JMIR Medical Informatics*, 12, e58080. <https://doi.org/10.2196/58080>
- Takehima, T., Karibe, J., Kuroda, S., & Yumura, Y. (2024). Development of a deep-learning model for detecting positive tubules during sperm recovery for nonobstructive azoospermia. *Reproduction (Cambridge, England)*, 168(4). <https://doi.org/10.1530/REP-24-0181>
- Ten, J., Herrero, L., Linares, Á., Álvarez, E., Ortiz, J. A., Bernabeu, A., & Bernabéu, R. (2024). Enhancing predictive models for egg donation: time to blastocyst hatching and machine learning insights. *Reproductive Biology and Endocrinology: RB&E*, 22(1), 116. <https://doi.org/10.1186/s12958-024-01285-9>
- Wang, C., Johansson, A. L. V., Nyberg, C., Pareek, A., Almqvist, C., Hernandez-Diaz, S., & Oberg, A. S. (2024). Prediction of pregnancy-related complications in women undergoing assisted reproduction, using machine learning methods. *Fertility and Sterility*, 122(1), 95–105. <https://doi.org/10.1016/j.fertnstert.2024.02.024>
- Wang, S., Chen, L., Fang, J., & Sun, H. (2024). A compact, high-throughput semi-automated embryo vitrification system based on hydrogel. *Reproductive BioMedicine Online*, 48(5). <https://doi.org/10.1016/j.rbmo.2023.103769>
- Wang, X., Wei, Q., Huang, W., Yin, L., & Ma, T. (2024). Can time-lapse culture combined with artificial intelligence improve ongoing pregnancy rates in fresh transfer cycles of single cleavage stage embryos? *Frontiers in Endocrinology*, 15, 1449035. <https://doi.org/10.3389/fendo.2024.1449035>
- Wu, J., Li, T., Xu, L., Chen, L., Liang, X., Lin, A., Zhang, W., & Huang, R. (2024). Development of a machine learning-based prediction model for clinical pregnancy of intrauterine insemination in a large Chinese population. *Journal of Assisted Reproduction and Genetics*, 41(8), 2173–2183. <https://doi.org/10.1007/s10815-024-03153-2>

Xiao, Y. H., Hu, Y. L., Lv, X. Y., Huang, L. J., Geng, L. H., Liao, P., Ding, Y. Bin, & Niu, C. C. (2024). The construction of machine learning-based predictive models for high-quality embryo formation in poor ovarian response patients with progestin-primed ovarian stimulation. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01251-5>

Yang, H. Y., Leahy, B. D., Jang, W. D., Wei, D., Kalma, Y., Rahav, R., Carmon, A., Kopel, R., Azem, F., Venturas, M., Kelleher, C. P., Cam, L., Pfister, H., Needleman, D. J., & Ben-Yosef, D. (2024). BlastAssist: a deep learning pipeline to measure interpretable features of human embryos. *Human Reproduction*, 39(4), 698–708. <https://doi.org/10.1093/humrep/deae024>

Yang, S.-R., Chien, J.-T., & Lee, C.-Y. (2025). Advancements in Clinical Evaluation and Regulatory Frameworks for AI-Driven Software as a Medical Device (SaMD). *IEEE Open Journal of Engineering in Medicine and Biology*, 6, 147–151. <https://doi.org/10.1109/OJEMB.2024.3485534>

Yao, M. W. M., Jenkins, J., Nguyen, E. T., Swanson, T., & Menabrito, M. (2024). Patient-Centric In Vitro Fertilization Prognostic Counseling Using Machine Learning for the Pragmatist. *Seminars in Reproductive Medicine*. <https://doi.org/10.1055/s-0044-1791536>

Artificial wombs for early or whole gestation (ectogenesis)

Scope: This topic is focused on research relevant to the partial or complete gestation of an embryo or fetus ex utero, including the sustained in vitro growth. As culturing of human embryos for research purposes is limited to 14-days or the appearance of the primitive streak under the HFE Act, research typically utilises animal or stem-cell based models to sustain growth or embryos or fetus' ex utero past 14-days. Although late-gestation support (termed artificial placentas and/or artificial wombs) falls outside the scope of the HFE Act, research in this field is monitored within this topic.

Note: With the introduction of 'Reproductive organoids' as a horizon scanning topic and the monitoring of advances in embryo culture systems through topics of 'Scientific developments relevant to the 14-day rule', the Executive have proposed that the topic of 'Artificial wombs for early or whole gestation (ectogenesis)' is removed from the prioritised list of horizon scanning topics and considered as a 'watching brief' topic. For further information, see associated paper (HFEA (03/02/2025) 008).

Accoe, D., & Pennings, G. (2024). Navigating conflicts of reproductive rights: Unbundling parenthood and balancing competing interests. *Bioethics*, 38(5), 425–430. <https://doi.org/10.1111/BIOE.13282>

AlJahsh, M. A. I. (2024). Science and Islamic ethics: Navigating artificial womb technology through Quranic principles. *Heliyon*, 10(17). <https://doi.org/10.1016/J.HELIYON.2024.E36793>

Blauvelt, D., & Roy, S. (2024). What is the feasibility of a clinical-scale and anticoagulation-free artificial placenta device? *Expert Review of Medical Devices*, 21(11). <https://doi.org/10.1080/17434440.2024.2419963>

Cavolo, A., de Boer, A., De Proost, L., Verweij, E. J., & Gastmans, C. (2024). Navigating the Ethical Landscape of the Artificial Placenta: A Systematic Review. *Prenatal Diagnosis*. <https://doi.org/10.1002/PD.6711>

Cohen, J. L., De Bie, F., Viaene, A. N., O'Grady, N., Rentas, S., Coons, B., Moon, J. K., Monson, E. E., Myers, R. A., Kalish, J. M., & Flake, A. W. (2024). Extrauterine support of pre-term lambs achieves similar transcriptomic profiling to late pre-term lamb brains. *Scientific Reports*, 14(1). <https://doi.org/10.1038/S41598-024-79095-7>

Filippi, L., Innocenti, F., Pascarella, F., Scaramuzza, R. T., Morganti, R., Bagnoli, P., Cammalleri, M., Dal Monte, M., Calvani, M., & Pini, A. (2024). β 3-Adrenoceptor Agonism to Mimic the Biological Effects of Intrauterine Hypoxia: Taking Great Strides Toward a Pharmacological Artificial Placenta. *Medicinal Research Reviews*. <https://doi.org/10.1002/MED.22092>

Heyer, J., Schubert, F., Seitz, A. L., Steinle, Y., Arens, J., Orlikowsky, T., Steinseifer, U., Schmitz-Rode, T., Jansen, S. V., & Schoberer, M. (2024). A Volume-Adjustable Artificial Womb for Extremely Preterm Infants. *Transplant International: Official Journal of the European Society for Organ Transplantation*, 37. <https://doi.org/10.3389/TI.2024.12947>

- Ikedo, H., Watanabe, S., Sato, S., Fee, E. L., Carter, S. W. D., Kumagai, Y., Takahashi, T., Kawamura, S., Hanita, T., Illanes, S. E., Choolani, M. A., Saito, M., Kikuchi, A., Kemp, M. W., & Usuda, H. (2024). Upregulation of hepatic nuclear receptors in extremely preterm ovine fetuses undergoing artificial placenta therapy. *The Journal of Maternal-Fetal & Neonatal Medicine : The Official Journal of the European Association of Perinatal Medicine, the Federation of Asia and Oceania Perinatal Societies, the International Society of Perinatal Obstetricians*, 37(1). <https://doi.org/10.1080/14767058.2023.2301651>
- Jonas, R. A. (2024). Management of fetal hemoglobin and risk of oxygen toxicity in the pump driven venovenous artificial placenta. *Perfusion*. <https://doi.org/10.1177/02676591241268368>
- Kendal, E. (2024). Whose (germ) line is it anyway? Reproductive technologies and kinship. *Bioethics*, 38(7), 632–642. <https://doi.org/10.1111/BIOE.13254>
- Kennedy, S. (2024). Ectogenesis and the value of gestational ties. *Bioethics*, 38(7), 643–649. <https://doi.org/10.1111/BIOE.13260>
- Kühle, H., Cho, S. K. S., Charest-Pekeski, A. J., Chow, J. S. M., Lee, F. T., Aujla, T., Saini, B. S., Lim, J. M., Darby, J. R. T., Mroczek, D., Floh, A. A., McVey, M. J., Morrison, J. L., Seed, M., Sun, L., & Haller, C. (2024). Echocardiographic assessment of cardiovascular physiology of preterm miniature piglets supported with a pumped artificial placenta system. *Prenatal Diagnosis*, 44(6–7), 888–898. <https://doi.org/10.1002/PD.6612>
- Nelson, A., Romanis, E. C., Adkins, V., Weis, C., & Kuberska, K. (2024). Death and the artificial placenta. *Journal of Law and the Biosciences*, 11(2). <https://doi.org/10.1093/JLB/LSAE013>
- Okpaise, O. O., Fils, A. J., Tonni, G., & Ruano, R. (2024). Artificial Ex Utero Systems to Treat Severe Periviable Fetal Growth Restriction-A Possible Future Indication? *Journal of Clinical Medicine*, 13(22). <https://doi.org/10.3390/JCM13226789>
- Romanis, E. C., & Adkins, V. (2024). Artificial placentas, pregnancy loss and loss-sensitive care. *Journal of Medical Ethics*, 50(5), 299–307. <https://doi.org/10.1136/JME-2023-109412>
- Romanis, E. C., Segers, S., & De Jong, B. D. (2024). Value sensitive design and the artificial placenta. *Journal of Medical Ethics*. <https://doi.org/10.1136/JME-2024-110066>
- Saraei, N., Dabaghi, M., Fusch, G., Rochow, N., Fusch, C., & Selvaganapathy, P. R. (2024). Scaled-up Microfluidic Lung Assist Device for Artificial Placenta Application with High Gas Exchange Capacity. *ACS Biomaterials Science & Engineering*, 10(7), 4612–4625. <https://doi.org/10.1021>

Emerging technologies in gamete and embryo testing

Scope: This topic includes research advancements in embryo testing methods, including but not limited to: non-invasive embryo testing (such as testing for aneuploidy and other parameters), advances in whole genome and exome sequencing, morphological grading with biopsy, novel blastocyst scoring systems, analysis of follicular fluid, novel PGT methods (eg mitochondrial copy number, segmental aneuploidy, PGT-P, etc), and emerging and novel oocyte/semen testing and selection methods. Previously male infertility testing (genetic testing through blood and sperm testing for infertility diagnosis) has been considered within scope.

Adolfsson, E., Ingberg, J., Igersten, E., & Bohlin, T. (2024). Clinical validation and experiences of the microfluidics sperm selection device ZyMöt™ for standard IVF. *JBRA Assisted Reproduction*. <https://doi.org/10.5935/1518-0557.20240104>

Ardehjani, N. A., Agha-Hosseini, M., Nashtaei, M. S., Khodarahmian, M., Shabani, M., Jabarpour, M., Fereidouni, F., Rastegar, T., & Amidi, F. (2024). Resveratrol ameliorates mitochondrial biogenesis and reproductive outcomes in women with polycystic ovary syndrome undergoing assisted reproduction: a randomized, triple-blind, placebo-controlled clinical trial. *Journal of Ovarian Research*, 17(1). <https://doi.org/10.1186/s13048-024-01470-9>

Ardestani, G., Banti, M., García-Pascual, C. M., Navarro-Sánchez, L., Van Zyl, E., Castellón, J. A., Simón, C., Sakkas, D., & Rubio, C. (2024). Culture time to optimize embryo cell-free DNA analysis for frozen-thawed blastocysts undergoing noninvasive preimplantation genetic testing for aneuploidy. *Fertility and Sterility*, 122(3), 465–473. <https://doi.org/10.1016/j.fertnstert.2024.04.037>

- Ardestani, G., Martins, M., Ocali, O., Sanchez, T. H., Gulliford, C., Barrett, C. B., & Sakkas, D. (2024). Effect of time post warming to embryo transfer on human blastocyst metabolism and pregnancy outcome. *Journal of Assisted Reproduction and Genetics*. <https://doi.org/10.1007/s10815-024-03115-8>
- Ariad, D., Madjunkova, S., Madjunkov, M., Chen, S., Abramov, R., Librach, C., & McCoy, R. C. (2024). Aberrant landscapes of maternal meiotic crossovers contribute to aneuploidies in human embryos. *Genome Research*, 34(1), 70–84. <https://doi.org/10.1101/gr.278168.123>
- Banti, M., Van Zyl, E., & Kafetzis, D. (2024). Sperm Preparation with Microfluidic Sperm Sorting Chip May Improve Intracytoplasmic Sperm Injection Outcomes Compared to Density Gradient Centrifugation. *Reproductive Sciences*, 31(6), 1695–1704. <https://doi.org/10.1007/s43032-024-01483-1>
- Barlevy, D., Cenolli, I., Campbell, T., Furrer, R., Mukherjee, M., Kostick-Quenet, K., Carmi, S., Lencz, T., Lázaro-Muñoz, G., & Pereira, S. (2024). Patient interest in and clinician reservations on polygenic embryo screening: a qualitative study of stakeholder perspectives. *Journal of Assisted Reproduction and Genetics*, 41(5), 1221–1231. <https://doi.org/10.1007/s10815-024-03074-0>
- Bednarska-Czerwińska, A., Smoleń-Dzirba, J., Strychalska, A., Sierka, W., Wróblewska, U., Mermer, P., Masarczyk, B., Jodłowiec-Lubańska, N., Kokot, A., Simka-Lampa, K., Zmarzły, N., Morawiec, E., Orczyk, A., & Grabarek, B. O. (2024). Comparison of Non-Invasive and Minimally Invasive Preimplantation Genetic Testing for Aneuploidy Using Samples Derived from the Same Embryo Culture. *Journal of Clinical Medicine*, 14(1). <https://doi.org/10.3390/JCM14010033>
- Bouloorchi Tabalvandani, M., Saeidpour, Z., Habibi, Z., Javadizadeh, S., Firoozabadi, S. A., & Badieirostami, M. (2024). Microfluidics as an emerging paradigm for assisted reproductive technology: A sperm separation perspective. *Biomedical Microdevices*, 26(2), 23. <https://doi.org/10.1007/s10544-024-00705-2>
- Boylan, C. F., Sambo, K. M., Neal-Perry, G., & Brayboy, L. M. (2024). Ex ovo omnia—why don't we know more about egg quality via imaging? *Biology of Reproduction*, 110(6), 1201–1212. <https://doi.org/10.1093/biolre/iaoe080>
- Brannigan, R. E., Hermanson, L., Kaczmarek, J., Kim, S. K., Kirkby, E., & Tanrikut, C. (2024). Updates to Male Infertility: AUA/ASRM Guideline (2024). *Journal of Urology*. <https://doi.org/10.1097/JU.0000000000004180>
- Cabello-Pinedo, S., Abdulla, H., Mas, S., Fraire, A., Maroto, B., Seth-Smith, M., Escriba, M., Teruel, J., Crespo, J., Munné, S., & Horcajadas, J. A. (2024). Development of a Novel Non-invasive Metabolomics Assay to Predict Implantation Potential of Human Embryos. *Reproductive Sciences*, 31(9), 2706–2717. <https://doi.org/10.1007/s43032-024-01583-y>
- Cambiasso, M. Y., Romanato, M., Gotfried, L., Valzacchi, G. R., Calvo, L., Calvo, J. C., & Fontana, V. A. (2024). Sperm histone modifications may predict success in human assisted reproduction: a pilot study. *Journal of Assisted Reproduction and Genetics*. <https://doi.org/10.1007/s10815-024-03280-w>
- Campos, G., & Nel-Themaat, L. (2024). Blastocoel fluid as an alternative source of DNA for minimally invasive PGT and biomarker of embryo competence. *Reproductive Biomedicine Online*, 49(4), 104322. <https://doi.org/10.1016/j.rbmo.2024.104322>
- Capalbo, A., de Wert, G., Mertes, H., Klausner, L., Coonen, E., Spinella, F., Van de Velde, H., Viville, S., Sermon, K., Vermeulen, N., Lencz, T., & Carmi, S. (2024). Screening embryos for polygenic disease risk: a review of epidemiological, clinical, and ethical considerations. *Human Reproduction Update*, 30(5), 529–557. <https://doi.org/10.1093/humupd/dmae012>
- Caroppo, E., & Skinner, M. K. (2024). Could the sperm epigenome become a diagnostic tool for evaluation of the infertile man? *Human Reproduction*, 39(3), 478–485. <https://doi.org/10.1093/humrep/dead266>
- Chavli, E. A., Klaasen, S. J., Van Opstal, D., Laven, J. S. E., Kops, G. J. P. L., & Baart, E. B. (2024). Single-cell DNA sequencing reveals a high incidence of chromosomal abnormalities in human blastocysts. *Journal of Clinical Investigation*, 134(6). <https://doi.org/10.1172/JCI174483>
- Chen, Y., Xie, M., Wu, S., Deng, Z., Tang, Y., Guan, Y., Ye, Y., He, Q., & Li, L. (2024). Multi-omics approach to reveal follicular metabolic changes and their effects on oocyte competence in PCOS patients. *Frontiers in Endocrinology*, 15, 1426517. <https://doi.org/10.3389/fendo.2024.1426517>

- Chien, C.-W., Tang, Y.-A., Jeng, S.-L., Pan, H.-A., & Sun, H. S. (2024). Blastocyst telomere length predicts successful implantation after frozen-thawed embryo transfer. *Human Reproduction Open*, 2024(2), hoae012. <https://doi.org/10.1093/hropen/hoae012>
- Chin, A. H. B., Al-Balas, Q., Ahmad, M. F., Alsomali, N., & Ghaly, M. (2024). Islamic Perspectives on Polygenic Testing and Selection of IVF Embryos (PGT-P) for Optimal Intelligence and Other Non-Disease-Related Socially Desirable Traits. *Journal of Bioethical Inquiry*, 21(3), 441–448. <https://doi.org/10.1007/s11673-023-10293-0>
- Chow, J. F. C., Lam, K. K. W., Cheng, H. H. Y., Lai, S. F., Yeung, W. S. B., & Ng, E. H. Y. (2024a). Optimizing non-invasive preimplantation genetic testing: investigating culture conditions, sample collection, and IVF treatment for improved non-invasive PGT-A results. *Journal of Assisted Reproduction and Genetics*, 41(2), 465–472. <https://doi.org/10.1007/s10815-023-03015-3>
- Chow, J. F. C., Lam, K. K. W., Cheng, H. H. Y., Lai, S. F., Yeung, W. S. B., & Ng, E. H. Y. (2024b). Optimizing non-invasive preimplantation genetic testing: investigating culture conditions, sample collection, and IVF treatment for improved non-invasive PGT-A results. *Journal of Assisted Reproduction and Genetics*, 41(2), 465–472. <https://doi.org/10.1007/s10815-023-03015-3>
- Chuang, T.-H., Chou, H.-H., Kuan, C.-S., Liu, S.-C., Kao, C.-W., Wu, Y.-H., Lai, H.-H., Hsieh, C.-L., Liang, Y.-T., Chen, C.-Y., & Chen, S.-U. (2024). Dependency of mitochondrial quantity on blastocyst timeline obscures its actual effect to pregnancy outcomes. *Frontiers in Endocrinology*, 15, 1415865. <https://doi.org/10.3389/fendo.2024.1415865>
- Córdova-Oriz, I., Cuadrado-Torroglosa, I., Madero-Molina, M., Rodríguez-García, A., Balmori, C., Medrano, M., Polonio, A. M., Chico-Sordo, L., Pacheco, A., García-Velasco, J. A., & Varela, E. (2024). Telomeric RNAs, TERRA, as a Potential Biomarker for Spermatozoa Quality. *Reproductive Sciences (Thousand Oaks, Calif.)*, 31(11), 3475–3484. <https://doi.org/10.1007/s43032-024-01690-w>
- Dai, M., Hong, L., Yin, T., & Liu, S. (2024). Disturbed Follicular Microenvironment in Polycystic Ovary Syndrome: Relationship to Oocyte Quality and Infertility. *Endocrinology (United States)*, 165(4). <https://doi.org/10.1210/endocr/bqae023>
- Daoud, S., Abdelkefi, O., Sellami, A., Bensalem, A., Chakroun, N., & Rebai, T. (2024). Association between hypo-osmotic swelling test-induced tail swelling patterns and sperm quality. *Future Science OA*, 10(1). <https://doi.org/10.1080/20565623.2024.2410696>
- Deng, S., Xu, Y., Warden, A. R., Xu, L., Duan, X., He, J., Bao, K., Xiao, R., Azmat, M., Hong, L., Jiang, L., Shen, G., Zhang, Z., & Ding, X. (2024). Quantitative Proteomics and Metabolomics of Culture Medium from Single Human Embryo Reveal Embryo Quality-Related Multiomics Biomarkers. *Analytical Chemistry*, 96(29), 11832–11844. <https://doi.org/10.1021/acs.analchem.4c01494>
- Fjeldstad, J., Qi, W., Mercuri, N., Siddique, N., Meriano, J., Krivoi, A., & Nayot, D. (2024). An artificial intelligence tool predicts blastocyst development from static images of fresh mature oocytes. *Reproductive BioMedicine Online*, 48(6). <https://doi.org/10.1016/j.rbmo.2024.103842>
- Furrer, R. A., Barlevy, D., Pereira, S., Carmi, S., Lencz, T., & Lázaro-Muñoz, G. (2024). Public Attitudes, Interests, and Concerns Regarding Polygenic Embryo Screening. *JAMA Network Open*, E2410832. <https://doi.org/10.1001/jamanetworkopen.2024.10832>
- Ganeva, R., Parvanov, D., Vidolova, N., Handzhiyska, M., Ruseva, M., Vasileva, M., Nikolova, K., Ivanova, I., Shaban, M., Shabarkova, J., Hristova, R., Miladinova, M., & Stamenov, G. (2024). Sperm selection by zona adhesion improves assisted reproductive treatment outcomes. *Andrology*, 12(6), 1373–1380. <https://doi.org/10.1111/andr.13590>
- Gao, Y., Liu, L., Tian, S., Liu, C., Lv, M., Wu, H., Tang, D., Song, B., Shen, Q., Xu, Y., Zhou, P., Wei, Z., Zhang, F., Cao, Y., & He, X. (2024). Whole-exome sequencing identifies ADGB as a novel causative gene for male infertility in humans: from motility to fertilization. *Andrology*. <https://doi.org/10.1111/andr.13605>
- Garrido, N., & Gil Juliá, M. (2024). The Use of Non-Apoptotic Sperm Selected by Magnetic Activated Cell Sorting (MACS) to Enhance Reproductive Outcomes: What the Evidence Says. *Biology*, 13(1). <https://doi.org/10.3390/biology13010030>

- Gayete-Lafuente, S., Vilà Famada, A., Albayrak, N., Espinós Gómez, J. J., Checa Vizcaíno, M. Á., & Moreno-Sepulveda, J. (2024). Indirect markers of oocyte quality in patients with ovarian endometriosis undergoing IVF/ICSI: a systematic review and meta-analysis. *Reproductive BioMedicine Online*, 49(3). <https://doi.org/10.1016/j.rbmo.2024.104075>
- Ginod, P., & Dahan, M. H. (2024). Preimplantation Genetic Testing for Polygenetic Conditions: A Legal, Ethical, and Scientific Challenge. *Seminars in Reproductive Medicine*. <https://doi.org/10.1055/s-0044-1782618>
- Goswami, N., Winston, N., Choi, W., Lai, N. Z. E., Arcanjo, R. B., Chen, X., Sobh, N., Nowak, R. A., Anastasio, M. A., & Popescu, G. (2024). EVATOM: an optical, label-free, machine learning assisted embryo health assessment tool. *Communications Biology*, 7(1). <https://doi.org/10.1038/s42003-024-05960-w>
- Handayani, N., Aubry, D., Boediono, A., Bowolaksono, A., Sini, I., Haq, N. M. D., Sirait, B., Periastringrum, G., Mutia, K., & Wiweko, B. (2024a). Non-invasive pre-implantation genetic testing's reliability for aneuploidy using Cell-free DNA in embryo culture media. *Journal of Gynecology Obstetrics and Human Reproduction*, 53(8). <https://doi.org/10.1016/j.jogoh.2024.102808>
- Handayani, N., Aubry, D., Boediono, A., Bowolaksono, A., Sini, I., Haq, N. M. D., Sirait, B., Periastringrum, G., Mutia, K., & Wiweko, B. (2024b). Non-invasive pre-implantation genetic testing's reliability for aneuploidy using Cell-free DNA in embryo culture media. *Journal of Gynecology Obstetrics and Human Reproduction*, 53(8). <https://doi.org/10.1016/j.jogoh.2024.102808>
- Handzhiyska, M., Ganeva, R., Parvanov, D., Ruseva, M., Eftimov, P., Georgieva, V., Velikova, D., & Stamenov, G. (2024). Cumulus matrix selection leads to isolation of spermatozoa with better motility, morphology, and lower DNA fragmentation. *Reproduction & Fertility*. <https://doi.org/10.1530/RAF-23-0052>
- Huang, B., Li, Z., Ren, X., Bai, J., Yue, J., Dong, X., Yang, L., Ma, B., Wang, J., Zhou, W., Wang, X., Guo, Y., Si, K., Shi, Z., & Jin, L. (2024). The density of the inner cell mass is a new indicator of the quality of a human blastocyst: A valid supplement to the Gardner scoring system. *Human Reproduction*, 39(9), 1942–1951. <https://doi.org/10.1093/humrep/deae158>
- Huang, T. K., Huang, C. H., Chen, P. A., Chen, C. H., Lu, F., Yang, W. J., Huang, J. Y. J., & Li, B. R. (2024). Development of a thermotaxis and rheotaxis microfluidic device for motile spermatozoa sorting. *Biosensors and Bioelectronics*, 258. <https://doi.org/10.1016/j.bios.2024.116353>
- Izadi, M., Khalili, M. A., Rezvani, M. E., Anbari, F., Maleki, B., Woodward, B., & Aflatoonian, B. (2024). Sperm Selection Using Zona Pellucida–Binding Enhanced Embryo Morphokinetic and Clinical Outcomes in ICSI: A Sibling Oocytes Study. *American Journal of Men's Health*, 18(1). <https://doi.org/10.1177/15579883241228236>
- Jahangiri, A. R., Ziarati, N., Dadkhah, E., Bucak, M. N., Rahimizadeh, P., Shahverdi, A., Sadighi Gilani, M. A., & Topraggaleh, T. R. (2024). Microfluidics: The future of sperm selection in assisted reproduction. *Andrology*, 12(6), 1236–1252. <https://doi.org/10.1111/andr.13578>
- Juchnewitsch, A.-G., Pomm, K., Dutta, A., Tamp, E., Valkna, A., Lillepea, K., Mahyari, E., Tjagur, S., Belova, G., Kübarsepp, V., Castillo-Madeen, H., Riera-Escamilla, A., Pölluaas, L., Nagirnaja, L., Poolamets, O., Vihljajev, V., Sütt, M., Versbraegen, N., Papadimitriou, S., ... Laan, M. (2024). Undiagnosed RASopathies in infertile men. *Frontiers in Endocrinology*, 15, 1312357. <https://doi.org/10.3389/fendo.2024.1312357>
- Kang, X., Wen, M., Zheng, J., Peng, F., Zeng, N., Chen, Z., Wu, Y., & Sun, H. (2024). Influence of the number of washings for embryos on non-invasive preimplantation chromosome screening results. *Frontiers in Endocrinology*, 15, 1363851. <https://doi.org/10.3389/fendo.2024.1363851>
- Karoi, D. H., Azizi, H., & Skutella, T. (2024). Whole transcriptome analysis to identify non-coding RNA regulators and hub genes in sperm of non-obstructive azoospermia by microarray, single-cell RNA sequencing, weighted gene co-expression network analysis, and mRNA-miRNA-lncRNA interaction analysis. *BMC Genomics*, 25(1). <https://doi.org/10.1186/s12864-024-10506-9>
- Li, Q., Guo, J., Huang, G., Wu, N., Chen, S., Dai, J., Zhang, X., Zhang, G., Zhi, W., Yan, J., Zheng, R., Yan, F., Yan, Z., Wu, L., Wu, S., Ji, Z., Zeng, J., Lin, G., Li, B., & Xu, W. (2024). Novel PLCZ1 compound

heterozygous mutations indicate gene dosage effect involved in TFF after ICSI. *Reproduction*.
<https://doi.org/10.1530/rep-23-0466>

Li, S., Zhang, Z., Xie, L., Zhao, Y., Chen, H., Zhang, S., Cai, Y., Ren, B., Liu, W., Tang, S., & Sha, Y. (2024). Novel bi-allelic DNAM3 variants cause oligoasthenoteratozoospermia. *Frontiers in Endocrinology*, 15, 1462509. <https://doi.org/10.3389/fendo.2024.1462509>

Liang, Y., Li, M., Fei, J., & Chen, Z. (2024). Should non-invasive prenatal testing be recommended for patients who achieve pregnancy with PGT? *BMC Pregnancy and Childbirth*, 24(1).
<https://doi.org/10.1186/s12884-024-06284-7>

Ma, J., Xie, Q., Zhang, Y., Xiao, Q., Liu, X., Qiao, C., & Tian, Y. (2024). Advances in microfluidic technology for sperm screening and in vitro fertilization. *Analytical and Bioanalytical Chemistry*, 416(16), 3717–3735.
<https://doi.org/10.1007/s00216-023-05120-9>

Madjunkov, M., Sharma, P., Baratz, A., Glass, K., Abramov, R., Logan, N., Madjunkova, S., & Librach, C. (2024). Prenatal cell-free DNA screening for chromosomal aneuploidies after euploid embryo transfer shows high concordance with preimplantation genetic testing for aneuploidy results and low positive predictive values. *Fertility and Sterility*. <https://doi.org/10.1016/j.fertnstert.2024.07.029>

Mantravadi, K. C., & Rao, D. (2024). In cases with raised sperm DNA fragmentation, can sperm selection by magnetic-activated cell sorting or testicular sperm aspiration help improve reproductive outcomes? *Journal of Assisted Reproduction and Genetics*, 41(6), 1507–1515. <https://doi.org/10.1007/s10815-024-03128-3>

Mateizel, I., Racca, A., Aligianni, E., Distasi, E., Baert, Y., Segers, I., Jankovic, D., Schoemans, C., Wouters, K., Tournaye, H., & De Munck, N. (2024). Optimized sperm selection: a highly efficient device for the isolation of progressive motile sperm with low DNA fragmentation index. *Journal of Assisted Reproduction and Genetics*, 41(8), 2201–2209. <https://doi.org/10.1007/s10815-024-03168-9>

Morimoto, A., Rose, R. D., Smith, K. M., Dinh, D. T., Umehara, T., Winstanley, Y. E., Shibahara, H., Russell, D. L., & Robker, R. L. (2024). Granulosa cell metabolism at ovulation correlates with oocyte competence and is disrupted by obesity and aging. *Human Reproduction*, 39(9), 2053–2066.
<https://doi.org/10.1093/humrep/deae154>

Moustakli, E., Zikopoulos, A., Skentou, C., Bouba, I., Dafopoulos, K., & Georgiou, I. (2024). Evolution of Minimally Invasive and Non-Invasive Preimplantation Genetic Testing: An Overview. *Journal of Clinical Medicine*, 13(8), 2160. <https://doi.org/10.3390/jcm13082160>

Nakhuda, G., Rodriguez, S., Tormasi, S., & Welch, C. (2024). A pilot study to investigate the clinically predictive values of copy number variations detected by next-generation sequencing of cell-free deoxyribonucleic acid in spent culture media. *Fertility and Sterility*, 122(1), 42–51.
<https://doi.org/10.1016/j.fertnstert.2024.02.030>

Ortega-Jaén, D., Mora-Martinez, C., Capalbo, A., Mifsud, A., Boluda-Navarro, M., Mercader, A., Martín, Á., Pardiñas, M. L., Gil, J., & de los Santos, M. J. (2024). A pilot study of transcriptomic preimplantation genetic testing (PGT-T): towards a new step in embryo selection? *Human Reproduction (Oxford, England)*.
<https://doi.org/10.1093/HUMREP/DEAE265>

Ovčar, A., & Kovačič, B. (2024). Biogenesis of Extracellular Vesicles (EVs) and the Potential Use of Embryo-Derived EVs in Medically Assisted Reproduction. *International Journal of Molecular Sciences*, 26(1). <https://doi.org/10.3390/IJMS26010042>

Parvin, A., Erabi, G., Alemi, A., Rezanezhad, A., Maleksabet, A., Sadeghpour, S., Taheri-Anganeh, M., & Ghasemnejad-Berenji, H. (2024). Seminal plasma proteomics as putative biomarkers for male infertility diagnosis. *Clinica Chimica Acta*, 561. <https://doi.org/10.1016/j.cca.2024.119757>

Pavuluri, H., Bakhtyari, Z., Panner Selvam, M. K., & Hellstrom, W. J. G. (2024). Oxidative Stress-Associated Male Infertility: Current Diagnostic and Therapeutic Approaches. *Medicina (Lithuania)*, 60(6).
<https://doi.org/10.3390/medicina60061008>

Płaczkowska, S., Kokot, I., Gilowska, I., & Kratz, E. M. (2024). Screening of cytokine expression in human seminal plasma in associations with sperm disorders and markers of oxidative-antioxidant balance. *Cytokine*, 182. <https://doi.org/10.1016/j.cyto.2024.156701>

- Przewocki, J., Kosiński, D., Łukaszuk, A., Jakiel, G., Wocławek-Potocka, I., Ołdziej, S., & Łukaszuk, K. (2024). Follicular Fluid Proteomic Analysis to Identify Predictive Markers of Normal Embryonic Development. *International Journal of Molecular Sciences*, 25(15). <https://doi.org/10.3390/ijms25158431>
- Raad, G., Tanios, J., Serdarogullari, M., Bazzi, M., Mourad, Y., Azoury, J., Yarkiner, Z., Liperis, G., Fakih, F., & Fakih, C. (2024). Mature oocyte dysmorphisms may be associated with progesterone levels, mitochondrial DNA content, and vitality in luteal granulosa cells. *Journal of Assisted Reproduction and Genetics*, 41(3), 795–813. <https://doi.org/10.1007/s10815-024-03053-5>
- Rashki Ghaleno, L., Pennisi, C. P., Shahverdi, A., Dardmeh, F., Alipour, H., & Rezazadeh Valojerdi, M. (2024). Exploring the Role of Hyaluronic Acid in Reproductive Biology and Beyond: Applications in Assisted Reproduction and Tissue Engineering. *Advanced Biology*, 8(6). <https://doi.org/10.1002/adbi.202300621>
- Sakkas, D., Navarro-Sánchez, L., Ardestani, G., Barroso, G., Bisioli, C., Boynukalin, K., Cimadomo, D., Frantz, N., Kopcow, L., Andrade, G. M., Ozturk, B., Rienzi, L., Weiser, A., Valbuena, D., Simón, C., & Rubio, C. (2024a). The impact of implementing a non-invasive preimplantation genetic testing for aneuploidies (niPGT-A) embryo culture protocol on embryo viability and clinical outcomes. *Human Reproduction*, 39(9), 1952–1959. <https://doi.org/10.1093/humrep/deae156>
- Sakkas, D., Navarro-Sánchez, L., Ardestani, G., Barroso, G., Bisioli, C., Boynukalin, K., Cimadomo, D., Frantz, N., Kopcow, L., Andrade, G. M., Ozturk, B., Rienzi, L., Weiser, A., Valbuena, D., Simón, C., & Rubio, C. (2024b). The impact of implementing a non-invasive preimplantation genetic testing for aneuploidies (niPGT-A) embryo culture protocol on embryo viability and clinical outcomes. *Human Reproduction*, 39(9), 1952–1959. <https://doi.org/10.1093/humrep/deae156>
- Sheibak, N., Amjadi, F., Shamloo, A., Zarei, F., & Zandieh, Z. (2024). Microfluidic sperm sorting selects a subpopulation of high-quality sperm with a higher potential for fertilization. *Human Reproduction*, 39(5), 902–911. <https://doi.org/10.1093/humrep/deae045>
- Siermann, M., Valcke, O., Vermeesch, J. R., Raivio, T., Tšuiiko, O., & Borry, P. (2024). “Are we not going too far?”: Socio-ethical considerations of preimplantation genetic testing using polygenic risk scores according to healthcare professionals. *Social Science and Medicine*, 343. <https://doi.org/10.1016/j.socscimed.2024.116599>
- Siermann, M., Van Der Schoot, V., Bunnik, E. M., & Borry, P. (2024). Ready for polygenic risk scores? An analysis of regulation of preimplantation genetic testing in European countries. *Human Reproduction*, 39(5), 1117–1130. <https://doi.org/10.1093/humrep/deae049>
- Siermann, M., Vermeesch, J. R., Raivio, T., Tšuiiko, O., & Borry, P. (2024). Polygenic embryo screening: quo vadis? *Journal of Assisted Reproduction and Genetics*, 41(7), 1719–1726. <https://doi.org/10.1007/s10815-024-03169-8>
- Siermann, M., Vermeesch, J. R., Raivio, T., Vanhie, A., Peeraer, K., Tšuiiko, O., & Borry, P. (2024). Perspectives of preimplantation genetic testing patients in Belgium on the ethics of polygenic embryo screening. *Reproductive BioMedicine Online*, 49(3). <https://doi.org/10.1016/j.rbmo.2024.104294>
- Sun, Q., Xu, J., Yao, Y., Huang, X., Zhao, D., Lu, S., Yao, B., & Chen, L. (2024). Efficacy of non-invasive chromosome screening, preimplantation genetic testing for aneuploidy, and morphological grading in selecting embryos of patients with advanced maternal age: a three-armed prospective cohort study. *BMC Pregnancy and Childbirth*, 24(1). <https://doi.org/10.1186/s12884-024-06736-0>
- Toschi, M., Bori, L., Rocha, J. C., Hickman, C., Fábio Gouveia Nogueira, M., Satoshi Ferreira, A., Costa Maffei, M., Malmsten, J., Zhan, Q., Zaninovic, N., & Meseguer, M. (2024). A Combination of Artificial Intelligence with Genetic Algorithms on Static Time-Lapse Images Improves Consistency in Blastocyst Assessment, An Interpretable Tool to Automate Human Embryo Evaluation: A Retrospective Cohort Study. *International Journal of Fertility and Sterility*, 18(4), 378–383. <https://doi.org/10.22074/IJFS.2024.2008339.1510>
- Tvrđonova, K., Belaskova, S., Rumpikova, T., Rumpik, D., Fucikova, A. M., & Malir, F. (2024). Prediction of live birth - selection of embryos using morphokinetic parameters. *Biomedical Papers of the Medical Faculty of the University Palacky, Olomouc, Czechoslovakia*, 168(1), 74–80. <https://doi.org/10.5507/BP.2022.052>

- Vargas-Ordaz, E., Newman, H., Austin, C., Catt, S., Nosrati, R., Cadarso, V. J., Neild, A., & Horta, F. (2024). Novel application of metabolic imaging of early embryos using a light-sheet on-a-chip device: a proof-of-concept study. *Human Reproduction (Oxford, England)*. <https://doi.org/10.1093/humrep/deae249>
- Venturas, M., Racowsky, C., & Needleman, D. J. (2024). Metabolic imaging of human cumulus cells reveals associations with pregnancy and live birth. *Human Reproduction*, 39(6), 1176–1185. <https://doi.org/10.1093/humrep/deae087>
- Volovsky, M., Scott, R. T., & Seli, E. (2024). Non-invasive preimplantation genetic testing for aneuploidy: is the promise real? *Human Reproduction*, 39(9), 1899–1908. <https://doi.org/10.1093/humrep/deae151>
- Wang, X., Shen, G., Yang, Y., Jiang, C., Ruan, T., Yang, X., Zhuo, L., Zhang, Y., Ou, Y., Zhao, X., Long, S., Tang, X., Lin, T., & Shen, Y. (2024). DNAH3 deficiency causes flagellar inner dynein arm loss and male infertility in humans and mice. *ELife*, 13. <https://doi.org/10.7554/eLife.96755>
- Widen, E., Lello, L., Eccles, J., Marin, D., & Treff, N. R. (2024). Correspondence on “Clinical utility of polygenic risk scores for embryo selection: A points to consider statement of the American College of Medical Genetics and Genomics (ACMG)” by Grebe et al. *Genetics in Medicine*, 26(8). <https://doi.org/10.1016/j.gim.2024.101155>
- Yang, K., Sun, X., Zheng, Q., Pan, C., Wang, S., Lu, Q., Xu, C., & Lu, Y. (2024). Transcriptome analysis of human spermatozoa with different DNA fragmentation index using RNA sequencing. *Reproductive Biology*, 24(4), 100964. <https://doi.org/10.1016/j.repbio.2024.100964>
- Yang, S., Xu, B., Zhuang, Y., Zhang, Q., Li, J., & Fu, X. (2024). Current research status and clinical applications of noninvasive preimplantation genetic testing: A review. *Medicine*, 103(40), e39964. <https://doi.org/10.1097/MD.00000000000039964>
- Yang, T., Yu, L., Xu, J., Ying, L., Jia, Y., Zheng, Y., Zhou, B., & Li, F. (2024). Correlation between standard sperm parameters and sperm DNA fragmentation from 11,339 samples. *Systems Biology in Reproductive Medicine*, 70(1), 91–100. <https://doi.org/10.1080/19396368.2024.2333285>
- Yildirim, R. M., & Seli, E. (2024a). Mitochondria as determinants of reproductive senescence and competence: implications for diagnosis of embryo competence in assisted reproduction. *Human Reproduction*. <https://doi.org/10.1093/humrep/deae171>
- Yildirim, R. M., & Seli, E. (2024b). The role of mitochondrial dynamics in oocyte and early embryo development. *Seminars in Cell and Developmental Biology*, 159–160, 52–61. <https://doi.org/10.1016/j.semcdb.2024.01.007>
- Zhang, X., Chao, S., Ye, N., & Ouyang, D. (2024). Emerging trends in sperm selection: enhancing success rates in assisted reproduction. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01239-1>
- Zhong, W., Wang, Q., Peng, D., Zou, Y., Chen, Y., Xia, Y., Zhang, X., Shu, M., Song, C., Wang, Y., Fu, Y., Wang, S., Ma, Y., Bu, X., Liang, Y., Chen, Y., Bai, W., Chen, Y., Deng, C., ... Shang, W. (2024). Genetic risk stratification and risk factors of early menopause in women: a multi-center study utilizing polygenic risk scores. *Frontiers in Endocrinology*, 15, 1518288. <https://doi.org/10.3389/fendo.2024.1518288>
- Zou, Y., Sui, Y., Fu, J., Ge, N., Sun, X., & Sun, Y. (2024). The morphokinetic signature of human blastocysts with mosaicism and the clinical outcomes following transfer of embryos with low-level mosaicism. *Journal of Ovarian Research*, 17(1). <https://doi.org/10.1186/s13048-023-01324-w>

Germline/Heritable Genome Editing

Scope: This topic is inclusive of developments in techniques used to manipulate the nuclear, mitochondrial, or epigenome for the alteration of DNA expression. If research is being conducted with the intention to refine a technique or delivery method, the publication will be included in literature scan. Research using models to understand physiology is excluded, however an overview of the applications will be given in the next paper. Literature on human and mammalian models is within scope.

Acharya, S., Ansari, A. H., Kumar Das, P., Hirano, S., Aich, M., Rauthan, R., Mahato, S., Maddileti, S., Sarkar, S., Kumar, M., Phutela, R., Gulati, S., Rahman, A., Goel, A., Afzal, C., Paul, D., Agrawal, T., Pulimamidi, V. K., Jalali, S., ... Chakraborty, D. (2024). PAM-flexible Engineered FnCas9 variants for

robust and ultra-precise genome editing and diagnostics. *Nature Communications*, 15(1).
<https://doi.org/10.1038/S41467-024-49233-W>

Ai, X., & Tang, Z. (2025). Aptazyme-directed A-to-I RNA editing. *Methods in Enzymology*, 710.
<https://doi.org/10.1016/BS.MIE.2024.11.022>

Alex, K., & Winkler, E. C. (2024). Comparative ethical evaluation of epigenome editing and genome editing in medicine: first steps and future directions. *Journal of Medical Ethics*, 50(6), 398–406.
<https://doi.org/10.1136/JME-2022-108888>

Allemailem, K. S., Almatroudi, A., Alrumaihi, F., Alradhi, A. E., Theyab, A., Algahtani, M., Alhawas, M. O., Dobie, G., Moawad, A. A., Rahmani, A. H., & Khan, A. A. (2024). Current Updates of CRISPR/Cas System and Anti-CRISPR Proteins: Innovative Applications to Improve the Genome Editing Strategies. *International Journal of Nanomedicine*, 19, 10185–10212. <https://doi.org/10.2147/IJN.S479068>

An, M., Raguram, A., Du, S. W., Banskota, S., Davis, J. R., Newby, G. A., Chen, P. Z., Palczewski, K., & Liu, D. R. (2024). Engineered virus-like particles for transient delivery of prime editor ribonucleoprotein complexes in vivo. *Nature Biotechnology*, 42(10). <https://doi.org/10.1038/S41587-023-02078-Y>

Aquino-Jarquín, G. (2024). Regulatory and governance gaps for human genome editing in Mexico. *Trends in Biotechnology*, 42(6), 665–670. <https://doi.org/10.1016/J.TIBTECH.2023.11.013>

Arivarasan, V. K., Diwakar, D., Kamarudheen, N., & Loganathan, K. (2025). Current approaches in CRISPR-Cas systems for diabetes. *Progress in Molecular Biology and Translational Science*, 210.
<https://doi.org/10.1016/BS.PMBTS.2024.08.002>

Aslan, C., Zolbanin, N. M., Faraji, F., & Jafari, R. (2024). Exosomes for CRISPR-Cas9 Delivery: The Cutting Edge in Genome Editing. *Molecular Biotechnology*, 66(11). <https://doi.org/10.1007/S12033-023-00932-7>

Bao-Xia, M., Sen, Y., Ming, L., Yu-Ren, W., Li-Ye, C., Yi-Fan, H., Jian-Gang, W., Yang, G., & Kun, X. (2024). Comparison and optimization of different CRISPR/Cas9 donor-adapting systems for gene editing. *Yi Chuan = Hereditas*, 46(6), 466–477. <https://doi.org/10.16288/J.YCZZ.23-273>

Barriga, F. M., & Lowe, S. W. (2024). Engineering megabase-sized genomic deletions with MACHETE (Molecular Alteration of Chromosomes with Engineered Tandem Elements). *Nature Protocols*, 19(5), 1381–1399. <https://doi.org/10.1038/S41596-024-00953-9>

Bekaert, B., Boel, A., Rybouchkin, A., Cosemans, G., Declercq, S., Chuva de Sousa Lopes, S. M., Parrington, J., Stoop, D., Coucke, P., Menten, B., & Heindryckx, B. (2024). Various repair events following CRISPR/Cas9-based mutational correction of an infertility-related mutation in mouse embryos. *Journal of Assisted Reproduction and Genetics*, 41(6), 1605–1617. <https://doi.org/10.1007/S10815-024-03095-9>

Briski, O., La Motta, G. E., Ratner, L. D., Allegroni, F. A., Pillado, S., Álvarez, G., Gutierrez, B., Tarragona, L., Zaccagnini, A., Acerbo, M., Ciampi, C., Fernández-Martin, R., & Salamone, D. F. (2024). Comparison of ICSI, IVF, and in vivo derived embryos to produce CRISPR-Cas9 gene-edited pigs for xenotransplantation. *Theriogenology*, 220, 43–55. <https://doi.org/10.1016/J.THERIOGENOLOGY.2024.02.028>

Callaway, E. (2024). “ChatGPT for CRISPR” creates new gene-editing tools. *Nature*, 629(8011), 272.
<https://doi.org/10.1038/D41586-024-01243-W>

Capdeville, N., Schindele, P., & Puchta, H. (2024). Increasing deletion sizes and the efficiency of CRISPR/Cas9-mediated mutagenesis by SunTag-mediated TREX1 recruitment. *The Plant Journal : For Cell and Molecular Biology*, 118(1), 277–287. <https://doi.org/10.1111/TPJ.16586>

Capin, J., Harrison, A., Raele, R. A., Yadav, S. K. N., Baiwir, D., Mazzucchelli, G., Quinton, L., Satchwell, T. J., Toye, A. M., Schaffitzel, C., Berger, I., & Aulicino, F. (2024). An engineered baculoviral protein and DNA co-delivery system for CRISPR-based mammalian genome editing. *Nucleic Acids Research*, 52(6), 3450–3468. <https://doi.org/10.1093/NAR/GKAE142>

Castillo, S. R., Simone, B. W., Clark, K. J., Devaux, P., & Ekker, S. C. (2024). Unconstrained Precision Mitochondrial Genome Editing with α DdCBEs. *Human Gene Therapy*, 35(19–20).
<https://doi.org/10.1089/HUM.2024.073>

Chen, L., Hong, M., Luan, C., Gao, H., Ru, G., Guo, X., Zhang, D., Zhang, S., Li, C., Wu, J., Randolph, P. B., Sousa, A. A., Qu, C., Zhu, Y., Guan, Y., Wang, L., Liu, M., Feng, B., Song, G., ... Li, D. (2024). Adenine

- transversion editors enable precise, efficient A•T-to-C•G base editing in mammalian cells and embryos. *Nature Biotechnology*, 42(4), 638–650. <https://doi.org/10.1038/S41587-023-01821-9>
- Chen, X. D., Chen, Z., Wythes, G., Zhang, Y., Orr, B. C., Sun, G., Chao, Y.-K., Navarro Torres, A., Thao, K., Vallurupalli, M., Sun, J., Borji, M., Tkacik, E., Chen, H., Bernstein, B. E., & Chen, F. (2024). Helicase-assisted continuous editing for programmable mutagenesis of endogenous genomes. *Science (New York, N.Y.)*, 386(6718). <https://doi.org/10.1126/SCIENCE.ADN5876>
- Chin, A. H. B., Nguma, J. D. B., & Ahmad, M. F. (2024). Stringent criteria needed for germline genome editing of human IVF embryos. *Journal of Assisted Reproduction and Genetics*, 41(7), 1727–1731. <https://doi.org/10.1007/s10815-024-03125-6>
- Chin, A. H. B., & Sun, N. (2024). Do not overlook the possibility of genome-edited somatic cells ending up in the human germline. *Journal of Community Genetics*. <https://doi.org/10.1007/s12687-024-00741-8>
- Cho, S. I., Lim, K., Hong, S., Lee, J., Kim, A., Lim, C. J., Ryou, S., Lee, J. M., Mok, Y. G., Chung, E., Kim, S., Han, S., Cho, S. M., Kim, J., Kim, E. K., Nam, K. H., Oh, Y., Choi, M., An, T. H., ... Kim, J. S. (2024). Engineering TALE-linked deaminases to facilitate precision adenine base editing in mitochondrial DNA. *Cell*, 187(1), 95-109.e26. <https://doi.org/10.1016/J.CELL.2023.11.035>
- Cohen, S., Bergman, S., Lynn, N., & Tuller, T. (2024). A tool for CRISPR-Cas9 sgRNA evaluation based on computational models of gene expression. *Genome Medicine*, 16(1), 152. <https://doi.org/10.1186/S13073-024-01420-6>
- Daliri, K., Hescheler, J., & Pfannkuche, K. P. (2024). Prime Editing and DNA Repair System: Balancing Efficiency with Safety. *Cells*, 13(10). <https://doi.org/10.3390/CELLS13100858>
- Darle, A., Mahiet, T., Aubin, D., Doyen, M., El Kassar, L., Parfait, B., Lemaitre, G., Baldeschi, C., Allouche, J., & Holic, N. (2024). Generation of heterozygous and homozygous NF1 lines from human-induced pluripotent stem cells using CRISPR/Cas9 to investigate bone defects associated with neurofibromatosis type 1. *Frontiers in Cell and Developmental Biology*, 12. <https://doi.org/10.3389/FCELL.2024.1359561>
- Del Arco, J., Acosta, J., & Fernández-Lucas, J. (2024). Biotechnological applications of purine and pyrimidine deaminases. *Biotechnology Advances*, 77. <https://doi.org/10.1016/J.BIOTECHADV.2024.108473>
- Dubey, S., Chen, Z., Jiang, Y. J., Talis, A., Molotkov, A., Ali, A., Mintz, A., & Momen-Heravi, F. (2024). Small extracellular vesicles (sEVs)-based gene delivery platform for cell-specific CRISPR/Cas9 genome editing. *Theranostics*, 14(7), 2777–2793. <https://doi.org/10.7150/THNO.92133>
- Duddy, G., Courtis, K., Horwood, J., Olsen, J., Horsler, H., Hodgson, T., Varsani-Brown, S., Abdullah, A., Denti, L., Lane, H., Delaqua, F., Janzen, J., Strom, M., Rosewell, I., Crawley, K., & Davies, B. (2024). Donor template delivery by recombinant adeno-associated virus for the production of knock-in mice. *BMC Biology*, 22(1). <https://doi.org/10.1186/S12915-024-01834-Z>
- Eghbalsaiied, S., Lawler, C., Petersen, B., Hajjiyev, R. A., Bischoff, S. R., & Frankenberg, S. (2024). CRISPR/Cas9-mediated base editors and their prospects for mitochondrial genome engineering. *Gene Therapy*, 31(5–6), 209–223. <https://doi.org/10.1038/S41434-023-00434-W>
- Elrick, H., Peterson, K. A., Willis, B. J., Lanza, D. G., Acar, E. F., Ryder, E. J., Teboul, L., Kasperek, P., Birling, M. C., Adams, D. J., Bradley, A., Braun, R. E., Brown, S. D., Caulder, A., Codner, G. F., DeMayo, F. J., Dickinson, M. E., Doe, B., Duddy, G., ... Nutter, L. M. J. (2024). Impact of essential genes on the success of genome editing experiments generating 3313 new genetically engineered mouse lines. *Scientific Reports*, 14(1), 22626. <https://doi.org/10.1038/S41598-024-72418-8>
- Ely, Z. A., Mathey-Andrews, N., Naranjo, S., Gould, S. I., Mercer, K. L., Newby, G. A., Cabana, C. M., Rideout, W. M., Jaramillo, G. C., Khirallah, J. M., Holland, K., Randolph, P. B., Freed-Pastor, W. A., Davis, J. R., Kulstad, Z., Westcott, P. M. K., Lin, L., Anzalone, A. V., Horton, B. L., ... Jacks, T. (2024). A prime editor mouse to model a broad spectrum of somatic mutations in vivo. *Nature Biotechnology*, 42(3), 424–436. <https://doi.org/10.1038/S41587-023-01783-Y>
- Erhardt, V., Hartig, E., Lorenzo, K., Megathlin, H. R., Tarchini, B., & Hosur, V. (2024). Large-Scale Genome-Wide Optimization and Prediction of the Cre Recombinase System for Precise Genome Manipulation in Mice. *BioRxiv : The Preprint Server for Biology*. <https://doi.org/10.1101/2024.06.14.599022>

- Fu, L., Wang, S., Liu, L., Shibata, Y., Okada, M., Luu, N., & Shi, Y. B. (2024). Simplifying Genotyping of Mutants from Genome Editing with a Parallel qPCR-Based iGenotype Index. *Cells*, 13(3). <https://doi.org/10.3390/cells13030247>
- Fu, Z.-C., Gao, B.-Q., Nan, F., Ma, X.-K., & Yang, L. (2024). DEMINING: A deep learning model embedded framework to distinguish RNA editing from DNA mutations in RNA sequencing data. *Genome Biology*, 25(1), 258. <https://doi.org/10.1186/S13059-024-03397-2>
- Gandadireja, A. P., Vos, P. D., Siira, S. J., Filipovska, A., & Rackham, O. (2024). Hyperactive Nickase Activity Improves Adenine Base Editing. *ACS Synthetic Biology*, 13(10). <https://doi.org/10.1021/ACSSYNBIO.4C00407>
- Gao, S., Guan, H., Bloomer, H., Wich, D., Song, D., Khirallah, J., Ye, Z., Zhao, Y., Chen, M., Xu, C., Liu, L., & Xu, Q. (2024). Harnessing non-Watson-Crick's base pairing to enhance CRISPR effectors cleavage activities and enable gene editing in mammalian cells. *Proceedings of the National Academy of Sciences of the United States of America*, 121(2). <https://doi.org/10.1073/PNAS.2308415120>
- Ge, W., Zhao, X., Gou, S., Jin, Q., Chen, F., Ouyang, Z., Lai, C., Cui, T., Mai, B., Lu, S., Zhong, K., Liang, Y., Chen, T., Wu, H., Li, N., Ye, Y., Lai, L., & Wang, K. (2023). Evaluation of guide-free Cas9-induced genomic damage and transcriptome changes in pig embryos. *Molecular Therapy. Nucleic Acids*, 34. <https://doi.org/10.1016/J.OMTN.2023.102035>
- Geuverink, W. P., Houtman, D., Retel Helmrich, I. R. A., van Baalen, S., van Beers, B. C., van El, C. G., Henneman, L., Kasprzak, M. D., Arets, D., & Riedijk, S. R. (2024). The need to set explicit goals for human germline gene editing public dialogues. *Journal of Community Genetics*, 15(3), 259–265. <https://doi.org/10.1007/S12687-024-00710-1>
- Guo, J., Gong, L., Yu, H., Li, M., An, Q., Liu, Z., Fan, S., Yang, C., Zhao, D., Han, J., & Xiang, H. (2024). Engineered minimal type I CRISPR-Cas system for transcriptional activation and base editing in human cells. *Nature Communications*, 15(1). <https://doi.org/10.1038/S41467-024-51695-X>
- Han, Y., Jia, Z., Xu, K., Li, Y., Lu, S., & Guan, L. (2024). CRISPR-Cpf1 system and its applications in animal genome editing. *Molecular Genetics and Genomics : MGG*, 299(1). <https://doi.org/10.1007/S00438-024-02166-X>
- Hausmann, I. U., Dix, T. C., McQuarrie, D. W. J., Dezi, V., Hans, A. I., Arnold, R., & Soller, M. (2024). Structure-optimized sgRNA selection with PlatinumCRISPr for efficient Cas9 generation of knockouts. *Genome Research*, 34(12), 2279–2292. <https://doi.org/10.1101/GR.279479.124>
- He, K., Xue, Q., Zhou, W., Wang, P., Hu, X., Lin, T., Chen, N., Wang, B., Ma, T., & Ding, S. (2025). Extended pegRNAs enhance the editing capability of Prime editing. *Trends in Biotechnology*, 43(1). <https://doi.org/10.1016/J.TIBTECH.2024.09.004>
- He, S., Huang, Z., Liu, Y., Ha, T., & Wu, B. (2024). DNA break induces rapid transcription repression mediated by proteasome-dependent RNAPII removal. *Cell Reports*, 43(7). <https://doi.org/10.1016/J.CELREP.2024.114420>
- Hermantara, R., Richmond, L., Taqi, A. F., Chilaka, S., Jeantet, V., Guerrini, I., West, K., & West, A. (2024). Improving CRISPR-Cas9 directed faithful transgene integration outcomes by reducing unwanted random DNA integration. *Journal of Biomedical Science*, 31(1). <https://doi.org/10.1186/S12929-024-01020-X>
- Hertel, O., & Neuss, A. (2024). Enhancing Cell Line Stability by CRISPR/Cas9-Mediated Site-Specific Integration Based on Histone Modifications. *Methods in Molecular Biology (Clifton, N.J.)*, 2810, 211–233. https://doi.org/10.1007/978-1-0716-3878-1_14
- Hołubowicz, R., Du, S. W., Felgner, J., Smidak, R., Choi, E. H., Palczewska, G., Menezes, C. R., Dong, Z., Gao, F., Medani, O., Yan, A. L., Hołubowicz, M. W., Chen, P. Z., Bassetto, M., Risaliti, E., Salom, D., Workman, J. N., Kiser, P. D., Foik, A. T., ... Palczewski, K. (2025). Safer and efficient base editing and prime editing via ribonucleoproteins delivered through optimized lipid-nanoparticle formulations. *Nature Biomedical Engineering*, 9(1). <https://doi.org/10.1038/S41551-024-01296-2>
- Hong, T., Bae, S. M., Song, G., & Lim, W. (2024). Guide for generating single-cell-derived knockout clones in mammalian cell lines using the CRISPR/Cas9 system. *Molecules and Cells*, 47(7). <https://doi.org/10.1016/J.MOCELL.2024.100087>

- Hosur, V., Erhardt, V., Hartig, E., Lorenzo, K., Megathlin, H., & Tarchini, B. (2024). Large-Scale Genome-Wide Optimization and Prediction of the Cre Recombinase System for Precise Genome Manipulation in Mice. *Research Square*. <https://doi.org/10.21203/RS.3.RS-4595968/V1>
- Hsiao, S., Chen, S., Jiang, Y., Wang, Q., Yang, Y., Lai, Y., Zhong, T., Liao, J., & Wu, Y. (2024). Library-Assisted Evolution in Eukaryotic Cells Yield Adenine Base Editors with Enhanced Editing Specificity. *Advanced Science (Weinheim, Baden-Wuerttemberg, Germany)*, 11(30). <https://doi.org/10.1002/ADVS.202309004>
- Huang, C. H., Chiu, S. Y., Chou, Y. C., & Wu, K. J. (2024). A refined Uni-vector prime editing system improves genome editing outcomes in mammalian cells. *Biotechnology Journal*, 19(2). <https://doi.org/10.1002/BIOT.202300353>
- Ikegawa, M., Kano, N., Ori, D., Fukuta, M., Hirano, M., Hewson, R., Yoshii, K., Kawai, T., & Kawasaki, T. (2024). HuR (ELAVL1) regulates the CCHFV minigenome and HAZV replication by associating with viral genomic RNA. *PLoS Neglected Tropical Diseases*, 18(9), e0012553. <https://doi.org/10.1371/JOURNAL.PNTD.0012553>
- Ilyas, M., Shah, Q., Gul, A., Ibrahim, H., Fatima, R., Babar, M. M., & Rajadas, J. (2024). Advances in CRISPR-Cas systems for epigenetics. *Progress in Molecular Biology and Translational Science*, 208, 185–209. <https://doi.org/10.1016/BS.PMBTS.2024.07.003>
- Inen, J., Han, C. M., Farrel, D. M., Bilousova, G., & Kogut, I. (2024). CIRCLE-Seq for Interrogation of Off-Target Gene Editing. *Journal of Visualized Experiments : JoVE*, 213. <https://doi.org/10.3791/67069>
- Jain, S., Xun, G., & Zhao, H. (2024). Impact of Chromatin Organization and Epigenetics on CRISPR-Cas and TALEN Genome Editing. *ACS Synthetic Biology*, 13(10). <https://doi.org/10.1021/ACSSYNBIO.4C00099>
- Kandlbinder, A. E. (2024). A critical view on using “life not worth living” in the bioethics of assisted reproduction. *Medicine, Health Care and Philosophy*, 27(2), 189–203. <https://doi.org/10.1007/s11019-023-10191-7>
- Kanke, K. L., Rayner, R. E., Bozik, J., Abel, E., Venugopalan, A., Suu, M., Nouri, R., Stack, J. T., Guo, G., Vetter, T. A., Cormet-Boyaka, E., Hester, M. E., & Vaidyanathan, S. (2024). Single-stranded DNA with internal base modifications mediates highly efficient knock-in in primary cells using CRISPR-Cas9. *Nucleic Acids Research*, 52(22), 13561–13576. <https://doi.org/10.1093/NAR/GKAE1069>
- Kantor, B., O'Donovan, B., Rittiner, J., Hodgson, D., Lindner, N., Guerrero, S., Dong, W., Zhang, A., & Chiba-Falek, O. (2024). The therapeutic implications of all-in-one AAV-delivered epigenome-editing platform in neurodegenerative disorders. *Nature Communications*, 15(1). <https://doi.org/10.1038/S41467-024-50515-6>
- Katayama, S., Watanabe, M., Kato, Y., Nomura, W., & Yamamoto, T. (2024). Engineering of Zinc Finger Nucleases Through Structural Modeling Improves Genome Editing Efficiency in Cells. *Advanced Science (Weinheim, Baden-Wuerttemberg, Germany)*, 11(23). <https://doi.org/10.1002/ADVS.202310255>
- Kato-Inui, T., Takahashi, G., Ono, T., & Miyaoka, Y. (2024). Fusion of histone variants to Cas9 suppresses non-homologous end joining. *PloS One*, 19(5). <https://doi.org/10.1371/JOURNAL.PONE.0288578>
- Kawai, H., Sato, K., Kato, T., & Kamiya, H. (2024). Correction of substitution, deletion, and insertion mutations by 5'-tailed duplexes. *Journal of Bioscience and Bioengineering*, 137(3), 157–164. <https://doi.org/10.1016/J.JBIOSEC.2023.12.011>
- Khan, S., & Drabiak, K. (2024). Eight Strategies to Engineer Acceptance of Human Germline Modifications. *Journal of Bioethical Inquiry*, 21(1), 81–94. <https://doi.org/10.1007/S11673-023-10266-3>
- Kim, J. S., & Chen, J. (2024). Base editing of organellar DNA with programmable deaminases. *Nature Reviews. Molecular Cell Biology*, 25(1), 34–45. <https://doi.org/10.1038/S41580-023-00663-2>
- Kim, Y.-J., Yun, D., Lee, J. K., Jung, C., & Chung, A. J. (2024). Highly efficient CRISPR-mediated genome editing through microfluidic droplet cell mechanoporation. *Nature Communications*, 15(1), 8099. <https://doi.org/10.1038/S41467-024-52493-1>

- Klermund, J., Rhiel, M., Kocher, T., Chmielewski, K. O., Bischof, J., Andrieux, G., el Gaz, M., Hainzl, S., Boerries, M., Cornu, T. I., Koller, U., & Cathomen, T. (2024). On- and off-target effects of paired CRISPR-Cas nickase in primary human cells. *Molecular Therapy : The Journal of the American Society of Gene Therapy*, 32(5), 1298–1310. <https://doi.org/10.1016/J.YMTHE.2024.03.006>
- Kose, A. M., Kocadagli, O., Taştan, C., Aktan, C., Ünalı, O. M., Güzenge, E., & Erdil, H. E. (2024). Unveiling Off-Target Mutations in CRISPR Guide RNAs: Implications for Gene Region Specificity. *The CRISPR Journal*, 7(3), 168–178. <https://doi.org/10.1089/CRISPR.2024.0002>
- Lam, S., Thomas, J. C., & Jackson, S. P. (2024). Genome-aware annotation of CRISPR guides validates targets in variant cell lines and enhances discovery in screens. *Genome Medicine*, 16(1), 139. <https://doi.org/10.1186/S13073-024-01414-4>
- Lampe, G. D., King, R. T., Halpin-Healy, T. S., Klompe, S. E., Hogan, M. I., Vo, P. L. H., Tang, S., Chavez, A., & Sternberg, S. H. (2024). Targeted DNA integration in human cells without double-strand breaks using CRISPR-associated transposases. *Nature Biotechnology*, 42(1), 87–98. <https://doi.org/10.1038/S41587-023-01748-1>
- Langley, J., Baudrier, L., & Billon, P. (2024). A protocol for the detection of precision genome editing in human cells using One-pot DTECT. *STAR Protocols*, 5(3), 103307. <https://doi.org/10.1016/J.XPRO.2024.103307>
- Lanza, D. G., Mao, J., Lorenzo, I., Liao, L., Seavitt, J. R., Ljungberg, M. C., Simpson, E. M., DeMayo, F. J., & Heaney, J. D. (2024). An oocyte-specific Cas9-expressing mouse for germline CRISPR/Cas9-mediated genome editing. *Genesis (New York, N.Y. : 2000)*, 62(2). <https://doi.org/10.1002/DVG.23589>
- Lau, C. H., Liang, Q. Le, & Zhu, H. (2024). Next-generation CRISPR technology for genome, epigenome and mitochondrial editing. *Transgenic Research*, 33(5). <https://doi.org/10.1007/S11248-024-00404-X>
- Lazar, N. H., Celik, S., Chen, L., Fay, M. M., Irish, J. C., Jensen, J., Tillinghast, C. A., Urbanik, J., Bone, W. P., Gibson, C. C., & Haque, I. S. (2024). High-resolution genome-wide mapping of chromosome-arm-scale truncations induced by CRISPR-Cas9 editing. *Nature Genetics*, 56(7), 1482–1493. <https://doi.org/10.1038/S41588-024-01758-Y>
- Leal, A. F., Herreno-Pachón, A. M., Benincore-Flórez, E., Karunathilaka, A., & Tomatsu, S. (2024). Current Strategies for Increasing Knock-In Efficiency in CRISPR/Cas9-Based Approaches. *International Journal of Molecular Sciences*, 25(5). <https://doi.org/10.3390/IJMS25052456>
- Leandro, K., Rufino-Ramos, D., Breyne, K., Di Ianni, E., Lopes, S. M., Jorge Nobre, R., Kleinstiver, B. P., Perdigão, P. R. L., Breakefield, X. O., & Pereira de Almeida, L. (2024). Exploring the potential of cell-derived vesicles for transient delivery of gene editing payloads. *Advanced Drug Delivery Reviews*, 211. <https://doi.org/10.1016/J.ADDR.2024.115346>
- Lenharo, M. (2024). Move over, CRISPR: RNA-editing therapies pick up steam. *Nature*, 626(8001), 933–934. <https://doi.org/10.1038/D41586-024-00275-6>
- Li, P., Dong, D., Gao, F., Xie, Y., Huang, H., Sun, S., Ma, Z., He, C., Lai, J., Du, X., & Wu, S. (2024). Versatile and efficient mammalian genome editing with Type I-C CRISPR System of *Desulfovibrio vulgaris*. *Science China. Life Sciences*, 67(11). <https://doi.org/10.1007/S11427-023-2682-5>
- Li, V. R., Wu, T., Tadych, A., Wong, A., & Zhang, Z. (2024). Widespread Impact of Natural Genetic Variations in CRISPR-Cas9 Outcomes. *The CRISPR Journal*, 7(5), 283–292. <https://doi.org/10.1089/CRISPR.2024.0020>
- Li, X., Chen, W., Martin, B. K., Calderon, D., Lee, C., Choi, J., Chardon, F. M., McDiarmid, T. A., Daza, R. M., Kim, H., Lalanne, J. B., Nathans, J. F., Lee, D. S., & Shendure, J. (2024). Chromatin context-dependent regulation and epigenetic manipulation of prime editing. *Cell*, 187(10), 2411–2427.e25. <https://doi.org/10.1016/J.CELL.2024.03.020>
- Liang, R., He, Z., Zhao, K. T., Zhu, H., Hu, J., Liu, G., Gao, Q., Liu, M., Zhang, R., Qiu, J. L., & Gao, C. (2024). Prime editing using CRISPR-Cas12a and circular RNAs in human cells. *Nature Biotechnology*, 42(12). <https://doi.org/10.1038/S41587-023-02095-X>

- Liang, S., Ma, N., Li, X., Yun, K., Meng, Q. F., Ma, K., Yue, L., Rao, L., Chen, X., & Wang, Z. (2024). A Guanidinobenzol-Rich Polymer Overcoming Cascade Delivery Barriers for CRISPR-Cas9 Genome Editing. *Nano Letters*, 24(23), 6872–6880. <https://doi.org/10.1021/ACS.NANOLETT.4C00533>
- Liao, H., Choi, J., & Shendure, J. (2024). Molecular recording using DNA Typewriter. *Nature Protocols*, 19(10). <https://doi.org/10.1038/S41596-024-01003-0>
- LIN, Q., TAKEBAYASHI, K., TORIGOE, N., LIU, B., NAMULA, Z., HIRATA, M., TANIHARA, F., NAGAHARA, M., & OTOI, T. (2024). Genome editing of porcine zygotes via lipofection of two guide RNAs using a CRISPR/Cas9 system. *The Journal of Reproduction and Development*, 70(6). <https://doi.org/10.1262/JRD.2024-054>
- Lin, Q., Torigoe, N., Liu, B., Nakayama, Y., Nakai, A., Namula, Z., Nagahara, M., Tanihara, F., Hirata, M., & Otoi, T. (2024). Efficient gene editing of pig embryos by combining electroporation and lipofection. *Veterinary World*, 17(11), 2701–2707. <https://doi.org/10.14202/VETWORLD.2024.2701-2707>
- Lin, Z., Yao, Q., Lai, K., Jiao, K., Zeng, X., Lei, G., Zhang, T., & Dai, H. (2024). Cas12f1 gene drives propagate efficiently in herpesviruses and induce minimal resistance. *Genome Biology*, 25(1). <https://doi.org/10.1186/S13059-024-03455-9>
- Liu, B., Dong, X., Zheng, C., Keener, D., Chen, Z., Cheng, H., Watts, J. K., Xue, W., & Sontheimer, E. J. (2024). Targeted genome editing with a DNA-dependent DNA polymerase and exogenous DNA-containing templates. *Nature Biotechnology*, 42(7), 1039–1045. <https://doi.org/10.1038/S41587-023-01947-W>
- Liu, Y., Kong, J., Liu, G., Li, Z., & Xiao, Y. (2024). Precise Gene Knock-In Tools with Minimized Risk of DSBs: A Trend for Gene Manipulation. *Advanced Science (Weinheim, Baden-Wuerttemberg, Germany)*, 11(28). <https://doi.org/10.1002/ADVS.202401797>
- Liu, Z., Liu, H., Huang, C., Zhou, Q., & Luo, Y. (2024). Hybrid Cas12a Variants with Relaxed PAM Requirements Expand Genome Editing Compatibility. *ACS Synthetic Biology*, 13(6), 1809–1819. <https://doi.org/10.1021/ACSSYNBIO.4C00103>
- Luo, X., Germer, J., Burghardt, T., Grau, M., Lin, Y., Höhn, M., Lächelt, U., & Wagner, E. (2025). Dual pH-responsive CRISPR/Cas9 ribonucleoprotein xenopeptide complexes for genome editing. *European Journal of Pharmaceutical Sciences : Official Journal of the European Federation for Pharmaceutical Sciences*, 205. <https://doi.org/10.1016/J.EJPS.2024.106983>
- Maladan, Y., Retnaningrum, E., Daryono, B. S., Sarassari, R., Sari, R. F., Balqis, S. A., Wahid, G. A., & Safari, D. (2024). A New Serotyping Method of *Streptococcus pneumoniae* Based on CRISPR/Cas9-Targeted Sequencing. *The Journal of Molecular Diagnostics : JMD*, 26(12), 1045–1054. <https://doi.org/10.1016/J.JMOLDX.2024.08.009>
- Martin, R., Espinoza, C. Y., Large, C. R. L., Rosswork, J., Bruinisse, C. Van, Miller, A. W., Sanchez, J. C., Miller, M., Paskvan, S., Alvino, G. M., Dunham, M. J., Raghuraman, M. K., & Brewer, B. J. (2024). Template switching between the leading and lagging strands at replication forks generates inverted copy number variants through hairpin-capped extrachromosomal DNA. *PLoS Genetics*, 20(1). <https://doi.org/10.1371/JOURNAL.PGEN.1010850>
- Matoušková, M., Plachý, J., Kučerová, D., Pecnová, L., Reinišová, M., Geryk, J., Karafiát, V., Hron, T., & Hejnar, J. (2024). Rapid adaptive evolution of avian leukosis virus subgroup J in response to biotechnologically induced host resistance. *PLoS Pathogens*, 20(8). <https://doi.org/10.1371/JOURNAL.PPAT.1012468>
- Matsuzaki, S., Sakuma, T., & Yamamoto, T. (2024). REMOVER-PITCh: microhomology-assisted long-range gene replacement with highly multiplexed CRISPR-Cas9. *In Vitro Cellular & Developmental Biology. Animal*, 60(7), 697–707. <https://doi.org/10.1007/S11626-024-00850-1>
- Meltzer, W. A., Gupta, A., Lin, P. N., Brown, R. A., Benyamien-Roufaeil, D. S., Khatri, R., Mahurkar, A. A., Song, Y., Taylor, R. J., & Zalzman, M. (2024). Reprogramming Chromosome Ends by Functional Histone Acetylation. *International Journal of Molecular Sciences*, 25(7). <https://doi.org/10.3390/IJMS25073898>
- Mikhailov, N., & Hämäläinen, R. H. (2024). Modulating Mitochondrial DNA Heteroplasmy with Mitochondrially Targeted Endonucleases. *Annals of Biomedical Engineering*, 52(9), 2627–2640. <https://doi.org/10.1007/S10439-022-03051-7>

- Misek, S. A., Fultineer, A., Kalfon, J., Noorbakhsh, J., Boyle, I., Roy, P., Dempster, J., Petronio, L., Huang, K., Saadat, A., Green, T., Brown, A., Doench, J. G., Root, D. E., McFarland, J. M., Beroukhim, R., & Boehm, J. S. (2024). Germline variation contributes to false negatives in CRISPR-based experiments with varying burden across ancestries. *Nature Communications*, 15(1). <https://doi.org/10.1038/S41467-024-48957-Z>
- Moraes, C. T. (2024). Tools for editing the mammalian mitochondrial genome. *Human Molecular Genetics*, 33(R1), R92–R99. <https://doi.org/10.1093/HMG/DDAE037>
- Morita, S., Horii, T., & Hatada, I. (2024). Optimized Protocol for the Regulation of DNA Methylation and Gene Expression Using Modified dCas9-SunTag Platforms. *Methods in Molecular Biology (Clifton, N.J.)*, 2842, 155–165. https://doi.org/10.1007/978-1-0716-4051-7_7
- Mu, S., Chen, H., Li, Q., Gou, S., Liu, X., Wang, J., Zheng, W., Chen, M., Jin, Q., Lai, L., Wang, K., & Shi, H. (2024). Enhancing prime editor flexibility with coiled-coil heterodimers. *Genome Biology*, 25(1). <https://doi.org/10.1186/S13059-024-03257-Z>
- Murjani, K., Tripathi, R., & Singh, V. (2024). An overview and potential of CRISPR-Cas systems for genome editing. *Progress in Molecular Biology and Translational Science*, 208, 1–17. <https://doi.org/10.1016/BS.PMBTS.2024.07.009>
- Naert, T., Yamamoto, T., Han, S., Horn, M., Bethge, P., Vladimirov, N., Voigt, F. F., Figueiro-Silva, J., Bachmann-Gagescu, R., Helmchen, F., & Lienkamp, S. S. (2024). Pythia: Non-random DNA repair allows predictable CRISPR/Cas9 integration and gene editing. *BioRxiv : The Preprint Server for Biology*. <https://doi.org/10.1101/2024.09.23.614424>
- Nakahara, T., Tabata, H., Kato, Y., Fuse, R., Nakamura, M., Yamaji, M., Hattori, N., Kiyono, T., Saito, I., & Nakanishi, T. (2024). Construction and Stability of All-in-One Adenovirus Vectors Simultaneously Expressing Four and Eight Multiplex Guide RNAs and Cas9 Nickase. *International Journal of Molecular Sciences*, 25(16). <https://doi.org/10.3390/IJMS25168783>
- Nakane, T., Nakagawa, R., Ishiguro, S., Okazaki, S., Mori, H., Shuto, Y., Yamashita, K., Yachie, N., Nishimasu, H., & Nureki, O. (2024). Structure and engineering of *Brevibacillus laterosporus* Cas9. *Communications Biology*, 7(1). <https://doi.org/10.1038/S42003-024-06422-Z>
- Ngo, W., Peukes, J., Baldwin, A., Xue, Z. W., Hwang, S., Stickels, R. R., Lin, Z., Satpathy, A. T., Wells, J. A., Schekman, R., Nogales, E., & Doudna, J. A. (2025). Mechanism-guided engineering of a minimal biological particle for genome editing. *Proceedings of the National Academy of Sciences of the United States of America*, 122(1). <https://doi.org/10.1073/PNAS.2413519121>
- Nikitchina, N., Ulashchik, E., Shmanai, V., Heckel, A. M., Tarassov, I., Mazunin, I., & Entelis, N. (2024). Targeting of CRISPR-Cas12a crRNAs into human mitochondria. *Biochimie*, 217, 74–85. <https://doi.org/10.1016/J.BIOCHI.2023.09.006>
- Núñez-Álvarez, Y., Espie-Caullet, T., Buhagiar, G., Rubio-Zulaika, A., Alonso-Marañón, J., Luna-Pérez, E., Blazquez, L., & Luco, R. F. (2024). A CRISPR-dCas13 RNA-editing tool to study alternative splicing. *Nucleic Acids Research*, 52(19). <https://doi.org/10.1093/NAR/GKAE682>
- Olszewska, M., Malcher, A., Stokowy, T., Pollock, N., Berman, A. J., Budkiewicz, S., Kamieniczna, M., Jackowiak, H., Suszynska-Zajczyk, J., Jedrzejczak, P., Yatsenko, A. N., & Kurpisch, M. (2024). Effects of Tcte1 knockout on energy chain transportation and spermatogenesis: implications for male infertility. *Human Reproduction Open*, 2024(2). <https://doi.org/10.1093/HROPEN/HOAE020>
- Palanki, R., Han, E. L., Murray, A. M., Maganti, R., Tang, S., Swingle, K. L., Kim, D., Yamagata, H., Safford, H. C., Mrksich, K., Peranteau, W. H., & Mitchell, M. J. (2024). Optimized microfluidic formulation and organic excipients for improved lipid nanoparticle mediated genome editing. *Lab on a Chip*, 24(16), 3790–3801. <https://doi.org/10.1039/D4LC00283K>
- Pandit, B., Fang, L., Kool, E. T., & Royzen, M. (2024). Reversible RNA Acylation Using Bio-Orthogonal Chemistry Enables Temporal Control of CRISPR-Cas9 Nuclease Activity. *ACS Chemical Biology*, 19(8), 1719–1724. <https://doi.org/10.1021/ACSCHEMBIO.4C00117>

- Park, J., Yu, G., Seo, S. Y., Yang, J., & Kim, H. H. (2024). SynDesign: web-based prime editing guide RNA design and evaluation tool for saturation genome editing. *Nucleic Acids Research*, 52(W1), W121–W125. <https://doi.org/10.1093/NAR/GKAE304>
- Parums, D. V. (2024). Editorial: Genome Editing Goes Beyond CRISPR with the Emergence of “Bridge” RNA Editing. *Medical Science Monitor : International Medical Journal of Experimental and Clinical Research*, 30. <https://doi.org/10.12659/MSM.945933>
- Pillai, A., Verma, V., & Galande, S. (2025). CHARM and EvoETR: Precision epigenetic tools for gene silencing. *BioEssays : News and Reviews in Molecular, Cellular and Developmental Biology*, 47(1). <https://doi.org/10.1002/BIES.202400186>
- Ponta, S., Bonato, A., Neidenbach, P., Bruhin, V. F., Laurent, A., Applegate, L. A., Zenobi-Wong, M., & Barreto, G. (2024). Streamlined, single-step non-viral CRISPR-Cas9 knockout strategy enhances gene editing efficiency in primary human chondrocyte populations. *Arthritis Research & Therapy*, 26(1). <https://doi.org/10.1186/S13075-024-03294-W>
- Porreca, I., Blassberg, R., Harbottle, J., Joubert, B., Mielczarek, O., Stombaugh, J., Hemphill, K., Sumner, J., Pazeraitis, D., Touza, J. L., Francescatto, M., Firth, M., Selmi, T., Collantes, J. C., Strezoska, Z., Taylor, B., Jin, S., Wiggins, C. M., van Brabant Smith, A., & Lambourne, J. J. (2024). An aptamer-mediated base editing platform for simultaneous knockin and multiple gene knockout for allogeneic CAR-T cells generation. *Molecular Therapy : The Journal of the American Society of Gene Therapy*, 32(8), 2692–2710. <https://doi.org/10.1016/J.YMTHE.2024.06.033>
- PR, A., X, C., S, S., A, S., AC, P., M, S., Ł, N., A, L., & G, P. (2024). Dimerization of the deaminase domain and locking interactions with Cas9 boost base editing efficiency in ABE8e. *Nucleic Acids Research*, 52(22), 13–14. <https://doi.org/10.1093/NAR/GKAE1066>
- Prokhorova, D. V., Kupryushkin, M. S., Zhukov, S. A., Zharkov, T. D., Dovydenko, I. S., Yakovleva, K. I., Pereverzev, I. M., Matveeva, A. M., Pyshnyi, D. V., & Stepanov, G. A. (2024). Effect of the Phosphoryl Guanidine Modification in Chimeric DNA-RNA crRNAs on the Activity of the CRISPR-Cas9 System In Vitro. *ACS Chemical Biology*, 19(6), 1311–1319. <https://doi.org/10.1021/ACSCHEMPIO.4C00147>
- Radford, E. J., Tan, H. K., Andersson, M. H. L., Stephenson, J. D., Gardner, E. J., Ironfield, H., Waters, A. J., Gitterman, D., Lindsay, S., Abascal, F., Martincorena, I., Kolesnik-Taylor, A., Ng-Cordell, E., Firth, H. V., Baker, K., Perry, J. R. B., Adams, D. J., Gerety, S. S., & Hurles, M. E. (2023). Saturation genome editing of DDX3X clarifies pathogenicity of germline and somatic variation. *Nature Communications*, 14(1). <https://doi.org/10.1038/S41467-023-43041-4>
- Rahman, S., Ikram, A. R., Azeem, F., Tahir ul Qamar, M., Shaheen, T., & Mehboob-ur-Rahman. (2024). Precision Genome Editing with CRISPR-Cas9. *Methods in Molecular Biology (Clifton, N.J.)*, 2788, 355–372. https://doi.org/10.1007/978-1-0716-3782-1_21
- Ramos, P. D., Almeida, M. S., & Olsson, I. A. S. (2023). What do people think about genetic engineering? A systematic review of questionnaire surveys before and after the introduction of CRISPR. *Frontiers in Genome Editing*, 5. <https://doi.org/10.3389/FGCEED.2023.1284547>
- Rimskaya, B., Shebanov, N., Entelis, N., & Mazunin, I. (2025). Enzymatic tools for mitochondrial genome manipulation. *Biochimie*, 229. <https://doi.org/10.1016/J.BIOCHI.2024.10.013>
- Rueda, J., Segers, S., Hopster, J., Kudlek, K., Liedo, B., Marchiori, S., & Danaher, J. (2024). Anticipatory gaps challenge the public governance of heritable human genome editing. *Journal of Medical Ethics*. <https://doi.org/10.1136/JME-2023-109801>
- Ryu, J., Barkal, S., Yu, T., Jankowiak, M., Zhou, Y., Francoeur, M., Phan, Q. V., Li, Z., Tognon, M., Brown, L., Love, M. I., Bhat, V., Lettre, G., Ascher, D. B., Cassa, C. A., Sherwood, R. I., & Pinello, L. (2024). Joint genotypic and phenotypic outcome modeling improves base editing variant effect quantification. *Nature Genetics*, 56(5), 925–937. <https://doi.org/10.1038/S41588-024-01726-6>
- Ryu, J., Statz, J. P., Chan, W., Oyama, K., Custer, M., Wienisch, M., Chen, R., Hanna, C. B., & Hennebold, J. D. (2024). Generation of Rhesus Macaque Embryos with Expanded CAG Trinucleotide Repeats in the Huntingtin Gene. *Cells*, 13(10). <https://doi.org/10.3390/cells13100829>

- Sakovina, L., Vokhtantsev, I., Akhmetova, E., Vorobyeva, M., Vorobjev, P., Zharkov, D. O., & Novopashina, D. (2024). Photocleavable Guide crRNAs for a Light-Controllable CRISPR/Cas9 System. *International Journal of Molecular Sciences*, 25(22), 12392. <https://doi.org/10.3390/IJMS252212392>
- Salomonsson, S. E., & Clelland, C. D. (2024). Building CRISPR Gene Therapies for the Central Nervous System: A Review. *JAMA Neurology*, 81(3), 283–290. <https://doi.org/10.1001/JAMANEUROL.2023.4983>
- Scheinerman, N. (2023). Public Engagement through Inclusive Deliberation: The Human Genome International Commission and Citizens' Juries. *The American Journal of Bioethics : AJOB*, 23(12), 66–76. <https://doi.org/10.1080/15265161.2022.2146786>
- Schep, R., Trauernicht, M., Vergara, X., Friskes, A., Morris, B., Gregoricchio, S., Manzo, S. G., Zwart, W., Beijersbergen, R. L., Medema, R. H., & Van Steensel, B. (2024). Chromatin context-dependent effects of epigenetic drugs on CRISPR-Cas9 editing. *Nucleic Acids Research*, 52(15), 8815–8832. <https://doi.org/10.1093/NAR/GKAE570>
- Seem, K., Kaur, S., Kumar, S., & Mohapatra, T. (2024). Epigenome editing for targeted DNA (de)methylation: a new perspective in modulating gene expression. *Critical Reviews in Biochemistry and Molecular Biology*, 59(1–2), 69–98. <https://doi.org/10.1080/10409238.2024.2320659>
- Sheikh, M. A., Afandi, F. H., Iannello, G., Corneo, B., Emerald, B. S., & Ansari, S. A. (2024). CRISPR-Cas9 Mediated Gene Deletion in Human Pluripotent Stem Cells Cultured Under Feeder-Free Conditions. *Journal of Visualized Experiments : JoVE*, 213. <https://doi.org/10.3791/67296>
- Shuto, Y., Nakagawa, R., Zhu, S., Hoki, M., Omura, S. N., Hirano, H., Itoh, Y., Zhang, F., & Nureki, O. (2024). Structural basis for pegRNA-guided reverse transcription by a prime editor. *Nature*, 631(8019), 224–231. <https://doi.org/10.1038/S41586-024-07497-8>
- Singh, K., Bhushan, B., Kumar, S., Singh, S., Macadangdang, R. R., Pandey, E., Varma, A. K., & Kumar, S. (2024). Precision Genome Editing Techniques in Gene Therapy: Current State and Future Prospects. *Current Gene Therapy*, 24(5), 377–394. <https://doi.org/10.2174/0115665232279528240115075352>
- Singh, S., Raj, D., Mathur, A., Mani, N., & Kumar, D. (2025). Current approaches in CRISPR-Cas systems for hereditary diseases. *Progress in Molecular Biology and Translational Science*, 210. <https://doi.org/10.1016/BS.PMBTS.2024.07.015>
- Singh, V., & Schimenti, J. C. (2024). Relevance, strategies, and added value of mouse models in androgenetics. *Andrology*. <https://doi.org/10.1111/ANDR.13761>
- Sledzinski, P., Nowaczyk, M., Smielowska, M. I., & Olejniczak, M. (2024). CRISPR/Cas9-induced double-strand breaks in the huntingtin locus lead to CAG repeat contraction through DNA end resection and homology-mediated repair. *BMC Biology*, 22(1), 282. <https://doi.org/10.1186/S12915-024-02079-6>
- Smith, K. R. (2024). Germline genome editing of human IVF embryos should not be subject to overly stringent restrictions. In *Journal of Assisted Reproduction and Genetics* (Vol. 41, Issue 7, pp. 1733–1737). Springer. <https://doi.org/10.1007/s10815-024-03174-x>
- Song, J., Zhuang, Y., & Yi, C. (2024). Programmable RNA base editing via targeted modifications. *Nature Chemical Biology*, 20(3), 277–290. <https://doi.org/10.1038/S41589-023-01531-Y>
- Stevens, C. S., Carmichael, J. C., Watkinson, R., Kowdle, S., Reis, R. A., Hamane, K., Jang, J., Park, A., Pernet, O., Khamaikawin, W., Hong, P., Thibault, P., Gowlikar, A., An, D. S., & Lee, B. (2024). A temperature-sensitive and less immunogenic Sendai virus for efficient gene editing. *Journal of Virology*, 98(12). <https://doi.org/10.1128/JVI.00832-24>
- Sun, Q., Zhang, H., Ding, F., Gao, X., Zhu, Z., & Yang, C. (2024). Development of ionizable lipid nanoparticles and a lyophilized formulation for potent CRISPR-Cas9 delivery and genome editing. *International Journal of Pharmaceutics*, 652. <https://doi.org/10.1016/J.IJPHARM.2024.123845>
- Taki, T., Morimoto, K., Mizuno, S., & Kuno, A. (2024). KOnezumi-AID: Automation Software for Efficient Multiplex Gene Knockout Using Target-AID. *International Journal of Molecular Sciences*, 25(24), 13500. <https://doi.org/10.3390/IJMS252413500>
- Terradas, G., Macias, V. M., Peterson, H., Mckeand, S., Krawczyk, G., & Rasgon, J. L. (2023). The Development and Expansion of in vivo Germline Editing Technologies in Arthropods: Receptor-Mediated

- Ovary Transduction of Cargo (ReMOT Control) and Beyond. *Integrative and Comparative Biology*, 63(6), 1550–1563. <https://doi.org/10.1093/ICB/ICAD123>
- Tompkins, J. D. (2023). Transgenerational Epigenetic DNA Methylation Editing and Human Disease. *Biomolecules*, 13(12). <https://doi.org/10.3390/BIOM13121684>
- Tong, M., Palmer, N., Dailamy, A., Kumar, A., Khaliq, H., Han, S., Finburgh, E., Wing, M., Hong, C., Xiang, Y., Miyasaki, K., Portell, A., Rainaldi, J., Suhardjo, A., Nourreddine, S., Chew, W. L., Kwon, E. J., & Mali, P. (2025). Robust genome and cell engineering via in vitro and in situ circularized RNAs. *Nature Biomedical Engineering*, 9(1). <https://doi.org/10.1038/S41551-024-01245-Z>
- Tran, N. T., & Han, R. (2024). Rapidly evolving genome and epigenome editing technologies. *Molecular Therapy : The Journal of the American Society of Gene Therapy*, 32(9). <https://doi.org/10.1016/J.YMTHE.2024.08.011>
- Tsuji-Hosokawa, A., Tsuchiya, I., Shimizu, K., Terao, M., Yasuhara, M., Miyamoto, N., Kikuchi, S., Ogawa, Y., Nakamura, K., Matsubara, Y., & Takada, S. (2024). Genetically humanized phenylketonuria mouse model as a testing tool for human genome editing in fertilized eggs. *Journal of Inherited Metabolic Disease*. <https://doi.org/10.1002/JIMD.12803>
- van den Berg van Saparoea, A. C. H., van Loosen, Q. C., Sarno, F., Ntini, E., Rots, M. G., Gjaltema, R. A. F., & Verschure, P. J. (2024). Plasmid Delivery and Single-Cell Plasmid Expression Analysis for CRISPR/dCas9-Based Epigenetic Editing. *Methods in Molecular Biology (Clifton, N.J.)*, 2842, 255–265. https://doi.org/10.1007/978-1-0716-4051-7_13
- Verma, A., Sharma, T., & Awasthi, A. (2024). CRISPR and Gene Editing: A Game-changer in Drug Development. *Current Pharmaceutical Design*, 30(15), 1133–1135. <https://doi.org/10.2174/0113816128298080240328053845>
- Villiger, L., Joung, J., Koblan, L., Weissman, J., Abudayyeh, O. O., & Gootenberg, J. S. (2024). CRISPR technologies for genome, epigenome and transcriptome editing. *Nature Reviews. Molecular Cell Biology*, 25(6), 464–487. <https://doi.org/10.1038/S41580-023-00697-6>
- Vora, D. S., Bhandari, S. M., & Sundar, D. (2024). DNA shape features improve prediction of CRISPR/Cas9 activity. *Methods (San Diego, Calif.)*, 226, 120–126. <https://doi.org/10.1016/J.YMETH.2024.04.012>
- Vos, P. D., Gandadireja, A. P., Rossetti, G., Siira, S. J., Mantegna, J. L., Filipovska, A., & Rackham, O. (2024). Mutational rescue of the activity of high-fidelity Cas9 enzymes. *Cell Reports Methods*, 4(4). <https://doi.org/10.1016/J.CRMETH.2024.100756>
- Walther, J., Porenta, D., Wilbie, D., Seinen, C., Benne, N., Yang, Q., de Jong, O. G., Lei, Z., & Mastrobattista, E. (2024). Comparative analysis of lipid Nanoparticle-Mediated delivery of CRISPR-Cas9 RNP versus mRNA/sgRNA for gene editing in vitro and in vivo. *European Journal of Pharmaceutics and Biopharmaceutics : Official Journal of Arbeitsgemeinschaft Fur Pharmazeutische Verfahrenstechnik e.V.*, 196. <https://doi.org/10.1016/J.EJPB.2024.114207>
- Wang, F., Ma, S., Zhang, S., Ji, Q., & Hu, C. (2024). CRISPR beyond: harnessing compact RNA-guided endonucleases for enhanced genome editing. *Science China. Life Sciences*, 67(12). <https://doi.org/10.1007/S11427-023-2566-8>
- Wang, J., Lu, X., Zhang, W., & Liu, G. H. (2024). Endogenous retroviruses in development and health. *Trends in Microbiology*, 32(4), 342–354. <https://doi.org/10.1016/J.TIM.2023.09.006>
- Wei, Y., Jin, M., Huang, S., Yao, F., Ren, N., Xu, K., Li, S., Gao, P., Zhou, Y., Chen, Y., Yang, H., Li, W., Xu, C., Zhang, M., & Wang, X. (2024). Enhanced C-To-T and A-To-G Base Editing in Mitochondrial DNA with Engineered DdCBE and TALE. *Advanced Science (Weinheim, Baden-Wuerttemberg, Germany)*, 11(3). <https://doi.org/10.1002/ADVS.202304113>
- Wu, L., Jiang, S., Shi, M., Yuan, T., Li, Y., Huang, P., Li, Y., Zuo, E., Zhou, C., & Sun, Y. (2024). Adenine base editors induce off-target structure variations in mouse embryos and primary human T cells. *Genome Biology*, 25(1), 291. <https://doi.org/10.1186/S13059-024-03434-0>

- Xu, W., Zhang, S., Qin, H., & Yao, K. (2024). From bench to bedside: cutting-edge applications of base editing and prime editing in precision medicine. *Journal of Translational Medicine*, 22(1). <https://doi.org/10.1186/S12967-024-05957-3>
- Xuan, Q., Wang, J., Nie, Y., Fang, C., & Liang, W. (2024). Research Progress and Application of Miniature CRISPR-Cas12 System in Gene Editing. *International Journal of Molecular Sciences*, 25(23). <https://doi.org/10.3390/IJMS252312686>
- Xue, N., Hong, D., Zhang, D., Wang, Q., Zhang, S., Yang, L., Chen, X., Li, Y., Han, H., Hu, C., Liu, M., Song, G., Guan, Y., Wang, L., Zhu, Y., & Li, D. (2024). Engineering IscB to develop highly efficient miniature editing tools in mammalian cells and embryos. *Molecular Cell*, 84(16), 3128-3140.e4. <https://doi.org/10.1016/J.MOLCEL.2024.07.007>
- Xun, G., Zhu, Z., Singh, N., Lu, J., Jain, P. K., & Zhao, H. (2024). Harnessing noncanonical crRNA for highly efficient genome editing. *Nature Communications*, 15(1). <https://doi.org/10.1038/S41467-024-48012-X>
- Yaish, O., & Orenstein, Y. (2024). Generating, modeling and evaluating a large-scale set of CRISPR/Cas9 off-target sites with bulges. *Nucleic Acids Research*, 52(12), 6777–6790. <https://doi.org/10.1093/NAR/GKAE428>
- Yan, H., Tan, X., Zou, S., Sun, Y., Ke, A., & Tang, W. (2024). Assessing and engineering the IscB- ω RNA system for programmed genome editing. *Nature Chemical Biology*, 20(12). <https://doi.org/10.1038/S41589-024-01669-3>
- Yan, J., Oyler-Castrillo, P., Ravisankar, P., Ward, C. C., Levesque, S., Jing, Y., Simpson, D., Zhao, A., Li, H., Yan, W., Goudy, L., Schmidt, R., Solley, S. C., Gilbert, L. A., Chan, M. M., Bauer, D. E., Marson, A., Parsons, L. R., & Adamson, B. (2024). Improving prime editing with an endogenous small RNA-binding protein. *Nature*, 628(8008), 639–647. <https://doi.org/10.1038/S41586-024-07259-6>
- Yan, K., Dumenil, T., Stewart, R., Bishop, C. R., Tang, B., Nguyen, W., Suhrbier, A., & Rawle, D. J. (2024). TMEM106B-mediated SARS-CoV-2 infection allows for robust ACE2-independent infection in vitro but not in vivo. *Cell Reports*, 43(11). <https://doi.org/10.1016/J.CELREP.2024.114921>
- Yang, L., Huo, Y., Wang, M., Zhang, D., Zhang, T., Wu, H., Rao, X., Meng, H., Yin, S., Mei, J., Zhang, D., Chen, X., Lv, J., Liu, M., Cheng, Y., Guan, Y., Feng, B., Song, G., Yi, C., ... Li, D. (2024). Engineering APOBEC3A deaminase for highly accurate and efficient base editing. *Nature Chemical Biology*, 20(9), 1176–1187. <https://doi.org/10.1038/S41589-024-01595-4>
- Yang, Q., Abebe, J. S., Mai, M., Rudy, G., Kim, S. Y., Devinsky, O., & Long, C. (2024). T4 DNA polymerase prevents deleterious on-target DNA damage and enhances precise CRISPR editing. *The EMBO Journal*, 43(17), 3733–3751. <https://doi.org/10.1038/S44318-024-00158-6>
- Yang, Z. X., Deng, D. H., Gao, Z. Y., Zhang, Z. K., Fu, Y. W., Wen, W., Zhang, F., Li, X., Li, H. Y., Zhang, J. P., & Zhang, X. B. (2024). OliTag-seq enhances in cellulo detection of CRISPR-Cas9 off-targets. *Communications Biology*, 7(1). <https://doi.org/10.1038/S42003-024-06360-W>
- Yi, Z., Zhang, X., Tang, W., Yu, Y., Wei, X., Zhang, X., & Wei, W. (2024). Strand-selective base editing of human mitochondrial DNA using mitoBEs. *Nature Biotechnology*, 42(3), 498–509. <https://doi.org/10.1038/S41587-023-01791-Y>
- Yi, Z., Zhang, X., Wei, X., Li, J., Ren, J., Zhang, X., Zhang, Y., Tang, H., Chang, X., Yu, Y., & Wei, W. (2024). Programmable DNA pyrimidine base editing via engineered uracil-DNA glycosylase. *Nature Communications*, 15(1). <https://doi.org/10.1038/S41467-024-50012-W>
- Yuan, B., Bi, C., Tian, Y., Wang, J., Jin, Y., Alsayegh, K., Tehseen, M., Yi, G., Zhou, X., Shao, Y., Romero, F. V., Fischle, W., Izpisua Belmonte, J. C., Hamdan, S., Huang, Y., & Li, M. (2024). Modulation of the microhomology-mediated end joining pathway suppresses large deletions and enhances homology-directed repair following CRISPR-Cas9-induced DNA breaks. *BMC Biology*, 22(1). <https://doi.org/10.1186/S12915-024-01896-Z>
- Yuan, Q., Zeng, H., Daniel, T. C., Liu, Q., Yang, Y., Osikpa, E. C., Yang, Q., Peddi, A., Abramson, L. M., Zhang, B., Xu, Y., & Gao, X. (2024). Orthogonal and multiplexable genetic perturbations with an

engineered prime editor and a diverse RNA array. *Nature Communications*, 15(1). <https://doi.org/10.1038/S41467-024-55134-9>

Zhang, G., Luo, Y., Xie, H., & Dai, Z. (2024). Crispr-SGRU: Prediction of CRISPR/Cas9 Off-Target Activities with Mismatches and Indels Using Stacked BiGRU. *International Journal of Molecular Sciences*, 25(20), 10945. <https://doi.org/10.3390/IJMS252010945>

Zhang, G., Song, Z., Huang, S., Wang, Y., Sun, J., Qiao, L., Li, G., Feng, Y., Han, W., Tang, J., Chen, Y., Huang, X., Liu, F., Wang, X., & Liu, J. (2024). nCas9 Engineering for Improved Target Interaction Presents an Effective Strategy to Enhance Base Editing. *Advanced Science (Weinheim, Baden-Wurttemberg, Germany)*, 11(31). <https://doi.org/10.1002/ADVS.202405426>

Zhang, G., Xie, H., & Dai, X. (2024). DeepIndel: An Interpretable Deep Learning Approach for Predicting CRISPR/Cas9-Mediated Editing Outcomes. *International Journal of Molecular Sciences*, 25(20). <https://doi.org/10.3390/IJMS252010928>

Zhong, Z., Hu, X., Zhang, R., Liu, X., Chen, W., Zhang, S., Sun, J., & Zhong, T. P. (2025). Improving precision base editing of the zebrafish genome by Rad51DBD-incorporated single-base editors. *Journal of Genetics and Genomics = Yi Chuan Xue Bao*, 52(1). <https://doi.org/10.1016/J.JGG.2024.10.006>

Zhou, B. Q., Li, S. P., Wang, X., Meng, X. Y., Deng, J. R., Xing, J. L., Wang, J. G., & Xu, K. (2024). Dual-localization signals enhance mitochondrial targeted presentation of engineered proteins. *Yi Chuan = Hereditas*, 46(11), 937–946. <https://doi.org/10.16288/J.YCZZ.24-171>

Zhu, W., Xie, H., Chen, Y., & Zhang, G. (2024). CrnnCrispr: An Interpretable Deep Learning Method for CRISPR/Cas9 sgRNA On-Target Activity Prediction. *International Journal of Molecular Sciences*, 25(8). <https://doi.org/10.3390/IJMS25084429>

Health outcomes in children born from ART (including the impact of culture media)

Scope: This topic includes research into the short and long-term health outcomes of children conceived through any form of ART (such as fresh or frozen embryo transfer, embryo testing, use of donor gametes, following male-factor infertility). In the short-term this includes trends in obstetric, maternal, perinatal, and neonatal outcomes, such as twin pregnancies, congenital malformations and birth defects. In the long term, this may include cognitive development, cardio-metabolic outcomes, risk of cancer, metabolic and reproductive functions of ART-concieved men, and other health outcomes (asthma, diabetes, imprinting disorders, cerebral palsy, etc). The effect of culture media on outcomes (including animal and human studies) is in scope.

Angel, P., Hermansen, M., Ramlau-Hansen, C. H., Gaml-Sørensen, A., Kristensen, D. M., & Lindahl-Jacobsen, R. (2024). Neurodevelopmental or behavioural disorders in children conceived after assisted reproductive technologies: A nationwide cohort study. *Fertility and Sterility*. <https://doi.org/10.1016/j.fertnstert.2024.10.017>

Appiah, D., Sang, J., Olayemi, O. E., Broni, E. K., Baykoca-Arslan, B., Ebong, I. A., & Kim, C. (2024). Infertility treatments and cyanotic congenital heart defects among livebirths in the USA: Findings from a contemporary cohort. *Human Reproduction*, 39(9), 2115–2123. <https://doi.org/10.1093/humrep/deae161>

Asserhøj, L. L., Mizrak, I., Lebech Kjaer, A. S., Clausen, T. D., Hoffmann, E. R., Greisen, G., Main, K. M., Madsen, P. L., Pinborg, A., & Jensen, R. B. (2024). Blood pressure and lipid profiles in children born after ART with frozen embryo transfer. *Human Reproduction Open*, 2024(2), hoae016. <https://doi.org/10.1093/hropen/hoae016>

Bartsch, L., Hämmerle, M., Putschögl, S., Hartmann, B., & Kirchengast, S. (2024). Assisted reproductive technology (ART) is not an independent risk factor for breech presentation among singleton term births in Vienna, Austria. *Journal of Biosocial Science*, 1–5. <https://doi.org/10.1017/S0021932024000130>

Bone, J. N., Joseph, K. S., Magee, L. A., Wang, L. Q., John, S., Bedaiwy, M. A., Mayer, C., & Lisonkova, S. (2024). Obesity, Twin Pregnancy, and the Role of Assisted Reproductive Technology. *JAMA Network Open*. <https://doi.org/10.1001/jamanetworkopen.2023.50934>

- Carneiro, F. A. T., Leong, V., Nóbrega, S., Salinas-Quiroz, F., Costa, P. A., & Leal, I. (2024). Are the children alright? A systematic review of psychological adjustment of children conceived by assisted reproductive technologies. *European Child and Adolescent Psychiatry*, 33(8), 2527–2546. <https://doi.org/10.1007/s00787-022-02129-w>
- Cavero-Ibircu, A., Canelas-Fernández, J., Gómez-Acebo, I., Alonso-Molero, J., Martínez-Jiménez, D., Llorca, J., Cabero-Perez, M. J., & Dierssen-Sotos, T. (2024). Association Between Assisted Reproductive Technology and Cerebral Palsy: A Meta-Analysis. *Pediatric Neurology*, 152, 115–124. <https://doi.org/10.1016/j.pediatrneurol.2023.12.019>
- Chen, L., Dong, Q., & Weng, R. (2024). Maternal and neonatal outcomes of dichorionic twin pregnancies achieved with assisted reproductive technology: meta-analysis of contemporary data. *Journal of Assisted Reproduction and Genetics*, 41(3), 581–589. <https://doi.org/10.1007/s10815-024-03035-7>
- Faa, G., Manchia, M., & Fanos, V. (2024). Assisted Reproductive Technologies: A New Player in the Foetal Programming of Childhood and Adult Diseases? *Pediatric Reports*, 16(2), 329–338. <https://doi.org/10.3390/pediatric16020029>
- Hwang, S., Jung, J., Moon, H., Ko, D. S., Kim, H.-W., Yoon, J.-P., Kim, W. K., Seol, A., Kim, K., & Kim, Y. H. (2024). The impact of assisted reproductive technologies on ADHD: A systematic review and meta-analysis. *Asian Journal of Psychiatry*, 99, 104125. <https://doi.org/10.1016/j.ajp.2024.104125>
- Ilmuratova, S., Lokshin, V., Prodeus, A., Manzhueva, L., Nurgaliyeva, Z., Kussainova, F., Bazarbaeva, A., Nekhorosheva, V., & Abshekenova, A. (2024). Immune profiling of ART-conceived children in Kazakhstan: a case-control study. *Frontiers in Pediatrics*, 12, 1447956. <https://doi.org/10.3389/fped.2024.1447956>
- Islam, M. I., Chaffey, O. A., Chadwick, V., & Martiniuk, A. (2024). Mental health in children conceived by Assisted Reproductive Technologies (ARTs): Insights from a longitudinal study of Australian children. *PLoS ONE*, 19(6 June). <https://doi.org/10.1371/journal.pone.0304213>
- Kristjansson, D., Lee, Y., Page, C. M., Gjessing, H., Magnus, M. C., Jugessur, A., Lyle, R., & Håberg, S. E. (2024). Sex differences in DNA methylation variations according to ART conception-evidence from the Norwegian mother, father, and child cohort study. *Scientific Reports*, 14(1), 22904. <https://doi.org/10.1038/s41598-024-73845-3>
- Kyhl, F., Spangmose, A. L., Gissler, M., Rönö, K., Westvik-Johari, K., Henningsen, A.-K. A., Bergh, C., Wennerholm, U.-B., Opdahl, S., Forman, J., Svensson, J., Clausen, T., Vassard, D., & Pinborg, A. (2024). The risk of Type 1 diabetes in children born after ART: a Nordic cohort study from the CoNARTaS group. *Human Reproduction Open*, 2024(2), hoae021. <https://doi.org/10.1093/hropen/hoae021>
- Lin, J., Zhang, K., Wu, F., Wang, B., Chai, W., Zhu, Q., Huang, J., & Lin, J. (2024). Maternal and perinatal risks for monozygotic twins conceived following frozen-thawed embryo transfer: a retrospective cohort study. *Journal of Ovarian Research*, 17(1). <https://doi.org/10.1186/s13048-024-01349-9>
- Lin, L., Yao, T., Liao, Q., Liu, J., Huang, L., & Zheng, L. (2024). Neonatal outcomes among twins born through assisted reproduction, compared to those born naturally. *Medicine*, 103(47), e40630. <https://doi.org/10.1097/MD.00000000000040630>
- Liu, S., Xu, Q., Qian, J., Liu, D., Zhang, B., Chen, X., & Zheng, M. (2024). Pregnancy outcomes of monochorionic diamniotic and dichorionic diamniotic twin pregnancies conceived by assisted reproductive technology and conceived naturally: a study based on chorionic comparison. *BMC Pregnancy and Childbirth*, 24(1). <https://doi.org/10.1186/s12884-024-06521-z>
- Liu, S., Zhou, X., Wang, W., Zhang, M., Sun, Y., Hu, X., You, J., Huang, X., Yang, Y., Feng, G., Xing, L., Bai, L., Tang, M., & Zhu, Y. (2024). The risk of asthma in singletons conceived by ART: a retrospective cohort study. *Human Reproduction Open*, 2024(3), hoae041. <https://doi.org/10.1093/hropen/hoae041>
- Marleen, S., Kodithuwakku, W., Nandasena, R., Mohideen, S., Allotey, J., Fernández-García, S., Gaetano-Gil, A., Ruiz-Calvo, G., Aquilina, J., Khalil, A., Bhide, P., Zamora, J., & Thangaratinam, S. (2024). Maternal and perinatal outcomes in twin pregnancies following assisted reproduction: a systematic review and meta-analysis involving 802 462 pregnancies. *Human Reproduction Update*, 30(3), 309–322. <https://doi.org/10.1093/humupd/dmae002>

- Matsumoto, N., Mitsui, T., Kadowaki, T., Mitsuhashi, T., Hirota, T., Masuyama, H., & Yorifuji, T. (2024). In vitro fertilization and long-term child health and development: nationwide birth cohort study in Japan. *European Journal of Pediatrics*, 184(1), 24. <https://doi.org/10.1007/s00431-024-05883-y>
- Mertens, J., Belva, F., van Montfoort, A. P. A., Regin, M., Zambelli, F., Seneca, S., Couvreur de Deckersberg, E., Bonduelle, M., Tournaye, H., Stouffs, K., Barbé, K., Smeets, H. J. M., Van de Velde, H., Sermon, K., Blockeel, C., & Spits, C. (2024). Children born after assisted reproduction more commonly carry a mitochondrial genotype associating with low birthweight. *Nature Communications*, 15(1). <https://doi.org/10.1038/s41467-024-45446-1>
- Nguyen, N. A., Nguyen, N. T., Tran, V. T. T., Vo, T. T. M., Uong, T. S., Nguyen, H. T., Nguyen, N. T., Nguyen, D. L., Pham, T. D., Nguyen, D. T. N., Ho, T. M., & Vuong, L. N. (2024). Developmental outcomes of children born through ICSI versus conventional IVF (cIVF) in couples with non-male factor infertility. *Human Reproduction*, 39(7), 1558–1563. <https://doi.org/10.1093/humrep/deae120>
- Oakley, L. L., Kristjansson, D., Munthe-Kaas, M. C., Nguyen, H. T., Lee, Y., Hanevik, H. I., Romundstad, L. B., Lyle, R., & Håberg, S. E. (2024). Sex differences in childhood cancer risk following ART conception: a registry-based study. *Human Reproduction*. <https://doi.org/10.1093/humrep/deae285>
- Ono, M., Kuji, N., Ueno, K., Kojima, J., & Nishi, H. (2024). The Long-Term Outcome of Children Conceived Through Assisted Reproductive Technology. *Reproductive Sciences (Thousand Oaks, Calif.)*, 31(3), 583–590. <https://doi.org/10.1007/S43032-023-01339-0>
- Piemonti, L., Vettor, L., Balducci, A., Farina, A., & Contro, E. (2024). Assisted reproductive technology and the risk of fetal congenital heart disease: insights from a tertiary-care referral center. *Archives of Gynecology and Obstetrics*, 310(4), 2073–2080. <https://doi.org/10.1007/s00404-024-07669-x>
- Pinborg, A., Blockeel, C., Coticchio, G., Garcia-Velasco, J., Santulli, P., & Campbell, A. (2024). Speaking up for the safety of the children following frozen embryo transfer. *Human Reproduction Open*, 2024(4). <https://doi.org/10.1093/HROPEN/HOAE058>
- Pinborg, A., Wennerholm, U. B., & Bergh, C. (2023). Long-term outcomes for children conceived by assisted reproductive technology. *Fertility and Sterility*, 120(3 Pt 1), 449–456. <https://doi.org/10.1016/J.FERTNSTERT.2023.04.022>
- Purkayastha, M., Sutcliffe, A., Brison, D. R., Nelson, S. M., Lawlor, D., & Roberts, S. A. (2024). Perinatal health in a cohort of children conceived after assisted reproduction in the UK: a population-based record-linkage study. *BMJ Open*, 14(11), e091910. <https://doi.org/10.1136/bmjopen-2024-091910>
- Raja, E. A., Bhattacharya, S., Maheshwari, A., & McLernon, D. J. (2023). A comparison of perinatal outcomes following fresh blastocyst or cleavage stage embryo transfer in singletons and twins and between singleton siblings. *Human Reproduction Open*, 2023(2). <https://doi.org/10.1093/HROPEN/HOAD003>
- Rios, P., Herlemont, P., Fauque, P., Lacour, B., Jouannet, P., Weill, A., Zureik, M., Clavel, J., & Dray-Spira, R. (2024). Medically Assisted Reproduction and Risk of Cancer among Offspring. *JAMA Network Open*, 7(5), E249429. <https://doi.org/10.1001/jamanetworkopen.2024.9429>
- Sargisian, N., Petzold, M., Furenäs, E., Gissler, M., Spangmose, A. L., Malchau Lauesgaard, S., Opdahl, S., Pinborg, A., Henningsen, A.-K. A., Westvik-Johari, K., Rönö, K., Bergh, C., & Wennerholm, U.-B. (2024). Congenital heart defects in children born after assisted reproductive technology: a CoNARTaS study. *European Heart Journal*. <https://doi.org/10.1093/eurheartj/ehae572>
- Shalev-Ram, H., Hershko Klement, A., Haikin-Herzberger, E., Levi, M., Rahav-Koren, R., Wisner, A., & Miller, N. (2024). Perinatal Outcomes in Siblings from Different Conception Methods: In Vitro Fertilization with Autologous Oocyte or Donor Egg vs. Unassisted Medical Conception. *Fertility and Sterility*. <https://doi.org/10.1016/j.fertnstert.2024.12.012>
- Sundrani, D., Kapare, A., Yadav, H., Randhir, K., Gupte, S., & Joshi, S. (2025). Placental expression and methylation of angiogenic factors in assisted reproductive technology pregnancies from India. *Epigenomics*, 17(1). <https://doi.org/10.1080/17501911.2024.2438593>
- Talbot, C., Hodson, N., Rose, J., & Bewley, S. (2024). Comparing the psychological outcomes of donor and non-donor conceived people: A systematic review. *BJOG : An International Journal of Obstetrics and Gynaecology*, 131(13), 1747–1759. <https://doi.org/10.1111/1471-0528.17892>

- Tang, W.-Z., Cai, Q.-Y., Wang, Y.-X., Shao, L.-Z., Zhang, X., Li, Z.-M., Tian, H., Liu, T.-H., Chen, Y., & Wang, L. (2024). Comparative influence of inappropriate gestational weight gain on pregnancy outcomes in IVF-conceived and spontaneously conceived twin pregnancies. *International Journal of Gynaecology and Obstetrics: The Official Organ of the International Federation of Gynaecology and Obstetrics*. <https://doi.org/10.1002/ijgo.15879>
- Tocariu, R., Niculae, L. E., Niculae, A. Ștefan, Carp-Velișcu, A., & Brătilă, E. (2024). Fresh versus Frozen Embryo Transfer in In Vitro Fertilization/Intracytoplasmic Sperm Injection Cycles: A Systematic Review and Meta-Analysis of Neonatal Outcomes. *Medicina (Lithuania)*, 60(8). <https://doi.org/10.3390/medicina60081373>
- Venetis, C., Choi, S. K. Y., Jorm, L., Zhang, X., Ledger, W., Lui, K., Havard, A., Chapman, M., Norman, R. J., & Chambers, G. M. (2023). Risk for Congenital Anomalies in Children Conceived With Medically Assisted Fertility Treatment: A Population-Based Cohort Study. *Annals of Internal Medicine*, 176(10), 1308–1320. <https://doi.org/10.7326/M23-0872>
- Waldaufova, E., Stastna, A., & Fait, T. (2024). The low birth weights of newborns conceived using assisted reproduction technology. *Bratislava Medical Journal*, 125(2), 137–143. https://doi.org/10.4149/BLL_2024_023
- Wang, W., Meng, Q., Hu, L., Du, J., Xu, B., Han, X., Liu, X., Zhou, K., Ke, K., Gan, M., Zhu, X., Peng, Y., Xue, H., Xiao, S., Lv, H., Jiang, Y., Jiang, T., Ma, H., Ling, X., ... Lin, Y. (2024). Assisted reproductive technology and neurodevelopment in children at 1 year of age: a longitudinal birth cohort study. *American Journal of Obstetrics and Gynecology*. <https://doi.org/10.1016/j.ajog.2024.05.039>
- Williams, Dr. C. L., Bunch, Mrs. K. J., Stiller, Mr. C., Murphy, Dr. M. F., Botting, Dr. B. J., Davies, Professor. M. C., Luke, Professor. B., Lupo, Professor. P. J., & Sutcliffe, Professor. A. G. (2024). Langerhans cell histiocytosis in children born after assisted reproductive technology. *Reproductive BioMedicine Online*, 49(6), 104379. <https://doi.org/10.1016/J.RBMO.2024.104379>
- Ye, M., Reyes Palomares, A., Iwarsson, E., Oberg, A. S., & Rodriguez-Wallberg, K. A. (2024). Imprinting disorders in children conceived with assisted reproductive technology in Sweden. *Fertility and Sterility*. <https://doi.org/10.1016/j.fertnstert.2024.05.168>
- Yeung, E. H., Trees, I. R., Clayton, P. K., Polinski, K. J., Livinski, A. A., & Putnick, D. L. (2024). Infertility treatment and offspring blood pressure—a systematic review and meta-analysis. *Human Reproduction Update*. <https://doi.org/10.1093/humupd/dmae029>
- Zeng, Z., Wang, Z., Yu, P., Wang, Y., Pei, Y., Dai, Y., Liu, Y., & Yang, Y. (2024). The Association between Assisted Reproductive Technologies and Neurodevelopmental Disorders in Offspring: An Overview of Current Evidence. *Journal of Integrative Neuroscience*, 23(1). <https://doi.org/10.31083/j.jin2301015>
- Zhang, B., Ban, M., Chen, X., Zhang, Y., Wang, Z., Feng, W., Zhao, H., Li, J., Zhang, T., Hu, J., Hu, K., Cui, L., & Chen, Z.-J. (2024). Associations between Paternal Obesity and Cardiometabolic Alterations in Offspring via Assisted Reproductive Technology. *The Journal of Clinical Endocrinology and Metabolism*. <https://doi.org/10.1210/clinem/dgae096>
- Zhang, G., Mao, Y., Zhang, Y., Huang, H., & Pan, J. (2023). Assisted reproductive technology and imprinting errors: analyzing underlying mechanisms from epigenetic regulation. *Human Fertility (Cambridge, England)*, 26(4), 864–878. <https://doi.org/10.1080/14647273.2023.2261628>
- Zhang, S., Luo, Q., Meng, R., Yan, J., Wu, Y., & Huang, H. (2024). Long-term health risk of offspring born from assisted reproductive technologies. *Journal of Assisted Reproduction and Genetics*, 41(3), 527–550. <https://doi.org/10.1007/S10815-023-02988-5>
- Zhang, Y., Dai, K., Chen, X., Cui, L., & Chen, Z. J. (2024). Association between being large for gestational age and cardiovascular metabolic health in children conceived from assisted reproductive technology: a prospective cohort study. *BMC Medicine*, 22(1). <https://doi.org/10.1186/s12916-024-03419-7>
- Zhao, J., Li, S., Ban, M., Gao, S., Cui, L., Yan, J., Yang, X., Li, J., Zhang, Y., Guan, S., Zhou, W., Gao, X., & Chen, Z.-J. (2024). Metabolic Profiles of Offspring Born From Biopsied Embryos from Toddlerhood to Preschool Age. *The Journal of Clinical Endocrinology and Metabolism*. <https://doi.org/10.1210/clinem/dgae315>

Zhou, W., Feng, W., Chang, J., Hu, J., Li, F., Hu, K., Jiao, J., Xue, X., Lan, T., Wan, W., Chen, Z. J., & Cui, L. (2024). Metabolic profiles of children aged 2-5 years born after frozen and fresh embryo transfer: A Chinese cohort study. *PLoS Medicine*, 21(6 June). <https://doi.org/10.1371/journal.pmed.1004388>

Impact of long-term cryopreservation of gametes and embryos

Scope: Following [amendments](#) to HFE Act (1990), storage of eggs, sperm and/or embryos for use in a patient's own treatment or for donation is permitted for up to 55 years. This topic monitors any safety or viability concerns relating to the keeping of gametes or embryos in long-term storage. As the storage extension applies to gametes preserved within tissue, the impact of long-term cryopreservation of ovarian and testicular tissue is also monitored within this topic. Monitoring of the safety of cryopreservation is also relevant to maintaining the [authorised processes](#) list. Studies looking at utilisation rates of cryopreserved gametes are additionally included.

Cobo, A., Coello, A., De Los Santos, M. J., Remohi, J., & Bellver, J. (2024). Embryo long-term storage does not affect assisted reproductive technologies outcome: analysis of 58,001 vitrified blastocysts over 11 years. *American Journal of Obstetrics and Gynecology*, 231(2), 238.e1-238.e11. <https://doi.org/10.1016/j.ajog.2024.03.033>

Kazorgah, F. M., Govahi, A., Dadseresht, A., Kenari, F. N. P., Ajdary, M., Mehdizadeh, R., Derakhshan, R., & Mehdizadeh, M. (2024). The effect of temperature and storage time on DNA integrity after freeze-drying sperm from individuals with normozoospermia. *Clinical and Experimental Reproductive Medicine*, 51(1), 42–47. <https://doi.org/10.5653/cerm.2023.06093>

Liang, M.-Y., Lin, M., Qin, X., Yang, R., Hu, K.-L., & Li, R. (2024). Long-term embryo vitrification is associated with reduced success rates in women undergoing frozen embryo transfer following a failed fresh cycle. *European Journal of Obstetrics & Gynecology and Reproductive Biology*, 296, 244–249. <https://doi.org/10.1016/j.ejogrb.2024.03.002>

Stigliani, S., Amaro, A., Reggiani, F., Maccarini, E., Massarotti, C., Lambertini, M., Anserini, P., & Scaruffi, P. (2024). The storage time of cryopreserved human spermatozoa does not affect pathways involved in fertility. *Basic and Clinical Andrology*, 34(1), 15. <https://doi.org/10.1186/s12610-024-00231-4>

Wang, X., Xiao, Y., Sun, Z., & Xiong, W. (2024). Effect of post-vitrification cryopreservation duration on singleton birth-weight in frozen-thawed blastocysts transfer cycles. *Frontiers in Endocrinology*, 15. <https://doi.org/10.3389/fendo.2024.1366360>

Weibring, K., Lundberg, F. E., Cohn-Cedermark, G., & Rodriguez-Wallberg, K. A. (2024). Parenthood in a Swedish prospective cohort of 1,378 adolescents and young adults banking semen for fertility preservation at time of cancer diagnosis. *Frontiers in Endocrinology*, 15. <https://doi.org/10.3389/FENDO.2024.1502479>

White, J., Jackson, A., Druce, I., & Gale, J. (2024). Oocyte cryopreservation and reciprocal in vitro fertilization in a transgender man on long term testosterone gender-affirming hormone therapy: a case report. *F&S Reports*, 5(1), 111–113. <https://doi.org/10.1016/j.xfre.2023.11.004>

Zhan, S., Lin, C., Lin, Q., Gan, J., Wang, C., Luo, Y., Liu, J., Du, H., & Liu, H. (2024). Vitrification preservation of good-quality blastocysts for more than 5 years reduces implantation and live birth rates. *Human Reproduction*, 39(9), 1960–1968. <https://doi.org/10.1093/humrep/deae150>

Zhu, L., Sun, L., Liu, W., Han, W., Huang, G., & Li, J. (2024). Long-term storage does not affect the DNA methylation profiles of vitrified-warmed human embryos. *Molecular Reproduction and Development*, 91(8). <https://doi.org/10.1002/mrd.23713>

Impact of stress on fertility treatment outcomes

Scope: This topic looks at the impact of stress in people undergoing ART on fertility treatment outcomes. Research that does not examine ART outcomes is excluded. There are two main areas of research: (1) The impact of psychological factors including stress on treatment outcomes and (2) the impact of interventions to manage stress on treatment outcomes. Research on the impact of the stress hormone cortisol on treatment outcomes is included.

Bian, C., Cao, J., Chen, K., Xia, X., & Yu, X. (2024). Effectiveness of psychological interventions on pregnancy rates in infertile women undergoing assisted reproductive technologies: a meta-analysis of

randomised controlled trials. *Biotechnology and Genetic Engineering Reviews*, 40(4), 4512–4531. <https://doi.org/10.1080/02648725.2023.2213080>

Li, Y., McLeish, J., Hardy, P., Cole, C., Carson, C., Alderdice, F., & Maheshwari, A. (2024). Anxiety in couples undergoing IVF: evidence from E-Freeze randomised controlled trial. *Human Reproduction Open*, 2024(3). <https://doi.org/10.1093/hropen/hoae037>

Li, Y., Shen, W., Mo, F., Ma, Q., & Xing, L. (2024). The Effect of Auricular Acupressure on Women Psychological Distress during Controlled Ovarian Hyperstimulation for in vitro Fertilization: A Single-Blind, Randomized, and Sham-Controlled Study. *The Tohoku Journal of Experimental Medicine*, 2024.J076. <https://doi.org/10.1620/tjem.2024.J076>

Liao, T., Gao, Y., Yang, X., Tang, Y., Wang, B., Yang, Q., Gao, X., Tang, Y., He, K., Shen, J., Bao, S., Pan, G., Zhu, P., Tao, F., & Shao, S. (2024). Preconception depression reduces fertility: a couple-based prospective preconception cohort. *Human Reproduction Open*, 2024(3). <https://doi.org/10.1093/hropen/hoae032>

Lvy, Y., Zhang, F., Cai, Z., Zhong, D., & Xing, L. (2024). Correlation among irrational parenthood cognitions, fertility stress, and social support in patients with repeated implantation failure and the mediating effect of fertility stress: a cross-sectional survey. *Journal of Assisted Reproduction and Genetics*, 41(1), 205–212. <https://doi.org/10.1007/s10815-023-02953-2>

Mínguez-Alarcón, L., Williams, P. L., Souter, I., Ford, J. B., Hauser, R., & Chavarro, J. E. (2024). Women's preconception psychological stress and birth outcomes in a fertility clinic: the EARTH study. *Frontiers in Global Women's Health*, 5. <https://doi.org/10.3389/fgwh.2024.1293255>

Nikolaeva, M., Arefieva, A., Babayan, A., Aksenov, V., Zhukova, A., Kalinina, E., Krechetova, L., & Sukhikh, G. (2024). Stress Biomarkers Transferred Into the Female Reproductive Tract by Seminal Plasma Are Associated with ICSI Outcomes. *Reproductive Sciences*, 31(6), 1732–1746. <https://doi.org/10.1007/s43032-024-01486-y>

Nunes, G. M., Paiva, S. de P. C., Geber, S., Serra, A. S. V. de A., Sampaio, M. A. C., & Tavares, R. L. C. (2024). The impact of extremely brief meditation and brief mindfulness interventions on assisted reproductive technologies success rates: A randomised controlled trial. *EXPLORE*, 20(6), 103067. <https://doi.org/10.1016/j.explore.2024.103067>

Orvieto, R., Shamir, C., & Aizer, A. (2024). Does extreme psychological burden (Hamas terrorist attack on October 7th, 2023) affect in vitro fertilization outcome? *Journal of Assisted Reproduction and Genetics*, 41(6), 1585–1588. <https://doi.org/10.1007/s10815-024-03099-5>

Swift, A., Thomas, E., Larson, K., Swanson, M., & Fernandez-Pineda, M. (2024). Infertility-related stress, quality of life, and reasons for fertility treatment discontinuation among US women: A secondary analysis of a cross-sectional study. *Sexual & Reproductive Healthcare*, 39, 100955. <https://doi.org/10.1016/j.srhc.2024.100955>

Zanettoullis, A. T., Mastorakos, G., Vakas, P., Vlahos, N., & Valsamakis, G. (2024). Effect of Stress on Each of the Stages of the IVF Procedure: A Systematic Review. *International Journal of Molecular Sciences*, 25(2), 726. <https://doi.org/10.3390/ijms25020726>

Zhai, J., Zhao, S., & Hao, G. (2024). The impact of sociocultural and psychological stress on the outcome of assisted reproductive technology in remarried families. *Journal of Psychosomatic Obstetrics & Gynecology*, 45(1). <https://doi.org/10.1080/0167482X.2024.2351809>

Impact of the microbiome on fertility and fertility treatment outcomes

Scope: Research into the microbiome with relevance to fertility treatment broadly falls into three areas: (1) understanding the normative microbiota composition of the reproductive tract, (2) understanding an association between reproductive tract microbiota and its role in fertility/infertility, (3) developing interventions to improve fertility and fertility treatment outcomes. Lack of standardisation in the methodological processes has limited research applications, however, commercial tests are being developed for distribution on the UK fertility market. In time, microbiome testing and interventions may be considered for an add-ons rating.

- Alqawasmeh, O. A. M., Jiang, X. T., Cong, L., Wu, W., Leung, M. B. W., Chung, J. P. W., Yim, H. C. H., Fok, E. K. L., & Chan, D. Y. L. (2024). Vertical transmission of microbiomes into embryo culture media and its association with assisted reproductive outcomes. *Reproductive Biomedicine Online*, 49(2). <https://doi.org/10.1016/J.RBMO.2024.103977>
- Ashonibare, V. J., Akorede, B. A., Ashonibare, P. J., Akhigbe, T. M., & Akhigbe, R. E. (2024). Gut microbiota-gonadal axis: the impact of gut microbiota on reproductive functions. *Frontiers in Immunology*, 15. <https://doi.org/10.3389/FIMMU.2024.1346035>
- Atay, R., & Hacıoglu, O. (2024). Determination of microbiota awareness levels in women planning pregnancy. *Revista Da Associacao Medica Brasileira (1992)*, 70(5). <https://doi.org/10.1590/1806-9282.20231401>
- Bai, S., Xu, G., Mo, H., Qi, T., Fu, S., Zhu, L., Huang, B., Zhang, J., & Chen, H. (2024). Investigating into microbiota in the uterine cavity of the unexplained recurrent pregnancy loss patients in early pregnancy. *Placenta*, 152, 1–8. <https://doi.org/10.1016/J.PLACENTA.2024.05.125>
- Balla, B., Illés, A., Tobiás, B., Pikó, H., Beke, A., Sipos, M., Lakatos, P., & Kósa, J. P. (2024). The Role of the Vaginal and Endometrial Microbiomes in Infertility and Their Impact on Pregnancy Outcomes in Light of Recent Literature. *International Journal of Molecular Sciences*, 25(23). <https://doi.org/10.3390/IJMS252313227>
- Bekon, A. M., Martiny, D., Thomas, A. L., Miendje, Y., Debyttère, A. L., Autin, C., & Bertrand, E. (2024a). Optimization of the microbiological treatment of sperm for assisted reproductive technology (ART). *Journal of Microbiological Methods*, 224. <https://doi.org/10.1016/J.MIMET.2024.107004>
- Bekon, A. M., Martiny, D., Thomas, A. L., Miendje, Y., Debyttère, A. L., Autin, C., & Bertrand, E. (2024b). Optimization of the microbiological treatment of sperm for assisted reproductive technology (ART). *Journal of Microbiological Methods*, 224. <https://doi.org/10.1016/J.MIMET.2024.107004>
- Bellver, J., Gonzalez-Monfort, M., González, S., Toson, B., Labarta, E., Castellón, G., Mariani, G., Vidal, C., Giles, J., Cruz, F., Ballesteros, A., Ferrando, M., García-Velasco, J. A., Valbuena, D., Vilella, F., Parras-Molto, M., Tercero-Atencia, E., Simon, C., & Moreno, I. (2024). An Analysis of the Digestive and Reproductive Tract Microbiota in Infertile Women with Obesity. *International Journal of Molecular Sciences*, 25(23). <https://doi.org/10.3390/IJMS252312600>
- Cao, W., Fu, X., Zhou, J., Qi, Q., Ye, F., Li, L., & Wang, L. (2024). The effect of the female genital tract and gut microbiome on reproductive dysfunction. *Bioscience Trends*, 17(6), 458–474. <https://doi.org/10.5582/BST.2023.01133>
- Cariati, F., Conforti, A., Esteves, S. C., & Alviggi, C. (2024). Editorial: Reproductive microbiome and its interplay with the environment. *Frontiers in Endocrinology*, 15. <https://doi.org/10.3389/FENDO.2024.1470608>
- Chen, H., Tu, Y., Zhang, C., Li, J., Wu, T., Liu, S., He, L., Zhang, A., Li, Y., Li, L., Sui, Y., Wang, L., Chen, X., Xi, J., Wu, Y., Jin, L., & Huang, H. F. (2023). Effect of transvaginal *Lactobacillus* supplementation on reversing lower genital tract dysbiosis and improving perinatal outcomes in PCOS patients after IVF-FET: a study protocol for a multicenter randomized controlled trial. *Trials*, 24(1). <https://doi.org/10.1186/S13063-023-07825-9>
- Chopra, C., Kumar, V., Kumar, M., & Bhushan, I. (2024). Role of vaginal microbiota in idiopathic infertility: a prospective study. *Microbes and Infection*, 26(4). <https://doi.org/10.1016/J.MICINF.2024.105308>
- Davies, R., Minhas, S., & Jayasena, C. N. (2023). Next-Generation Sequencing to Elucidate the Semen Microbiome in Male Reproductive Disorders. *Medicina (Kaunas, Lithuania)*, 60(1). <https://doi.org/10.3390/MEDICINA60010025>
- Deng, R., Huang, Y., Tian, Z., & Zeng, Q. (2024). Association between gut microbiota and male infertility: a two-sample Mendelian randomization study. *International Microbiology : The Official Journal of the Spanish Society for Microbiology*, 27(6). <https://doi.org/10.1007/S10123-024-00512-Y>
- Dubey, I., K, N., G, V., Rohilla, G., Lalruatmawii, Naxine, P., P, J., Rachamalla, M., & Kushwaha, S. (2024). Exploring the hypothetical links between environmental pollutants, diet, and the gut-testis axis: The

- potential role of microbes in male reproductive health. *Reproductive Toxicology* (Elmsford, N.Y.), 130. <https://doi.org/10.1016/J.REPROTOX.2024.108732>
- Gao, H., Liu, Q., Wang, X., Li, T., Li, H., Li, G., Tan, L., & Chen, Y. (2024). Deciphering the role of female reproductive tract microbiome in reproductive health: a review. *Frontiers in Cellular and Infection Microbiology*, 14. <https://doi.org/10.3389/FCIMB.2024.1351540>
- Gao, X., Louwers, Y. V., Laven, J. S. E., & Schoenmakers, S. (2024). Clinical Relevance of Vaginal and Endometrial Microbiome Investigation in Women with Repeated Implantation Failure and Recurrent Pregnancy Loss. *International Journal of Molecular Sciences*, 25(1). <https://doi.org/10.3390/IJMS25010622>
- Grande, G., Graziani, A., De Toni, L., Garolla, A., & Ferlin, A. (2024). Male Tract Microbiota and Male Infertility. *Cells*, 13(15). <https://doi.org/10.3390/CELLS13151275>
- Han, X., Tian, H., Yang, L., & Ji, Y. (2024). Bidirectional Mendelian randomization to explore the causal relationships between the gut microbiota and male reproductive diseases. *Scientific Reports*, 14(1). <https://doi.org/10.1038/S41598-024-69179-9>
- Hao, Y., Du, X., Cai, C., Zhao, Y., & Ren, Y. (2025). Ammonia and hydrogen sulfide - new insights into gut microbiota and male infertility through meta-analysis. *Frontiers in Cellular and Infection Microbiology*, 14. <https://doi.org/10.3389/FCIMB.2024.1449453>
- Jendraszak, M., Skibińska, I., Kotwicka, M., & Andrusiewicz, M. (2024). The elusive male microbiome: revealing the link between the genital microbiota and fertility. Critical review and future perspectives. *Critical Reviews in Clinical Laboratory Sciences*, 61(7). <https://doi.org/10.1080/10408363.2024.2331489>
- Junxian, H., Chen, P., & Liu, G. (2024). ASSOCIATION BETWEEN SEMEN MICROBIOME DISORDER AND SPERM DNA DAMAGE. *Fertility and Sterility*, 122(4), e432. <https://doi.org/10.1016/J.FERTNSTERT.2024.08.295>
- Kadogami, D., Kimura, F., Hanada, T., Tsuji, S., Nakaoka, Y., Murakami, T., & Morimoto, Y. (2023). Impact of *Lactobacillus* in the uterine microbiota on in vitro fertilization outcomes. *Journal of Reproductive Immunology*, 160. <https://doi.org/10.1016/J.JRI.2023.104138>
- Leclaire, S., Bandekar, M., Rowe, M., Ritari, J., Jokiniemi, A., Partanen, J., Allinen, P., Kuusipalo, L., & Kekäläinen, J. (2024). Female reproductive tract microbiota varies with MHC profile. *Proceedings. Biological Sciences*, 291(2033), 20241334. <https://doi.org/10.1098/RSPB.2024.1334>
- Li, B., Xiong, Y., Guo, D., Deng, G., & Wu, H. (2025). The gut-reproductive axis: Bridging microbiota balances to reproductive health and fetal development. *International Immunopharmacology*, 144. <https://doi.org/10.1016/J.INTIMP.2024.113627>
- Maksimovic Celicanin, M., Haahr, T., Humaidan, P., & Skaft-Holm, A. (2024a). Vaginal dysbiosis - the association with reproductive outcomes in IVF patients: a systematic review and meta-analysis. *Current Opinion in Obstetrics & Gynecology*, 36(3), 155–164. <https://doi.org/10.1097/GCO.0000000000000953>
- Maksimovic Celicanin, M., Haahr, T., Humaidan, P., & Skaft-Holm, A. (2024b). Vaginal dysbiosis - the association with reproductive outcomes in IVF patients: a systematic review and meta-analysis. *Current Opinion in Obstetrics & Gynecology*, 36(3), 155–164. <https://doi.org/10.1097/GCO.0000000000000953>
- Maldonado-Barrueco, A., Almazán-Garate, E., Armijo-Suárez, O., Iniesta-Pérez, S., Sanz-González, C., Falces-Romero, I., Álvarez-López, C., Cacho-Calvo, J., & Quiles-Melero, I. (2024). Utility of culture and molecular methods using Allplex™ Bacterial Vaginosis Plus Assay (Seegene®) as a tool for endometriosis, infertility and recurrent pregnancy loss diagnosis. *Diagnostic Microbiology and Infectious Disease*, 110(3). <https://doi.org/10.1016/J.DIAGMICROBIO.2024.116437>
- Mongane, J., Hendwa, E., Sengeyi, D., Kajibwami, E., Kampara, F., Chentwali, S., Kalegamire, C., Barhishindi, I., Kujirakwinja, Y., Maningo, J. B., Kasago, B., & Mulinganya, G. (2024). Association between bacterial vaginosis, *Chlamydia trachomatis* infection and tubal factor infertility in Bukavu, Democratic Republic of Congo. *BMC Infectious Diseases*, 24(1). <https://doi.org/10.1186/S12879-024-09379-W>
- Odendaal, J., Black, N., Bennett, P. R., Brosens, J., Quenby, S., & MacIntyre, D. A. (2024). The endometrial microbiota and early pregnancy loss. *Human Reproduction* (Oxford, England), 39(4), 638–646. <https://doi.org/10.1093/HUMREP/DEAD274>

- Pérez-Prieto, I., Rodríguez-Santisteban, A., & Altmäe, S. (2024). Beyond the reproductive tract: gut microbiome and its influence on gynecological health. *Current Opinion in Clinical Nutrition and Metabolic Care*, 36(3), 134–147. <https://doi.org/10.1097/01.gco.0001016536.06312.72>
- Santana, A. S. A., & Póvoa, A. M. (2024). Female genital tract microbiome: the influence of probiotics on assisted reproduction. *Revista Brasileira de Ginecologia e Obstetricia : Revista Da Federacao Brasileira Das Sociedades de Ginecologia e Obstetricia*, 46. <https://doi.org/10.61622/RBGO/2024RBGO82>
- Teh, H. E., Pung, C. K., Arasoo, V. J. T., & Yap, P. S. X. (2024). A Landscape View of the Female Genital Tract Microbiome in Healthy Controls and Women With Reproductive Health Conditions Associated With Ectopic Pregnancy. *British Journal of Biomedical Science*, 80. <https://doi.org/10.3389/BJBS.2023.12098>
- Tian, Q., Jin, S., Zhang, G., Liu, Y., Liu, J., Tang, X., Li, Y., Liu, J., Liu, Y., & Wang, Z. (2024). Assessing vaginal microbiome through Vaginal Microecology Evaluation System as a predictor for in vitro fertilization outcomes: a retrospective study. *Frontiers in Endocrinology*, 15. <https://doi.org/10.3389/FENDO.2024.1380187>
- Tian, Z., Zhao, M., Sui, X., Li, X., Qin, L., Chen, Z. J., Zhao, S., & Zhao, H. (2024). Associations between vaginal microbiota and endometrial polypoid lesions in women of reproductive age: a cross-sectional study. *Reproductive Biomedicine Online*, 48(2). <https://doi.org/10.1016/J.RBMO.2023.103602>
- Väinämö, S., Saqib, S., Kalliala, I., Kervinen, K., Luiro, K., Niinimäki, M., Halttunen-Nieminen, M., Virtanen, S., Nieminen, P., Salonen, A., & Holster, T. (2023). Longitudinal analysis of vaginal microbiota during IVF fresh embryo transfer and in early pregnancy. *Microbiology Spectrum*, 11(6). <https://doi.org/10.1128/SPECTRUM.01650-23>
- van den Tweel, M. M., van den Munckhof, E. H. A., van der Zanden, M., Molijn, A. C., van Lith, J. M. M., Le Cessie, S., & Boers, K. E. (2024). Bacterial vaginosis in a subfertile population undergoing fertility treatments: a prospective cohort study. *Journal of Assisted Reproduction and Genetics*, 41(2), 441–450. <https://doi.org/10.1007/S10815-023-03000-W>
- van den Tweel, M. M., van den Munckhof, E. H. A., van der Zanden, M., Molijn, A., van Lith, J. M. M., & Boers, K. E. (2024). The Vaginal Microbiome Changes During Various Fertility Treatments. *Reproductive Sciences (Thousand Oaks, Calif.)*, 31(6), 1593–1600. <https://doi.org/10.1007/S43032-024-01484-0>
- van den Tweel, M., van den Munckhof, E., van der Zanden, M., Le Cessie, S., van Lith, J., & Boers, K. (2024). Testing on bacterial vaginosis in a subfertile population and time to pregnancy: a prospective cohort study. *Archives of Gynecology and Obstetrics*, 310(2), 1245–1253. <https://doi.org/10.1007/S00404-024-07542-X>
- Vomstein, K., Krog, M. C., Wrønding, T., & Nielsen, H. S. (2024). The microbiome in recurrent pregnancy loss - A scoping review. *Journal of Reproductive Immunology*, 163. <https://doi.org/10.1016/J.JRI.2024.104251>
- Wang, M., Zheng, L. W., Ma, S., Zhao, D. H., & Xu, Y. (2024). The gut microbiota: emerging biomarkers and potential treatments for infertility-related diseases. *Frontiers in Cellular and Infection Microbiology*, 14, 1450310. <https://doi.org/10.3389/FCIMB.2024.1450310>
- Wang, Y., Li, W., & Ha, C. (2024). A large-scale causal analysis of gut microbiota and endometriosis associated infertility: A Mendelian randomization study. *Medicine*, 103(12), E37383. <https://doi.org/10.1097/MD.00000000000037383>
- Wei, Q., Chen, H., Zou, H., Zhang, H., Liu, S., Zheng, J., Zhang, S., & Hu, L. (2024). Impact of vaginal microecological differences on pregnancy outcomes and endometrial microbiota in frozen embryo transfer cycles. *Journal of Assisted Reproduction and Genetics*, 41(4), 929–938. <https://doi.org/10.1007/S10815-024-03066-0>
- Wu, N., Liu, J., Sun, Y., Fan, X., Zang, T., Richardson, B. N., Bai, J., Xianyu, Y., & Liu, Y. (2024). Alterations of the gut microbiota and fecal short-chain fatty acids in women undergoing assisted reproduction. *Reproduction, Fertility, and Development*, 36(3). <https://doi.org/10.1071/RD23096>
- Xiao, L., Zuo, Z., & Zhao, F. (2024). Microbiome in Female Reproductive Health: Implications for Fertility and Assisted Reproductive Technologies. *Genomics, Proteomics & Bioinformatics*, 22(1). <https://doi.org/10.1093/GPBJNL/QZAD005>

Zhang, H., Zou, H., Zhang, C., & Zhang, S. (2024). Chronic endometritis and the endometrial microbiota: implications for reproductive success in patients with recurrent implantation failure. *Annals of Clinical Microbiology and Antimicrobials*, 23(1). <https://doi.org/10.1186/S12941-024-00710-6>

Zhang, J. X., Li, Q. L., Wang, X. Y., Zhang, C. C., Chen, S. T., Liu, X. H., Dong, X. Y., Zhao, H., & Huang, D. H. (2024). Causal Link between Gut Microbiota and Infertility: A Two-sample Bidirectional Mendelian Randomization Study. *Current Medical Science*, 44(6). <https://doi.org/10.1007/S11596-024-2931-X>

Zhao, X., Shi, W., Li, Z., & Zhang, W. (2024). Linking reproductive tract microbiota to premature ovarian insufficiency: Pathophysiological mechanisms and therapies. *Journal of Reproductive Immunology*, 166. <https://doi.org/10.1016/J.JRI.2024.104325>

Zou, H., Xu, N., Xu, H., Xing, X., Chen, Y., & Wu, S. (2024a). Inflammatory cytokines may mediate the causal relationship between gut microbiota and male infertility: a bidirectional, mediating, multivariate Mendelian randomization study. *Frontiers in Endocrinology*, 15. <https://doi.org/10.3389/FENDO.2024.1368334>

Zou, H., Xu, N., Xu, H., Xing, X., Chen, Y., & Wu, S. (2024b). Inflammatory cytokines may mediate the causal relationship between gut microbiota and male infertility: a bidirectional, mediating, multivariate Mendelian randomization study. *Frontiers in Endocrinology*, 15. <https://doi.org/10.3389/FENDO.2024.1368334>

In vitro derived gametes

Scope: This topic encompasses research into methods for achieving in vitro gametogenesis (organoids, 3D systems, scaffolds, etc) in both human and animal models. This includes the use of embryonic stem cells, induced pluripotent stem cells, immature gametes, and studies which attempt to recreate early stages of gametogenesis (ie primordial germ cells and primordial germ cell-like cells, and their induction to germ cells). In vitro maturation is also considered within scope. Opinion articles on the ethical and legal perspectives relevant to this topic are included.

Adashi, E. Y., Hayashi, K., & Cohen, I. G. (2024). Ethical and legal challenges in assisted same-sex conception through in vitro gametogenesis. In *Nature Medicine* (Vol. 30, Issue 2, pp. 322–323). Nature Research. <https://doi.org/10.1038/s41591-023-02689-7>

Aizawa, E., Peters, A. H. F. M., & Wutz, A. (2024). In vitro gametogenesis: Towards competent oocytes: Limitations and future improvements for generating oocytes from pluripotent stem cells in culture. *BioEssays : News and Reviews in Molecular, Cellular and Developmental Biology*, e2400106. <https://doi.org/10.1002/bies.202400106>

Akyash, F., Aflatoonian, R., Farashahi-Yazd, E., Hajizadeh-Tafti, F., Golzadeh, J., Sadat Tahajjodi, S., & Aflatoonian, B. (2024). Testicular Cells Derived Conditioned Medium Supports Germ Cell Differentiation of Human Embryonic Stem Cells. *Royan Institute Cell Journal (Yakhteh)*, 26(6), 370–379. <https://doi.org/10.22074/CELLJ.2024.2012768.1419>

Alves-Lopes, J. P., Wong, F. C. K., & Surani, M. A. (2024). Human primordial germ cell-like cells specified from resetting precursors develop in human hindgut organoids. *Nature Protocols*, 19(4), 1149–1182. <https://doi.org/10.1038/s41596-023-00945-1>

Aponte, P. M., Gutierrez-Reinoso, M. A., & Garcia-Herreros, M. (2024). Bridging the Gap: Animal Models in Next-Generation Reproductive Technologies for Male Fertility Preservation. In *Life* (Vol. 14, Issue 1). Multidisciplinary Digital Publishing Institute (MDPI). <https://doi.org/10.3390/life14010017>

Asgari, F., Asgari, H., Najafi, M., Hajiaghalou, S., Pirhajati-Mahabadi, V., Mohammadi, A., Gholipourmalekabadi, M., & Koruji, M. (2024). In vitro proliferation and differentiation of mouse spermatogonial stem cells in decellularized human placenta matrix. *Journal of Biomedical Materials Research - Part B Applied Biomaterials*, 112(5). <https://doi.org/10.1002/jbm.b.35414>

Choong, E., Dawson, E. P., Bowman, K., & Adashi, E. Y. (2024). In vitro gametogenesis (IVG): reflections from a workshop. *Journal of Assisted Reproduction and Genetics*. <https://doi.org/10.1007/s10815-024-03266-8>

- Damyanova, K. B., Nixon, B., Johnston, S. D., Gambini, A., Benitez, P. P., & Lord, T. (2024). Spermatogonial stem cell technologies: applications from human medicine to wildlife conservation. *Biology of Reproduction*. <https://doi.org/10.1093/biolre/ioae109>
- Di Berardino, C., Peserico, A., Camerano Spelta Rapini, C., Liverani, L., Capacchietti, G., Russo, V., Berardinelli, P., Unalan, I., Damian-Buda, A. I., Boccaccini, A. R., & Barboni, B. (2024). Bioengineered 3D ovarian model for long-term multiple development of preantral follicle: bridging the gap for poly(ϵ -caprolactone) (PCL)-based scaffold reproductive applications. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01266-y>
- Esfahani, S. N., Zheng, Y., Arabpour, A., Irizarry, A. M. R., Kobayashi, N., Xue, X., Shao, Y., Zhao, C., Agranonik, N. L., Sparrow, M., Hunt, T. J., Faith, J., Lara, M. J., Wu, Q. Y., Silber, S., Petropoulos, S., Yang, R., Chien, K. R., Clark, A. T., & Fu, J. (2024). Derivation of human primordial germ cell-like cells in an embryonic-like culture. *Nature Communications*, 15(1). <https://doi.org/10.1038/s41467-023-43871-2>
- Frost, E. R., & Gilchrist, R. B. (2024). Making human eggs in a dish: are we close? In *Trends in Biotechnology* (Vol. 42, Issue 2, pp. 168–178). Elsevier Ltd. <https://doi.org/10.1016/j.tibtech.2023.07.007>
- Ghaleno, L. R., Hajari, M. A., Choshali, M. A., Heidari, E. A., Shahverdi, A., Alipour, H., & Valojerdi, M. R. (2024). Hyaluronic acid-alginate hydrogel stimulates the differentiation of neonatal mouse testicular cells into hepatocyte-like and other cell lineages in three-dimensional culture. *Biology of the Cell*. <https://doi.org/10.1111/BOC.202400049>
- Gizer, M., Önen, S., & Korkusuz, P. (2024). The Evolutionary Route of in vitro Human Spermatogenesis: What is the Next Destination? *Stem Cell Reviews and Reports*. <https://doi.org/10.1007/s12015-024-10726-2>
- Hashemi, E., Movahedin, M., Ghiaseddin, A., & Aghamir, S. M. K. (2024). In Vitro Spermatogenesis on Human Decellularized Testicular Matrix Plates Following Exosome Treatment in a Dynamic Culture System. *Stem Cell Reviews and Reports*. <https://doi.org/10.1007/s12015-024-10818-z>
- Jang, S. W., Kim, Y. R., Han, J. H., Jang, H., & Choi, H. W. (2024). Generation of mouse and rat xenogeneic ovaries in vitro for production of mouse oocyte. *Animal Cells and Systems*, 28(1), 303–314. <https://doi.org/10.1080/19768354.2024.2363601>
- Khampang, S., Lorthongpanich, C., Laowtammathron, C., Klaihmon, P., Meesa, S., Suksomboon, W., Jiamvoraphong, N., Kheolamai, P., Luanpitpong, S., Easley, C. A., Mahyari, E., & Issaragrisil, S. (2024). The dynamic expression of YAP is essential for the development of male germ cells derived from human embryonic stem cells. *Scientific Reports*, 14(1). <https://doi.org/10.1038/s41598-024-66852-x>
- Kwaspen, L., Kanbar, M., & Wyns, C. (2024). Mapping the Development of Human Spermatogenesis Using Transcriptomics-Based Data: A Scoping Review. In *International Journal of Molecular Sciences* (Vol. 25, Issue 13). Multidisciplinary Digital Publishing Institute (MDPI). <https://doi.org/10.3390/ijms25136925>
- Le Goff, A., Jeffries Hein, R., Hart, A. N., Roberson, I., & Landecker, H. L. (2024). Anticipating in vitro gametogenesis: Hopes and concerns for IVG among diverse stakeholders. *Stem Cell Reports*, 19(7), 933–945. <https://doi.org/10.1016/j.stemcr.2024.05.002>
- Lin, Y. H., Lehle, J. D., & McCarrey, J. R. (2024). Source cell-type epigenetic memory persists in induced pluripotent cells but is lost in subsequently derived germline cells. *Frontiers in Cell and Developmental Biology*, 12. <https://doi.org/10.3389/fcell.2024.1306530>
- Mikhailchenko, A., Marti Gutierrez, N., Frana, D., Safaei, Z., Dyken, C. Van, Li, Y., Ma, H., Koski, A., Liang, D., Lee, S.-G., Amato, P., & Mitalipov, S. (2024). Induction of somatic cell haploidy by premature cell division. In *Sci. Adv* (Vol. 10). <https://www.science.org>
- Mohammadi, A., Koruji, M., Azami, M., Shabani, R., Mohandesnezhad, S., Bashiri, Z., & Asgari, H. (2024). Polycaprolactone/Testicular Extracellular Matrix/Graphene Oxide-Based Electrospun Tubular Scaffolds for Reproductive Medicine: Biomimetic Architecture of Seminiferous Tubules. *Macromolecular Bioscience*, 24(2). <https://doi.org/10.1002/mabi.202300342>
- Murase, Y., Yokogawa, R., Yabuta, Y., Nagano, M., Katou, Y., Mizuyama, M., Kitamura, A., Puangsricharoen, P., Yamashiro, C., Hu, B., Mizuta, K., Tsujimura, T., Yamamoto, T., Ogata, K., Ishihama,

Y., & Saitou, M. (2024). In vitro reconstitution of epigenetic reprogramming in the human germ line. *Nature*, 631(8019), 170–178. <https://doi.org/10.1038/s41586-024-07526-6>

Nguyen, A.-L. V., Julian, S., Weng, N., & Flannigan, R. (2024). Advances in human In vitro spermatogenesis: A review. *Molecular Aspects of Medicine*, 100, 101320. <https://doi.org/10.1016/j.mam.2024.101320>

Ogawa, T., Matsumura, T., Yao, T., Kimura, H., Hashimoto, K., Ishikawa-Yamauchi, Y., & Sato, T. (2024). Improvements in in vitro spermatogenesis: oxygen concentration, antioxidants, tissue-form design, and space control. In *Journal of Reproduction and Development* (Vol. 70, Issue 1).

Richer, G., Goyvaerts, C., Davida Marchandise, L., Vanhaecke, T., Goossens, E., & Baert, Y. (2024). Spermatogenesis in mouse testicular organoids with testis-specific architecture, improved germ cell survival and testosterone production. *Biofabrication*. <https://doi.org/10.1088/1758-5090/ad618f>

Richer, G., Vanhaecke, T., Rogiers, V., Goossens, E., & Baert, Y. (2024). Mouse In Vitro Spermatogenesis on 3D Bioprinted Scaffolds. In *Methods in Molecular Biology* (Vol. 2770, pp. 135–149). Humana Press Inc. https://doi.org/10.1007/978-1-0716-3698-5_11

Salem M, Khadivi F, Feizollahi N, Khodarahmian M, Saedi Marghmaleki M, Ayub S, Chegini R, Bashiri Z, Abbasi Y, Naji M, & Abbasi M. (2024). Melatonin Promotes Differentiation of Human Spermatogonial Stem Cells Cultured on Three-Dimensional Decellularized Human Testis Matrix. *Urology Journal*, 21(4), 250–264.

Shirasawa, A., Hayashi, M., Shono, M., Ideta, A., Yoshino, T., & Hayashi, K. (2024). Efficient derivation of embryonic stem cells and primordial germ cell-like cells in cattle. In *Journal of Reproduction and Development* (Vol. 70, Issue 2).

Silber, S. J., Goldsmith, S., Castleman, L., & Hayashi, K. (2024). In Vitro Maturation, In Vitro Oogenesis, and Ovarian Longevity. *Reproductive Sciences*, 31(5), 1234–1245. <https://doi.org/10.1007/s43032-023-01427-1>

Tan, J., Li, J., Zhang, Y., Li, X., Han, S., Li, Z., & Zhou, X. (2024). Application of photocrosslinked gelatin, alginate and dextran hydrogels in the in vitro culture of testicular tissue. *International Journal of Biological Macromolecules*, 260. <https://doi.org/10.1016/j.ijbiomac.2024.129498>

Villalba, A. (2024). Artificial Gametes and Human Reproduction in the 21st Century: An Ethical Analysis. In *Reproductive Sciences*. Springer Nature. <https://doi.org/10.1007/s43032-024-01558-z>

von Rohden, E., Jensen, C. F. S., Andersen, C. Y., Sønksen, J., Fedder, J., Thorup, J., Ohl, D. A., Fode, M., Hoffmann, E. R., & Mamsen, L. S. (2024). Male fertility restoration: In vivo and In vitro stem cell-based strategies using cryopreserved testis tissue – A scoping review. *Fertility and Sterility*. <https://doi.org/10.1016/j.fertnstert.2024.07.010>

Mitochondrial donation

Scope: Under the HFE (Mitochondrial Donation) Regulations, maternal spindle transfer (MST) and pronuclear transfer (PNT) can be performed with HFEA approval in the UK. Relevant research describing the application of MST or PNT in the context of mitochondrial donation to inform SCAAC of its application and safety are the focus of this topic, including approaches to refine techniques to minimise mitochondrial heteroplasmy. The development of alternative techniques for preventing the inheritance of mitochondrial disease is also monitored within this topic, including both mitochondrial replacement techniques and *applications* of mitochondrial genome editing techniques. Expansion of mitochondrial donation techniques as a treatment for infertility (not indicated by mitochondrial disease) are additionally included in the scope of this topic, for example autologous cytoplasmic transfer.

Gil, J., Nohales, M., Ortega-Jaen, D., Martin, A., Pardiñas, M. L., Serra, V., Labarta, E., & de los Santos, M. J. (2024). Impact of autologous mitochondrial transfer on obstetric and neonatal health of offspring: A small single-center case series. *Placenta*, 158, 217–222. <https://doi.org/10.1016/J.PLACENTA.2024.10.007>

Hui, L., Hayman, P., Buckland, A., Fahey, M. C., Mackey, D. A., Mallett, A. J., Schweitzer, D. R., Stuart, C. P., Yau, W. Y., & Christodoulou, J. (2024). Pregnancy in women with mitochondrial disease-A literature

review and suggested guidance for preconception and pregnancy care. *The Australian & New Zealand Journal of Obstetrics & Gynaecology*. <https://doi.org/10.1111/AJO.13874>

Kendal, E. (2024). Whose (germ) line is it anyway? Reproductive technologies and kinship. *Bioethics*, 38(7), 632–642. <https://doi.org/10.1111/BIOE.13254>

Lan, X., Ao, W. L., & Li, J. (2024). Preimplantation genetic testing as a preventive strategy for the transmission of mitochondrial DNA disorders. *Systems Biology in Reproductive Medicine*, 70(1), 38–51. <https://doi.org/10.1080/19396368.2024.2306389>

Mangione, E. (2024). “Recombining” biological motherhoods. Towards two “complete” biological mothers. *Journal of Medical Ethics*. <https://doi.org/10.1136/JME-2023-109610>

Mikhailov, N., & Hämäläinen, R. H. (2024). Modulating Mitochondrial DNA Heteroplasmy with Mitochondrially Targeted Endonucleases. *Annals of Biomedical Engineering*, 52(9), 2627–2640. <https://doi.org/10.1007/S10439-022-03051-7>

Pillai, R., & Choudhary, M. (2024). Enucleated egg donation: why is it important to understand the attitudes of women on donating enucleated eggs? *Reproductive Biomedicine Online*, 49(3). <https://doi.org/10.1016/J.RBMO.2024.104323>

Yildirim, R. M., & Seli, E. (2024). Mitochondria as therapeutic targets in assisted reproduction. *Human Reproduction (Oxford, England)*, 39(10). <https://doi.org/10.1093/HUMREP/DEAE170>

Reproductive organoids

Scope: This topic covers both human and animal research looking at generating ovarian, fallopian tube, endometrial, uterine, cervical and testicular organoids to study infertility and associated treatments. The focus of this topic restricted to organoid methods, but not conventional methods which precede organoids (such as 2D cultures, organotypic cultures, or other 3D cultures). Use of organoids to model cancer are excluded. Assembloids used to study placentation and foetal-maternal interactions are in scope, whereas assembloids combining organoids or cells with blastoids are considered under the topic of SCBEM.

Abady, M. M., Saadeldin, I. M., Han, A., Bang, S., Kang, H., Seok, D. W., Kwon, H. J., Cho, J., & Jeong, J. S. (2024). Melatonin and resveratrol alleviate molecular and metabolic toxicity induced by Bisphenol A in endometrial organoids. *Reproductive Toxicology*, 128. <https://doi.org/10.1016/j.reprotox.2024.108628>

Abbas, Y., Brunel, L. G., Hollinshead, M. S., Fernando, R. C., Gardner, L., Duncan, I., Moffett, A., Best, S., Turco, M. Y., Burton, G. J., & Cameron, R. E. (2020). Generation of a three-dimensional collagen scaffold-based model of the human endometrium. *Interface Focus*, 10(2), 20190079. <https://doi.org/10.1098/rsfs.2019.0079>

Ahmad, V., Yeddula, S. G. R., Telugu, B. P., Spencer, T. E., & Kelleher, A. M. (2024). Development of polarity-reversed endometrial epithelial organoids. *Reproduction*, 167(3), 167–230. <https://doi.org/10.1530/REP-23-0478>

Alves-Lopes, J. P., Söder, O., & Stukenborg, J. B. (2017). Testicular organoid generation by a novel in vitro three-layer gradient system. *Biomaterials*, 130, 76–89. <https://doi.org/10.1016/j.biomaterials.2017.03.025>

Alves-Lopes, J. P., Söder, O., & Stukenborg, J. B. (2018). Use of a three-layer gradient system of cells for rat testicular organoid generation. *Nature Protocols*, 13(2), 248–259. <https://doi.org/10.1038/nprot.2017.140>

Alves-Lopes, J. P., & Stukenborg, J. B. (2018). Testicular organoids: A new model to study the testicular microenvironment in vitro? *Human Reproduction Update*, 24(2), 176–191. <https://doi.org/10.1093/humupd/dmx036>

Alzamil, L., Nikolakopoulou, K., & Turco, M. Y. (2021). Organoid systems to study the human female reproductive tract and pregnancy. *Cell Death and Differentiation*, 28(1), 35–51. <https://doi.org/10.1038/s41418-020-0565-5>

Baert, Y., De Kock, J., Alves-Lopes, J. P., Söder, O., Stukenborg, J. B., & Goossens, E. (2017). Primary Human Testicular Cells Self-Organize into Organoids with Testicular Properties. *Stem Cell Reports*, 8(1), 30–38. <https://doi.org/10.1016/j.stemcr.2016.11.012>

- Baert, Y., Rombaut, C., & Goossens, E. (2019). Scaffold-based and scaffold-free testicular organoids from primary human testicular cells. *Methods in Molecular Biology*, 1576, 283–290. https://doi.org/10.1007/7651_2017_48
- Baert, Y., Ruetschle, I., Cools, W., Oehme, A., Lorenz, A., Marx, U., Goossens, E., & Maschmeyer, I. (2020). A multi-organ-chip co-culture of liver and testis equivalents: A first step toward a systemic male reprotoxicity model. *Human Reproduction*, 35(5), 1029–1044. <https://doi.org/10.1093/humrep/deaa057>
- Bagheri, M. J., Valojerdi, M. R., & Salehnia, M. (2024a). Formation of ovarian organoid by co-culture of human endometrial mesenchymal stem cells and mouse oocyte in 3-dimensional culture system. *Cytotechnology*, 76(5), 571–584. <https://doi.org/10.1007/s10616-024-00639-w>
- Bagheri, M. J., Valojerdi, M. R., & Salehnia, M. (2024b). Formation of ovarian organoid by co-culture of human endometrial mesenchymal stem cells and mouse oocyte in 3-dimensional culture system. *Cytotechnology*, 76(5), 571–584. <https://doi.org/10.1007/s10616-024-00639-w>
- Banerjee, S., Xu, W., Chowdhury, I., Driss, A., Ali, M., Yang, Q., Al-Hendy, A., & Thompson, W. E. (2022). Human Myometrial and Uterine Fibroid Stem Cell-Derived Organoids for Intervening the Pathophysiology of Uterine Fibroid. *Reproductive Sciences*, 29(9), 2607–2619. <https://doi.org/10.1007/s43032-022-00960-9>
- Barton, S., Zhou, W., Santos, L. L., Menkhorst, E., Yang, G., Teh, W. T., Ang, C., Lucky, T., & Dimitriadis, E. (2023). miR-23b-3p regulates human endometrial epithelial cell adhesion implying a role in implantation. *Reproduction*, 165(4), 407–416. <https://doi.org/10.1530/REP-22-0338>
- Bishop, R. C., Boretto, M., Rutkowski, M. R., Vankelecom, H., & Derré, I. (2020). Murine Endometrial Organoids to Model Chlamydia Infection. *Frontiers in Cellular and Infection Microbiology*, 10. <https://doi.org/10.3389/fcimb.2020.00416>
- Boretto, M., Cox, B., Noben, M., Hendriks, N., Fassbender, A., Roose, H., Amant, F., Timmerman, D., Tomassetti, C., Vanhie, A., Meuleman, C., Ferrante, M., & Vankelecom, H. (2017). Development of organoids from mouse and human endometrium showing endometrial epithelium physiology and long-term expandability. *Development (Cambridge)*, 144(10), 1775–1786. <https://doi.org/10.1242/dev.148478>
- Bui, B. N., Ardisasmita, A. I., van de Vliert, F. H., Abendroth, M. S., van Hoesel, M., Mackens, S., Fuchs, S. A., Nieuwenhuis, E. E. S., Broekmans, F. J. M., & Steba, G. S. (2024). Enrichment of cell cycle pathways in progesterone-treated endometrial organoids of infertile women compared to fertile women. *Journal of Assisted Reproduction and Genetics*. <https://doi.org/10.1007/S10815-024-03173-Y>
- Bui, B. N., Boretto, M., Kobayashi, H., van Hoesel, M., Steba, G. S., van Hoogenhuijze, N., Broekmans, F. J. M., Vankelecom, H., & Torrance, H. L. (2020). Organoids can be established reliably from cryopreserved biopsy catheter-derived endometrial tissue of infertile women. *Reproductive BioMedicine Online*, 41(3), 465–473. <https://doi.org/10.1016/j.rbmo.2020.03.019>
- Burton, G. J., Jauniaux, E., Cindrova-Davies, T., & Turco, M. Y. (2024). The human gestational sac as a choriovitelline placenta during early pregnancy; the secondary yolk sac and organoid models. *Developmental Biology*, 518, 28–36. <https://doi.org/10.1016/j.ydbio.2024.11.007>
- Butt, Z., Tinning, H., O'Connell, M. J., Fenn, J., Alberio, R., & Forde, N. (2024). Understanding conceptus–maternal interactions: what tools do we need to develop? *Reproduction, Fertility and Development*, 36(1–2), 81–92. <https://doi.org/10.1071/RD23181>
- Cai, Y., Li, N., & Li, H. (2023). Combining Endometrial Assembloids and Blastoids to Delineate the Molecular Roadmap of Implantation. *Stem Cell Reviews and Reports*, 19(5), 1268–1282. <https://doi.org/10.1007/S12015-023-10527-Z>
- Cham, T. C., Chen, X., & Honaramooz, A. (2021). Current progress, challenges, and future prospects of testis organoids†. *Biology of Reproduction*, 104(5), 942–961. <https://doi.org/10.1093/biolre/ioab014>
- Cham, T. C., Ibtisham, F., Fayaz, M. A., & Honaramooz, A. (2021). Generation of a highly biomimetic organoid, including vasculature, resembling the native immature testis tissue. *Cells*, 10(7). <https://doi.org/10.3390/cells10071696>

- Chang, Y. H., Chu, T. Y., & Ding, D. C. (2020). Human fallopian tube epithelial cells exhibit stemness features, self-renewal capacity, and Wnt-related organoid formation. *Journal of Biomedical Science*, 27(1). <https://doi.org/10.1186/s12929-019-0602-1>
- Chang, Y. H., Wu, K. C., Harnod, T., & Ding, D. C. (2023). Comparison of the Cost and Effect of Combined Conditioned Medium and Conventional Medium for Fallopian Tube Organoid Cultures. *Cell Transplantation*, 32. <https://doi.org/10.1177/09636897231160216>
- Cheung, V. C., Peng, C. Y., Marinić, M., Sakabe, N. J., Aneas, I., Lynch, V. J., Ober, C., Nobrega, M. A., & Kessler, J. A. (2021). Pluripotent stem cell-derived endometrial stromal fibroblasts in a cyclic, hormone-responsive, coculture model of human decidua. *Cell Reports*, 35(7). <https://doi.org/10.1016/j.celrep.2021.109138>
- Chumduri, C., & Turco, M. Y. (2021). Organoids of the female reproductive tract. *Journal of Molecular Medicine*, 99(4), 531–553. <https://doi.org/10.1007/s00109-020-02028-0>
- Cindrova-Davies, T., Zhao, X., Elder, K., Jones, C. J. P., Moffett, A., Burton, G. J., & Turco, M. Y. (2021). Menstrual flow as a non-invasive source of endometrial organoids. *Communications Biology*, 4(1). <https://doi.org/10.1038/s42003-021-02194-y>
- Clevers, H. (2016). Modeling Development and Disease with Organoids. *Cell*, 165(7), 1586–1597. <https://doi.org/10.1016/J.CELL.2016.05.082>
- Cohen, A. B., Nikmehr, B., Abdelaal, O. A., Escott, M., Walker, S. J., Atala, A., & Sadri-Ardekani, H. (2024). MicroRNA Analysis of In Vitro Differentiation of Spermatogonial Stem Cells Using a 3D Human Testis Organoid System. *Biomedicine*, 12(8). <https://doi.org/10.3390/biomedicine12081774>
- Cortez, J., Leiva, B., Torres, C. G., Parraguez, V. H., De Los Reyes, M., Carrasco, A., & Peralta, O. A. (2022). Generation and Characterization of Bovine Testicular Organoids Derived from Primary Somatic Cell Populations. *Animals : An Open Access Journal from MDPI*, 12(17). <https://doi.org/10.3390/ani12172283>
- Cortez, J., Torres, C. G., Parraguez, V. H., De los Reyes, M., & Peralta, O. A. (2024). Bovine adipose tissue-derived mesenchymal stem cells self-assemble with testicular cells and integrates and modifies the structure of a testicular organoids. *Theriogenology*, 215, 259–271. <https://doi.org/10.1016/j.theriogenology.2023.12.013>
- Cui, Y., Zhao, H., Wu, S., & Li, X. (2020). Human Female Reproductive System Organoids: Applications in Developmental Biology, Disease Modelling, and Drug Discovery. *Stem Cell Reviews and Reports*, 16(6), 1173–1184. <https://doi.org/10.1007/s12015-020-10039-0>
- Dai, W., Liang, J., Guo, R., Zhao, Z., Na, Z., Xu, D., & Li, D. (2024). Bioengineering approaches for the endometrial research and application. *Materials Today. Bio*, 26, 101045. <https://doi.org/10.1016/j.mtbio.2024.101045>
- De Lima E Martins Lara, N., Sakib, S., & Dobrinski, I. (2021). Regulation of Cell Types within Testicular Organoids. *Endocrinology (United States)*, 162(4). <https://doi.org/10.1210/endocr/bqab033>
- De Vriendt, S., Casares, C. M., Rocha, S., & Vankelecom, H. (2023). Matrix scaffolds for endometrium-derived organoid models. *Frontiers in Endocrinology*, 14. <https://doi.org/10.3389/FENDO.2023.1240064>
- Deane, J. A., Cousins, F. L., & Gargett, C. E. (2017). Endometrial organoids: In vitro models for endometrial research and personalized medicine. *Biology of Reproduction*, 97(6), 781–783. <https://doi.org/10.1093/biolre/i0x139>
- Del Valle, J. S., Husetic, A., Diek, D., Rutgers, L. F., Asseler, J. D., Metzemaekers, J., van Mello, N. M., & Chuva de Sousa Lopes, S. M. (2024). Human Ovarian Surface Epithelium Organoids as a Platform to Study Tissue Regeneration. *Journal of Visualized Experiments : JoVE*, 210. <https://doi.org/10.3791/66797>
- Dipali, S. S., Gowett, M. Q., Kamat, P., Converse, A., Zaniker, E. J., Fennell, A., Chou, T., Pritchard, M. T., Zelinski, M., Phillip, J. M., & Duncan, F. E. (2024). Self-organizing ovarian somatic organoids preserve cellular heterogeneity and reveal cellular contributions to ovarian aging. <https://doi.org/10.1101/2024.08.10.607456>

- Dolat, L., Carpenter, V. K., Chen, Y. S., Suzuki, M., Smith, E. P., Kuddar, O., & Valdivia, R. H. (2022). Chlamydia repurposes the actin-binding protein EPS8 to disassemble epithelial tight junctions and promote infection. *Cell Host and Microbe*, 30(12), 1685-1700.e10. <https://doi.org/10.1016/j.chom.2022.10.013>
- Dolat, L., & Valdivia, R. H. (2021). An endometrial organoid model of interactions between Chlamydia and epithelial and immune cells. *Journal of Cell Science*, 134(5). <https://doi.org/10.1242/jcs.252403>
- Dong, Y., Li, J., Cao, D., Zhong, J., Liu, X., Duan, Y. G., Lee, K. F., Yeung, W. S. B., Lee, C. L., & Chiu, P. C. N. (2023). Integrated MicroRNA and Secretome Analysis of Human Endometrial Organoids Reveal the miR-3194-5p/Aquaporin/S100A9 Module in Regulating Trophoblast Functions. *Molecular and Cellular Proteomics*, 22(4). <https://doi.org/10.1016/j.mcpro.2023.100526>
- Edmonds, M. E., Forshee, M. D., & Woodruff, T. K. (2020). Extra cellular matrix-based and extra cellular matrix-free generation of murine testicular organoids. *Journal of Visualized Experiments*, 2020(164), 1–19. <https://doi.org/10.3791/61403>
- Edmonds, M. E., & Woodruff, T. K. (2020). Testicular organoid formation is a property of immature somatic cells, which self-assemble and exhibit long-term hormone-responsive endocrine function. *Biofabrication*, 12(4). <https://doi.org/10.1088/1758-5090/ab9907>
- Esfandiari, F., Favaedi, R., Heidari-Khoei, H., Chitsazian, F., Yari, S., Piryaei, A., Ghafari, F., Baharvand, H., & Shahhoseini, M. (2021). Insight into epigenetics of human endometriosis organoids: DNA methylation analysis of HOX genes and their cofactors. *Fertility and Sterility*, 115(1), 125–137. <https://doi.org/10.1016/j.fertnstert.2020.08.1398>
- Esfandiari, F., Heidari Khoei, H., Saber, M., Favaedi, R., Piryaei, A., Moini, A., Shahhoseini, M., Ramezanali, F., Ghaffari, F., & Baharvand, H. (2021). Disturbed progesterone signalling in an advanced preclinical model of endometriosis. *Reproductive BioMedicine Online*, 43(1), 139–147. <https://doi.org/10.1016/j.rbmo.2020.12.011>
- Esfandiari, F., Mansouri, N., Shahhoseini, M., Heidari Khoei, H., Mikaeeli, G., Vankelecom, H., & Baharvand, H. (2022). Endometriosis organoids: prospects and challenges. *Reproductive BioMedicine Online*, 45(1), 5–9. <https://doi.org/10.1016/j.rbmo.2022.03.016>
- Feng, L., Yang, W., Zhao, H., Bakkum-Gamez, J., Sherman, M. E., & Kannan, N. (2022). Protocol for the Detection of Organoid-Initiating Cell Activity in Patient-Derived Single Fallopian Tube Epithelial Cells. *Methods in Molecular Biology*, 2429, 445–454. https://doi.org/10.1007/978-1-0716-1979-7_30
- Filby, C. E., Wyatt, K. A., Mortlock, S., Cousins, F. L., McKinnon, B., Tyson, K. E., Montgomery, G. W., & Gargett, C. E. (2021). Comparison of Organoids from Menstrual Fluid and Hormone-Treated Endometrium: Novel Tools for Gynecological Research. *Journal of Personalized Medicine*, 11(12). <https://doi.org/10.3390/jpm11121314>
- Fitzgerald, H. C., Dhakal, P., Behura, S. K., Schust, D. J., & Spencer, T. E. (2019). Self-renewing endometrial epithelial organoids of the human uterus. *Proceedings of the National Academy of Sciences of the United States of America*, 116(46), 23132–23142. <https://doi.org/10.1073/pnas.1915389116>
- Fitzgerald, H. C., Kelleher, A. M., Ranjit, C., Schust, D. J., & Spencer, T. E. (2023). Basolateral secretions of human endometrial epithelial organoids impact stromal cell decidualization. *Molecular Human Reproduction*, 29(4). <https://doi.org/10.1093/molehr/gaad007>
- Fitzgerald, H. C., Schust, D. J., & Spencer, T. E. (2021). In vitro models of the human endometrium: Evolution and application for women's health+. *Biology of Reproduction*, 104(2), 282–293. <https://doi.org/10.1093/biolre/iaaa183>
- Francés-Herrero, E., Juárez-Barber, E., Campo, H., López-Martínez, S., de Miguel-Gómez, L., Faus, A., Pellicer, A., Ferrero, H., & Cervelló, I. (2021). Improved models of human endometrial organoids based on hydrogels from decellularized endometrium. *Journal of Personalized Medicine*, 11(6). <https://doi.org/10.3390/jpm11060504>
- Francés-Herrero, E., Lopez, R., Hellström, M., De Miguel-Gómez, L., Herraiz, S., Brännström, M., Pellicer, A., & Cervelló, I. (2022). Bioengineering trends in female reproduction: a systematic review. *Human Reproduction Update*, 28(6), 798–837. <https://doi.org/10.1093/HUMUPD/DMAC025>

- Garcia-Alonso, L., Handfield, L. F., Roberts, K., Nikolakopoulou, K., Fernando, R. C., Gardner, L., Woodhams, B., Arutyunyan, A., Polanski, K., Hoo, R., Sancho-Serra, C., Li, T., Kwakwa, K., Tuck, E., Lorenzi, V., Massalha, H., Prete, M., Kleshchevnikov, V., Tarkowska, A., ... Vento-Tormo, R. (2021). Mapping the temporal and spatial dynamics of the human endometrium in vivo and in vitro. *Nature Genetics*, 53(12), 1698–1711. <https://doi.org/10.1038/s41588-021-00972-2>
- Gebril, M., Aboelmaaty, A., Al Balah, O., Taha, T., Abbassy, A., & Elnoury, M. A. H. (2021). Bio-modulated mice epithelial endometrial organoids by low-level laser therapy serves as an invitro model for endometrial regeneration. *Reproductive Biology*, 21(4). <https://doi.org/10.1016/j.repbio.2021.100564>
- Giannakopoulos, S., Strange, D. P., Jiyarom, B., Abdelaal, O., Bradshaw, A. W., Nerurkar, V. R., Ward, M. A., Bakse, J., Yap, J., Vanapruks, S., Boisvert, W. A., Tallquist, M. D., Shikuma, C., Sadri-Ardekani, H., Clapp, P., Murphy, S. V., & Verma, S. (2023). In vitro evidence against productive SARSCoV-2 infection of human testicular cells: Bystander effects of infection mediate testicular injury. *PLoS Pathogens*, 19(5). <https://doi.org/10.1371/journal.ppat.1011409>
- Gnecco, J. S., Brown, A., Buttrey, K., Ives, C., Goods, B. A., Baugh, L., Hernandez-Gordillo, V., Loring, M., Isaacson, K. B., & Griffith, L. G. (2023). Organoid co-culture model of the human endometrium in a fully synthetic extracellular matrix enables the study of epithelial-stromal crosstalk. *Med*, 4(8), 554-579.e9. <https://doi.org/10.1016/j.medj.2023.07.004>
- Gołabek-Grenda, A., & Olejnik, A. (2022). In vitro modeling of endometriosis and endometriotic microenvironment – Challenges and recent advances. *Cellular Signalling*, 97. <https://doi.org/10.1016/j.cellsig.2022.110375>
- Goldsmith, T. M., Sakib, S., Webster, D., Carlson, D. F., Van der Hoorn, F., & Dobrinski, I. (2020). A reduction of primary cilia but not hedgehog signaling disrupts morphogenesis in testicular organoids. *Cell and Tissue Research*, 380(1), 191–200. <https://doi.org/10.1007/s00441-019-03121-8>
- Gómez-Álvarez, M., Bueno-Fernandez, C., Rodríguez-Eguren, A., Francés-Herrero, E., Agustina-Hernández, M., Faus, A., Gisbert Roca, F., Martínez-Ramos, C., Galán, A., Pellicer, A., Ferrero, H., & Cervelló, I. (2024). Hybrid Endometrial-Derived Hydrogels: Human Organoid Culture Models and In Vivo Perspectives. *Advanced Healthcare Materials*, 13(11). <https://doi.org/10.1002/adhm.202303838>
- Gu, Z. Y., Jia, S. Z., & Leng, J. H. (2020). Establishment of endometriotic models: the past and future. *Chinese Medical Journal*, 133(14), 1703–1710. <https://doi.org/10.1097/CM9.0000000000000885>
- Guo, J., Zhou, W., Sacco, M., Downing, P., Dimitriadis, E., & Zhao, F. (2023). Using organoids to investigate human endometrial receptivity. *Frontiers in Endocrinology*, 14, 1158515. <https://doi.org/10.3389/fendo.2023.1158515>
- Gurumurthy, R. K., Koster, S., Kumar, N., Meyer, T. F., & Chumduri, C. (2022). Patient-derived and mouse endo-ectocervical organoid generation, genetic manipulation and applications to model infection. *Nature Protocols*, 17(7), 1658–1690. <https://doi.org/10.1038/s41596-022-00695-6>
- Haider, S., & Beristain, A. G. (2023). Human organoid systems in modeling reproductive tissue development, function, and disease. *Human Reproduction*, 38(8), 1449–1463. <https://doi.org/10.1093/humrep/dead085>
- Haider, S., Gamperl, M., Burkard, T. R., Kunihs, V., Kaindl, U., Junttila, S., Fiala, C., Schmidt, K., Mendjan, S., Knöfler, M., & Latos, P. A. (2019). Estrogen Signaling Drives Ciliogenesis in Human Endometrial Organoids. *Endocrinology*, 160(10), 2282–2297. <https://doi.org/10.1210/en.2019-00314>
- Heidari Khoei, H., Javali, A., Kagawa, H., Sommer, T. M., Sestini, G., David, L., Slovakova, J., Novatchkova, M., Scholte op Reimer, Y., & Rivron, N. (2023). Generating human blastoids modeling blastocyst-stage embryos and implantation. *Nature Protocols*, 18(5), 1584–1620. <https://doi.org/10.1038/s41596-023-00802-1>
- Heidari-Khoei, H., Esfandiari, F., Hajari, M. A., Ghorbaninejad, Z., Piryaei, A., Piryaei, A., & Baharvand, H. (2020). Organoid technology in female reproductive biomedicine. *Reproductive Biology and Endocrinology*, 18(1). <https://doi.org/10.1186/s12958-020-00621-z>
- Heidari-Khoei, H., Esfandiari, F., Moini, A., Yari, S., Saber, M., Ghaffari Novin, M., Piryaei, A., & Baharvand, H. (2022). Derivation of hormone-responsive human endometrial organoids and stromal cells

from cryopreserved biopsies. *Experimental Cell Research*, 417(1).
<https://doi.org/10.1016/J.YEXCR.2022.113205>

Hemberger, M., & Dean, W. (2023). The placenta: epigenetic insights into trophoblast developmental models of a generation-bridging organ with long-lasting impact on lifelong health. *Physiological Reviews*, 103(4), 2523–2560. <https://doi.org/10.1152/physrev.00001.2023>

Hennes, A., Held, K., Boretto, M., De Clercq, K., Van den Eynde, C., Vanhie, A., Van Ranst, N., Benoit, M., Luyten, C., Peeraer, K., Tomassetti, C., Meuleman, C., Voets, T., Vankelecom, H., & Vriens, J. (2019). Functional expression of the mechanosensitive PIEZO1 channel in primary endometrial epithelial cells and endometrial organoids. *Scientific Reports*, 9(1). <https://doi.org/10.1038/s41598-018-38376-8>

Hewitt, S. C., Dickson, M. J., Edwards, N., Hampton, K., Garantziotis, S., & DeMayo, F. J. (2023). From cup to dish: how to make and use endometrial organoid and stromal cultures derived from menstrual fluid. *Frontiers in Endocrinology*, 14. <https://doi.org/10.3389/FENDO.2023.1220622>

Hewitt, S. C., Wu, S. P., Wang, T., Ray, M., Brolinson, M., Young, S. L., Spencer, T. E., Decherney, A., & Demayo, F. J. (2022). The Estrogen Receptor α Cistrome in Human Endometrium and Epithelial Organoids. *Endocrinology (United States)*, 163(9). <https://doi.org/10.1210/endo/bqac116>

Hewitt, S. C., Wu, S. P., Wang, T., Young, S. L., Spencer, T. E., & Demayo, F. J. (2022). Progesterone Signaling in Endometrial Epithelial Organoids. *Cells*, 11(11). <https://doi.org/10.3390/cells11111760>

Hu, R., Wang, Y., Li, W., Liu, H., Wu, R., Xu, X., Jiang, X., Xing, Q., Wang, J., & Wei, Z. (2024). Transplantation of human umbilical cord blood mononuclear cells promotes functional endometrium reconstruction via downregulating EMT in damaged endometrium. *Regenerative Therapy*, 27, 279–289. <https://doi.org/10.1016/J.RETH.2024.03.030>

Hwang, S. Y., Lee, D., Lee, G., Ahn, J., Lee, Y. G., Koo, H. S., & Kang, Y. J. (2024). Endometrial organoids: a reservoir of functional mitochondria for uterine repair. *Theranostics*, 14(3), 954–972. <https://doi.org/10.7150/thno.90538>

Jamaluddin, M. F. B., Ghosh, A., Ingle, A., Mohammed, R., Ali, A., Bahrami, M., Kaiko, G., Gibb, Z., Filipe, E. C., Cox, T. R., Boulton, A., O'Sullivan, R., Ius, Y., Karakoti, A., Vinu, A., Nahar, P., Jaaback, K., Bansal, V., & Tanwar, P. S. (2022). Bovine and human endometrium-derived hydrogels support organoid culture from healthy and cancerous tissues. *Proceedings of the National Academy of Sciences of the United States of America*, 119(44). <https://doi.org/10.1073/pnas.2208040119>

Jiang, R., Tang, X., Pan, J., Li, G., Yang, N., Tang, Y., Bi, S., Cai, H., Chen, Q., Chen, D., Wang, H., & Kong, S. (2022). CDC42 governs normal oviduct multiciliogenesis through activating AKT to ensure timely embryo transport. *Cell Death and Disease*, 13(9). <https://doi.org/10.1038/s41419-022-05184-y>

Jiang, Y., Palomares, A. R., Munoz, P., Nalvarte, I., Acharya, G., Inzunza, J., Varshney, M., & Rodriguez-Wallberg, K. A. (2024). Proof-of-Concept for Long-Term Human Endometrial Epithelial Organoids in Modeling Menstrual Cycle Responses. *Cells*, 13(21). <https://doi.org/10.3390/cells13211811>

Johnson, M. H. (2017). First evidence that endometrial-like organoids can develop from the endometrial mesenchymal stem/stromal cell population. *Reproductive BioMedicine Online*, 35(3), 239–240. <https://doi.org/10.1016/j.rbmo.2017.07.009>

Jones, A. S. K., & Shikanov, A. (2019). Follicle development as an orchestrated signaling network in a 3D organoid. *Journal of Biological Engineering*, 13(1), 2. <https://doi.org/10.1186/s13036-018-0134-3>

Jorban, A., Lunenfeld, E., & Huleihel, M. (2024). Effect of Temperature on the Development of Stages of Spermatogenesis and the Functionality of Sertoli Cells In Vitro. *International Journal of Molecular Sciences*, 25(4). <https://doi.org/10.3390/ijms25042160>

Juárez-barber, E., Francés-herrero, E., Corachán, A., Vidal, C., Giles, J., Alamá, P., Faus, A., Pellicer, A., Cervelló, I., & Ferrero, H. (2022). Establishment of Adenomyosis Organoids as a Preclinical Model to Study Infertility. *Journal of Personalized Medicine*, 12(2). <https://doi.org/10.3390/jpm12020219>

Juárez-Barber, E., Segura-Benítez, M., Carbajo-García, M. C., Bas-Rivas, A., Faus, A., Vidal, C., Giles, J., Labarta, E., Pellicer, A., Cervelló, I., & Ferrero, H. (2023). Extracellular vesicles secreted by adenomyosis

- endometrial organoids contain miRNAs involved in embryo implantation and pregnancy. *Reproductive Biomedicine Online*, 46(3), 470–481. <https://doi.org/10.1016/J.RBMO.2022.12.008>
- Kagawa, H., Javali, A., Khoei, H. H., Sommer, T. M., Sestini, G., Novatchkova, M., Scholte op Reimer, Y., Castel, G., Bruneau, A., Maenhoudt, N., Lammers, J., Loubersac, S., Freour, T., Vankelecom, H., David, L., & Rivron, N. (2022). Human blastoids model blastocyst development and implantation. *Nature*, 601(7894), 600–605. <https://doi.org/10.1038/S41586-021-04267-8>
- Kanatsu-Shinohara, M., Ogonuki, N., Matoba, S., Morimoto, H., Shiromoto, Y., Ogura, A., & Shinohara, T. (2022). Regeneration of spermatogenesis by mouse germ cell transplantation into allogeneic and xenogeneic testis primordia or organoids. *Stem Cell Reports*, 17(4), 924–935. <https://doi.org/10.1016/j.stemcr.2022.02.013>
- Kanbar, M., Vermeulen, M., & Wyns, C. (2021). Organoids as tools to investigate the molecular mechanisms of male infertility and its treatments. *Reproduction*, 161(5), R103–R112. <https://doi.org/10.1530/REP-20-0499>
- Kanton, S., & Paş Ca, S. P. (2022). Human assembloids. <https://doi.org/10.1242/dev.201120>
- Kawamura, K., Matsumura, Y., Kawamura, T., Araki, H., Hamada, N., Kuramoto, K., Yagi, H., Onoyama, I., Asanoma, K., & Kato, K. (2024). Endometrial senescence is mediated by interleukin 17 receptor B signaling. *Cell Communication and Signaling*, 22(1). <https://doi.org/10.1186/s12964-024-01740-5>
- Kaya, Y. A., de Zoete, M. R., & Steba, G. S. (2024). Advanced Technologies for Studying Microbiome–Female Reproductive Tract Interactions: Organoids, Organoids-on-a-Chip, and Beyond. *Seminars in Reproductive Medicine*, 41(5), 160–171. <https://doi.org/10.1055/s-0043-1778067>
- Kessler, M., Hoffmann, K., Brinkmann, V., Thieck, O., Jackisch, S., Toelle, B., Berger, H., Mollenkopf, H. J., Mangler, M., Sehoul, J., Fotopoulou, C., & Meyer, T. F. (2015). The Notch and Wnt pathways regulate stemness and differentiation in human fallopian tube organoids. *Nature Communications*, 6. <https://doi.org/10.1038/ncomms9989>
- Kiani, M., Movahedin, M., Halvaei, I., & Soleimani, M. (2021). Formation of organoid-like structures in the decellularized rat testis. *Iranian Journal of Basic Medical Sciences*, 24(11), 1523–1528. <https://doi.org/10.22038/IJBMS.2021.58294.12948>
- Kleinová, M., Varga, I., Čeháková, M., Valent, M., & Klein, M. (2024). Exploring the black box of human reproduction: endometrial organoids and assembloids - generation, implantation modeling, and future clinical perspectives. *Frontiers in Cell and Developmental Biology*, 12, 1482054. <https://doi.org/10.3389/fcell.2024.1482054>
- Knarston, I. M., Pachernegg, S., Robevska, G., Ghobrial, I., Er, P. X., Georges, E., Takasato, M., Combes, A. N., Jørgensen, A., Little, M. H., Sinclair, A. H., & Ayers, K. L. (2020). An In Vitro Differentiation Protocol for Human Embryonic Bipotential Gonad and Testis Cell Development. *Stem Cell Reports*, 15(6), 1377–1391. <https://doi.org/10.1016/j.stemcr.2020.10.009>
- Koster, S., Gurumurthy, R. K., Kumar, N., Prakash, P. G., Dhanraj, J., Bayer, S., Berger, H., Kurian, S. M., Drabkina, M., Mollenkopf, H. J., Goosmann, C., Brinkmann, V., Nagel, Z., Mangler, M., Meyer, T. F., & Chumduri, C. (2022). Modelling Chlamydia and HPV co-infection in patient-derived ectocervix organoids reveals distinct cellular reprogramming. *Nature Communications*, 13(1). <https://doi.org/10.1038/s41467-022-28569-1>
- Kriseman, M. L., Tang, S., Liao, Z., Jiang, P., Parks, S. E., Cope, D. I., Yuan, F., Chen, F., Masand, R. P., Castro, P. D., Ittmann, M. M., Creighton, C. J., Tan, Z., & Monsivais, D. (2023). SMAD2/3 signaling in the uterine epithelium controls endometrial cell homeostasis and regeneration. *Communications Biology*, 6(1). <https://doi.org/10.1038/s42003-023-04619-2>
- Lawson, E. F., Ghosh, A., Blanch, V., Grupen, C. G., Aitken, R. J., Lim, R., Drury, H. R., Baker, M. A., Gibb, Z., & Tanwar, P. S. (2023). Establishment and characterization of oviductal organoids from farm and companion animals†. *Biology of Reproduction*, 108(6), 854–865. <https://doi.org/10.1093/biolre/ioad030>
- Lawson, E. F., Ghosh, A., Grupen, C., Netherton, J., Aitken, R. J., Smith, N. D., Lim, R., Drury, H. R., Pickford, R., Gibb, Z., Baker, M., Tanwar, P. S., & Swegen, A. (2024). Investigations into the role of

- platelet-activating factor in the peri-conception period of the mare. *Reproduction* (Cambridge, England), 168(4). <https://doi.org/10.1530/REP-24-0049>
- Li, N., Du, X., Zhao, Y., Zeng, Q., Han, C., Xiong, D., He, L., Zhang, G., & Liu, W. (2024). Exploring stem cell technology: Pioneering new pathways for female fertility preservation and restoration. *Reproductive Biology*, 24(4), 100958. <https://doi.org/10.1016/j.repbio.2024.100958>
- Li, X., Kodithuwakku, S. P., Chan, R. W. S., Yeung, W. S. B., Yao, Y., Ng, E. H. Y., Chiu, P. C. N., & Lee, C. L. (2022). Three-dimensional culture models of human endometrium for studying trophoblast-endometrium interaction during implantation. *Reproductive Biology and Endocrinology*, 20(1). <https://doi.org/10.1186/s12958-022-00973-8>
- Li, X., Zheng, M., Xu, B., Li, D., Shen, Y., Nie, Y., Ma, L., & Wu, J. (2021). Generation of offspring-producing 3D ovarian organoids derived from female germline stem cells and their application in toxicological detection. *Biomaterials*, 279. <https://doi.org/10.1016/j.biomaterials.2021.121213>
- Liao, Z., Tang, S., Jiang, P., Geng, T., Cope, D. I., Dunn, T. N., Guner, J., Radilla, L. A., Guan, X., & Monsivais, D. (2024). Impaired bone morphogenetic protein (BMP) signaling pathways disrupt decidualization in endometriosis. *Communications Biology*, 7(1). <https://doi.org/10.1038/S42003-024-05898-Z>
- Lin, L., Bai, K., Li, J., Chiu, P. C. N., & Lee, C. L. (2023). Regulatory role of human endometrial gland secretome on macrophage differentiation. *Journal of Reproductive Immunology*, 160. <https://doi.org/10.1016/j.jri.2023.104158>
- Lin, Y. X., Wei, Y. Z., Jiang, M. Z., Tang, X., Huang, F., & Yang, X. Z. (2021). Organoid culture of mouse fallopian tube epithelial stem cells with a thermo-reversible gelation polymer. *Tissue and Cell*, 73. <https://doi.org/10.1016/j.tice.2021.101622>
- Liu, Y., Sun, Y., & Cheng, S. (2024). Advances in the use of organoids in endometrial diseases. *International Journal of Gynecology and Obstetrics*, 166(2), 502–511. <https://doi.org/10.1002/ijgo.15422>
- Löhmussaar, K., Oka, R., Espejo Valle-Inclan, J., Smits, M. H. H., Wardak, H., Korving, J., Begthel, H., Proost, N., van de Ven, M., Kranenburg, O. W., Jonges, T. G. N., Zweemer, R. P., Veersema, S., van Boxtel, R., & Clevers, H. (2021). Patient-derived organoids model cervical tissue dynamics and viral oncogenesis in cervical cancer. *Cell Stem Cell*, 28(8), 1380-1396.e6. <https://doi.org/10.1016/j.stem.2021.03.012>
- Lopez, I., & Truskey, G. A. (2024). Multi-cellular engineered living systems to assess reproductive toxicology. *Reproductive Toxicology* (Elmsford, N.Y.), 127. <https://doi.org/10.1016/J.REPROTOX.2024.108609>
- Lu, M., Han, Y., Zhang, Y., Yu, R., Su, Y., Chen, X., Liu, B., Li, T., Zhao, R., & Zhao, H. (2024). Investigating Aging-Related Endometrial Dysfunction Using Endometrial Organoids. *Cell Proliferation*. <https://doi.org/10.1111/CPR.13780>
- Luddi, A., Pavone, V., Governini, L., Capaldo, A., Landi, C., Ietta, F., Paccagnini, E., Morgante, G., De Leo, V., & Piomboni, P. (2021). Emerging role of embryo secretome in the paracrine communication at the implantation site: a proof of concept. *Fertility and Sterility*, 115(4), 1054–1062. <https://doi.org/10.1016/j.fertnstert.2020.10.058>
- Luddi, A., Pavone, V., Semplici, B., Governini, L., Criscuoli, M., Paccagnini, E., Gentile, M., Morgante, G., Leo, V. De, Belmonte, G., Zarovni, N., & Piomboni, P. (2020). Organoids of Human Endometrium: A Powerful In Vitro Model for the Endometrium-Embryo Cross-Talk at the Implantation Site. *Cells*, 9(5). <https://doi.org/10.3390/cells9051121>
- Luo, H., Li, X., Tian, G. G., Li, D., Hou, C., Ding, X., Hou, L., Lyu, Q., Yang, Y., Cooney, A. J., Xie, W., Xiong, J., Wang, H., Zhao, X., & Wu, J. (2021). Offspring production of ovarian organoids derived from spermatogonial stem cells by defined factors with chromatin reorganization. *Journal of Advanced Research*, 33, 81–98. <https://doi.org/10.1016/j.jare.2021.03.006>
- Maenhoudt, N., De Moor, A., & Vankelecom, H. (2022). Modeling Endometrium Biology and Disease. *Journal of Personalized Medicine*, 12(7). <https://doi.org/10.3390/jpm12071048>

- Mall, E. M., Rotte, N., Yoon, J., Sandhowe-Klaverkamp, R., Röpke, A., Wistuba, J., Hübner, K., Schöler, H. R., & Schlatt, S. (2020). A novel xeno-organoid approach: exploring the crosstalk between human iPSC-derived PGC-like and rat testicular cells. *Molecular Human Reproduction*, 26(12), 879–893. <https://doi.org/10.1093/molehr/gaaa067>
- Marinić, M., Rana, S., & Lynch, V. J. (2020). Derivation of endometrial gland organoids from term placenta. *Placenta*, 101, 75–79. <https://doi.org/10.1016/j.placenta.2020.08.017>
- Maru, Y., Tanaka, N., Tatsumi, Y., Nakamura, Y., Itami, M., & Hippo, Y. (2021). Kras activation in endometrial organoids drives cellular transformation and epithelial-mesenchymal transition. *Oncogenesis*, 10(6), 46. <https://doi.org/10.1038/s41389-021-00337-8>
- Mecca, R., Tang, S., Jones, C., & Coward, K. (2024). The limitations of testicular organoids: are they truly as promising as we believe? *Reproduction, Fertility and Development*, 36(11). <https://doi.org/10.1071/RD23216>
- Menjivar, N. G., Gad, A., Thompson, R. E., Meyers, M. A., Hollinshead, F. K., & Tesfaye, D. (2023). Bovine oviductal organoids: a multi-omics approach to capture the cellular and extracellular molecular response of the oviduct to heat stress. *BMC Genomics*, 24(1). <https://doi.org/10.1186/s12864-023-09746-y>
- Mu, C., Li, X., Yang, J., Tian, G. G., Bai, H., Lin, W., Wang, L., & Wu, J. (2024). Spatial Transcriptome and Single Nucleus Transcriptome Sequencing Reveals Tetrahydroxy Stilbene Glucoside Promotes Ovarian Organoids Development Through the Vegfa-Ephb2 Pair. *Advanced Science (Weinheim, Baden-Württemberg, Germany)*, e2410098. <https://doi.org/10.1002/advs.202410098>
- Murphy, A. R., Wiwatpanit, T., Lu, Z., Davaadelger, B., & Kim, J. J. (2019). Generation of multicellular human primary endometrial organoids. *Journal of Visualized Experiments*, 2019(152). <https://doi.org/10.3791/60384>
- Nason-Tomaszewski, C. E., Thomas, E. E., Matera, D. L., Baker, B. M., & Shikanov, A. (2024). Extracellular matrix-templating fibrous hydrogels promote ovarian tissue remodeling and oocyte growth. *Bioactive Materials*, 32, 292–303. <https://doi.org/10.1016/j.bioactmat.2023.10.001>
- Nengzhuang, W., Jiaming, S., Minghua, L. I. U., Long, M., Lina, Q. I. N., Xuemei, G. E., & Hongli, Y. A. N. (2022a). A brief history of testicular organoids: from theory to the wards. *Journal of Assisted Reproduction and Genetics*, 39(7), 1423–1431. <https://doi.org/10.1007/s10815-022-02529-6>
- Nengzhuang, W., Jiaming, S., Minghua, L. I. U., Long, M., Lina, Q. I. N., Xuemei, G. E., & Hongli, Y. A. N. (2022b). A brief history of testicular organoids: from theory to the wards. *Journal of Assisted Reproduction and Genetics*, 39(7), 1423–1431. <https://doi.org/10.1007/S10815-022-02529-6>
- Nikmahzar, A., Khadivi, F., Koruji, M., Jahanshahi, M., Dehghan Tarazjani, M., Shabani, M., Abbasi, Y., & Abbasi, M. (2023). Evaluation of Apoptosis-related Genes and Hormone Secretion Profiles Using Three Dimensional Culture System of Human Testicular Organoids. *Galen Medical Journal*, 12, 1–13. <https://doi.org/10.31661/gmj.v12i0.2805>
- Nikmahzar, A., Koruji, M., Jahanshahi, M., Khadivi, F., Shabani, M., Dehghani, S., Forouzes, M., Jabari, A., Feizollahi, N., Salem, M., Ghanami Gashti, N., Abbasi, Y., & Abbasi, M. (2023). Differentiation of human primary testicular cells in the presence of SCF using the organoid culture system. *Artificial Organs*, 47(12), 1818–1830. <https://doi.org/10.1111/aor.14643>
- Nikolakopoulou, K., & Turco, M. Y. (2021). Investigation of infertility using endometrial organoids. *Reproduction*, 161(5), R113–R127. <https://doi.org/10.1530/REP-20-0428>
- Oliver, E., Alves-Lopes, J. P., Hartevelde, F., Mitchell, R. T., Åkesson, E., Söder, O., & Stukenborg, J. B. (2021a). Self-organising human gonads generated by a Matrigel-based gradient system. *BMC Biology*, 19(1). <https://doi.org/10.1186/s12915-021-01149-3>
- Oliver, E., Alves-Lopes, J. P., Hartevelde, F., Mitchell, R. T., Åkesson, E., Söder, O., & Stukenborg, J. B. (2021b). Self-organising human gonads generated by a Matrigel-based gradient system. *BMC Biology*, 19(1). <https://doi.org/10.1186/s12915-021-01149-3>

- Pendergraft, S. S., Sadri-Ardekani, H., Atala, A., & Bishop, C. E. (2017). Three-dimensional testicular organoid: A novel tool for the study of human spermatogenesis and gonadotoxicity in vitro. *Biology of Reproduction*, 96(3), 720–732. <https://doi.org/10.1095/biolreprod.116.143446>
- Peng, T. L., Yang, S., Lian, W., Liu, X., Zheng, P., Qin, X., Liao, B., Zhou, P., Wang, Y., Liu, F., Yang, Z., Ye, Z., Shan, H., Liu, X., Yu, Y., & Li, R. (2024). Cytoskeletal and inter-cellular junction remodelling in endometrial organoids under oxygen–glucose deprivation: a new potential pathological mechanism for thin endometria. *Human Reproduction*, 39(8), 1778–1793. <https://doi.org/10.1093/humrep/deae137>
- Pryzhkova, M. V., Boers, R., & Jordan, P. W. (2022). Modeling Human Gonad Development in Organoids. *Tissue Engineering and Regenerative Medicine*, 19(6), 1185–1206. <https://doi.org/10.1007/s13770-022-00492-y>
- Qin, X., Cao, M., Peng, T., Shan, H., Lian, W., Yu, Y., Shui, G., & Li, R. (2024). Features, Potential Invasion Pathways, and Reproductive Health Risks of Microplastics Detected in Human Uterus. *Environmental Science and Technology*, 58(24), 10482–10493. <https://doi.org/10.1021/acs.est.4c01541>
- Qin, X., Hu, K.-L., Li, Q., Sun, Y., Peng, T., Liu, X., Li, J., Nan, W., Yu, Y., Qi, X., & Li, R. (2024). In Situ Sprayed Hydrogel Delivers Extracellular Vesicles Derived from Human Endometrial Organoids for Uterine Function Preservation and Fertility Restoration. *Advanced Healthcare Materials*, e2403604. <https://doi.org/10.1002/adhm.202403604>
- Rawlings, T. M., Makwana, K., Taylor, D. M., Molè, M. A., Fishwick, K. J., Tryfonos, M., Odendaal, J., Hawkes, A., Zernicka-Goetz, M., Hartshorne, G. M., Brosens, J. J., & Lucas, E. S. (2021). Modelling the impact of decidual senescence on embryo implantation in human endometrial assembloids. *ELife*, 10. <https://doi.org/10.7554/eLife.69603>
- Rawlings, T. M., Makwana, K., Tryfonos, M., & Lucas, E. S. (2021). Organoids to model the endometrium: implantation and beyond. *Reproduction and Fertility*, 2(3), R85–R101. <https://doi.org/10.1530/RAF-21-0023>
- Rawlings, T. M., Tryfonos, M., Makwana, K., Taylor, D. M., Brosens, J. J., & Lucas, E. S. (2024). Endometrial Assembloids to Model Human Embryo Implantation In Vitro. *Methods in Molecular Biology (Clifton, N.J.)*, 2767, 63–74. https://doi.org/10.1007/978-1-0716-3698-5_11
- Rezaei Topraggaleh, T., Rezazadeh Valojerdi, M., Montazeri, L., & Baharvand, H. (2019). A testis-derived macroporous 3D scaffold as a platform for the generation of mouse testicular organoids. *Biomaterials Science*, 7(4), 1422–1436. <https://doi.org/10.1039/c8bm01001c>
- Richer, G., Baert, Y., & Goossens, E. (2020). In-vitro spermatogenesis through testis modelling: Toward the generation of testicular organoids. *Andrology*, 8(4), 879–891. <https://doi.org/10.1111/andr.12741>
- Richer, G., Goyvaerts, C., Marchandise, L., Vanhaecke, T., Goossens, E., & Baert, Y. (2024). Spermatogenesis in mouse testicular organoids with testis-specific architecture, improved germ cell survival and testosterone production. *Biofabrication*, 16(4). <https://doi.org/10.1088/1758-5090/ad618f>
- Richer, G., Hobbs, R. M., Loveland, K. L., Goossens, E., & Baert, Y. (2021). Long-Term Maintenance and Meiotic Entry of Early Germ Cells in Murine Testicular Organoids Functionalized by 3D Printed Scaffolds and Air-Medium Interface Cultivation. *Frontiers in Physiology*, 12, 757565. <https://doi.org/10.3389/fphys.2021.757565>
- Richer, G., Vanhaecke, T., Rogiers, V., Goossens, E., & Baert, Y. (2024). Mouse In Vitro Spermatogenesis on 3D Bioprinted Scaffolds. *Methods in Molecular Biology*, 2770, 135–149. https://doi.org/10.1007/978-1-0716-3698-5_11
- Rizo, J. A., Davenport, K. M., Winuthayanon, W., Spencer, T. E., & Kelleher, A. M. (2023). Estrogen receptor alpha regulates uterine epithelial lineage specification and homeostasis. *IScience*, 26(9), 107568. <https://doi.org/10.1016/j.isci.2023.107568>
- Rizo, J. A., Spencer, T. E., & Kelleher, A. M. (2024). Protocol for the establishment and characterization of an endometrial-derived epithelial organoid and stromal cell co-culture system. *STAR Protocols*, 5(1). <https://doi.org/10.1016/j.xpro.2024.102894>

- Robinson, M., Bedford, E., Witherspoon, L., Willerth, S. M., & Flannigan, R. (2022). Using clinically derived human tissue to 3-dimensionally bioprint personalized testicular tubules for in vitro culturing: first report. *F and S Science*, 3(2), 130–139. <https://doi.org/10.1016/j.xfss.2022.02.004>
- Rose, I. M., Bidarimath, M., Webster, A., Godwin, A. K., Flesken-Nikitin, A., & Nikitin, A. Y. (2020). WNT and inflammatory signaling distinguish human Fallopian tube epithelial cell populations. *Scientific Reports*, 10(1). <https://doi.org/10.1038/s41598-020-66556-y>
- Saadeldin, I. M., Ehab, S., Noreldin, A. E., Swelum, A. A. A., Bang, S., Kim, H., Yoon, K. Y., Lee, S., & Cho, J. (2024). Current strategies using 3D organoids to establish in vitro maternal-embryonic interaction. *Journal of Veterinary Science*, 25(3). <https://doi.org/10.4142/JVS.24004>
- Saadeldin, I. M., Han, A., Bang, S., Kang, H., Kim, H., Abady, M. M., Jeong, J. S., Kwon, H. J., Lee, S., & Cho, J. (2024). Generation of porcine endometrial organoids and their use as a model for enhancing embryonic attachment and elongation. *Reproduction*, 167(2). <https://doi.org/10.1530/REP-23-0429>
- Sadeghi, E., Rezazadeh Valojerdi, M., & Salehnia, M. (2023). Co-Culture of Mouse Blastocysts on A Human Recellularized Endometrial Scaffold: An In Vitro Model for Future Implantation Studies. *Cell Journal*, 25(8), 579–590. <https://doi.org/10.22074/cellj.2023.1989926.1236>
- Sakib, S., Goldsmith, T., Voigt, A., & Dobrinski, I. (2020). Testicular organoids to study cell–cell interactions in the mammalian testis. *Andrology*, 8(4), 835–841. <https://doi.org/10.1111/andr.12680>
- Sakib, S., Lara, N. de L. E. M., Huynh, B. C., & Dobrinski, I. (2022). Organotypic Rat Testicular Organoids for the Study of Testicular Maturation and Toxicology. *Frontiers in Endocrinology*, 13, 892342. <https://doi.org/10.3389/fendo.2022.892342>
- Sakib, S., Uchida, A., Valenzuela-Leon, P., Yu, Y., Valli-Pulaski, H., Orwig, K., Ungrin, M., & Dobrinski, I. (2019). Formation of organotypic testicular organoids in microwell culture. *Biology of Reproduction*, 100(6), 1648–1660. <https://doi.org/10.1093/biolre/ioz053>
- Sakib, S., Voigt, A., Goldsmith, T., & Dobrinski, I. (2019). Three-dimensional testicular organoids as novel in vitro models of testicular biology and toxicology. *Environmental Epigenetics*, 5(3), dvz011. <https://doi.org/10.1093/eep/dvz011>
- Sakib, S., Yu, Y., Voigt, A., Ungrin, M., & Dobrinski, I. (2019). Generation of porcine testicular organoids with testis specific architecture using microwell culture. *Journal of Visualized Experiments*, 2019(152). <https://doi.org/10.3791/60387>
- Salisbury, E., Rawlings, T. M., Efstathiou, S., Tryfonos, M., Makwana, K., Fitzgerald, H. C., Gargett, C. E., Cameron, N. R., Haddleton, D. M., Brosens, J. J., & Eissa, A. M. (2024). Photo-Cross-linked Gelatin Methacryloyl Hydrogels Enable the Growth of Primary Human Endometrial Stromal Cells and Epithelial Gland Organoids. *ACS Applied Materials and Interfaces*, 16(30), 39140–39152. <https://doi.org/10.1021/acsami.4c08763>
- Santativongchai, P., Klæui, C. C., Kosonsiriluk, S., Saqui-Salces, M., Reed, K. M., Wileman, B. W., Studniski, M. M., & Boukherroub, K. S. (2024). Protocol to establish turkey oviductal organoids as an in vitro model. *STAR Protocols*, 5(4). <https://doi.org/10.1016/j.xpro.2024.103384>
- Schutgens, F., & Clevers, H. (2020). Human Organoids: Tools for Understanding Biology and Treating Diseases. *Annual Review of Pathology: Mechanisms of Disease*, 15(Volume 15, 2020), 211–234. <https://doi.org/10.1146/ANNUREV-PATHMECHDIS-012419-032611/CITE/REFWORKS>
- Shibata, S., Endo, S., Nagai, L. A. E., Kobayashi, E. H., Oike, A., Kobayashi, N., Kitamura, A., Hori, T., Nashimoto, Y., Nakato, R., Hamada, H., Kaji, H., Kikutake, C., Suyama, M., Saito, M., Yaegashi, N., Okae, H., & Arima, T. (2024). Modeling embryo-endometrial interface recapitulating human embryo implantation. *Science Advances*, 10(8). <https://doi.org/10.1126/sciadv.adi4819>
- Shibata, S., Kobayashi, E. H., Kobayashi, N., Oike, A., Okae, H., & Arima, T. (2020). Unique features and emerging in vitro models of human placental development. *Reproductive Medicine and Biology*, 19(4), 301–313. <https://doi.org/10.1002/rmb2.12347>
- Simintiras, C. A., Dhakal, P., Ranjit, C., Fitzgerald, H. C., Balboula, A. Z., & Spencer, T. E. (2021). Capture and metabolomic analysis of the human endometrial epithelial organoid secretome. *Proceedings of the*

National Academy of Sciences of the United States of America, 118(15).
<https://doi.org/10.1073/pnas.2026804118>

Sistani, M. N., Zavareh, S., Valojerdi, M. R., & Salehnia, M. (2024). Reconstruction of ovarian follicular-like structure by recellularization of a cell-free human ovarian scaffold with mouse fetal ovarian cells. *Cytotechnology*, 76(1), 27–38. <https://doi.org/10.1007/s10616-023-00595-x>

Sittadjody, S., Criswell, T., Jackson, J. D., Atala, A., & Yoo, J. J. (2021). Regenerative Medicine Approaches in Bioengineering Female Reproductive Tissues. *Reproductive Sciences*, 28(6), 1573–1595. <https://doi.org/10.1007/s43032-021-00548-9>

Skardal, A., Aleman, J., Forsythe, S., Rajan, S., Murphy, S., Devarasetty, M., Pourhabibi Zarandi, N., Nzou, G., Wicks, R., Sadri-Ardekani, H., Bishop, C., Soker, S., Hall, A., Shupe, T., & Atala, A. (2020). Drug compound screening in single and integrated multi-organoid body-on-a-chip systems. *Biofabrication*, 12(2). <https://doi.org/10.1088/1758-5090/ab6d36>

Smela, M. D. P., Kramme, C. C., Fortuna, P. R. J., Adams, J. L., Su, R., Dong, E., Kobayashi, M., Brix, G., Kavirayuni, V. S., Tysinger, E., Kohman, R. E., Shioda, T., Chatterjee, P., & Church, G. M. (2023). Directed differentiation of human iPSCs to functional ovarian granulosa-like cells via transcription factor overexpression. *ELife*, 12. <https://doi.org/10.7554/eLife.83291>

Stopel, A., Lev, C., Dahari, S., Adibi, O., Armon, L., & Gonen, N. (2024). Towards a “Testis in a Dish”: Generation of Mouse Testicular Organoids that Recapitulate Testis Structure and Expression Profiles. *International Journal of Biological Sciences*, 20(3), 1024–1041. <https://doi.org/10.7150/ijbs.89480>

Strange, D. P., Jiyarom, B., Zarandi, N. P., Xie, X., Baker, C., Sadri-Ardekani, H., Shi, P. Y., & Verma, S. (2019). Axl promotes Zika virus entry and modulates the antiviral state of human Sertoli cells. *MBio*, 10(4). <https://doi.org/10.1128/mBio.01372-19>

Strange, D. P., Zarandi, N. P., Trivedi, G., Atala, A., Bishop, C. E., Sadri-Ardekani, H., & Verma, S. (2018). Human testicular organoid system as a novel tool to study Zika virus pathogenesis correspondence. *Emerging Microbes and Infections*, 7(1). <https://doi.org/10.1038/s41426-018-0080-7>

Stratopoulou, C. A., Rossi, M., Beaussart, C., Zipponi, M., Camboni, A., Donnez, J., & Dolmans, M.-M. (2024). Generation of epithelial-stromal assembloids as an advanced in vitro model of impaired adenomyosis-related endometrial receptivity. *Fertility and Sterility*. <https://doi.org/10.1016/j.fertnstert.2024.08.339>

Tan, Y., Flynn, W. F., Sivajothi, S., Luo, D., Bozal, S. B., Davé, M., Luciano, A. A., Robson, P., Luciano, D. E., & Courtois, E. T. (2022). Single-cell analysis of endometriosis reveals a coordinated transcriptional programme driving immunotolerance and angiogenesis across eutopic and ectopic tissues. *Nature Cell Biology*, 24(8), 1306–1318. <https://doi.org/10.1038/s41556-022-00961-5>

Thompson, R. E., Bouma, G. J., & Hollinshead, F. K. (2022a). The Roles of Extracellular Vesicles and Organoid Models in Female Reproductive Physiology. *International Journal of Molecular Sciences*, 23(6). <https://doi.org/10.3390/ijms23063186>

Thompson, R. E., Bouma, G. J., & Hollinshead, F. K. (2022b). The Roles of Extracellular Vesicles and Organoid Models in Female Reproductive Physiology. *International Journal of Molecular Sciences*, 23(6). <https://doi.org/10.3390/IJMS23063186>

Thompson, R. E., Johnson, A. K., Dini, P., Turco, M. Y., Prado, T. M., Premanandan, C., Burton, G. J., Ball, B. A., Whitlock, B. K., & Pukazhenth, B. S. (2020). Hormone-responsive organoids from domestic mare and endangered Przewalski's horse endometrium. *Reproduction*, 160(6), 819–831. <https://doi.org/10.1530/REP-20-0266>

Thompson, R. E., Meyers, M. A., Premanandan, C., & Hollinshead, F. K. (2023). Generation and cryopreservation of feline oviductal organoids. *Theriogenology*, 196, 167–173. <https://doi.org/10.1016/j.theriogenology.2022.11.020>

Thompson, R. E., Meyers, M. A., Pukazhenth, B. S., & Hollinshead, F. K. (2022). Evaluation of growth, viability, and structural integrity of equine endometrial organoids following cryopreservation. *Cryobiology*, 104, 56–62. <https://doi.org/10.1016/j.cryobiol.2021.11.003>

- Thompson, R. E., Meyers, M. A., Veeramachaneni, D. N. R., Pukazhenthil, B. S., & Hollinshead, F. K. (2022). Equine Oviductal Organoid Generation and Cryopreservation. *Methods and Protocols*, 5(3). <https://doi.org/10.3390/mps5030051>
- Tian, J., Yang, J., Chen, T., Yin, Y., Li, N., Li, Y., Luo, X., Dong, E., Tan, H., Ma, Y., & Li, T. (2023). Generation of Human Endometrial Assembloids with a Luminal Epithelium using Air-Liquid Interface Culture Methods. *Advanced Science (Weinheim, Baden-Wurtemberg, Germany)*, 10(30). <https://doi.org/10.1002/ADVS.202301868>
- Turco, M. Y., Gardner, L., Hughes, J., Cindrova-Davies, T., Gomez, M. J., Farrell, L., Hollinshead, M., Marsh, S. G. E., Brosens, J. J., Critchley, H. O., Simons, B. D., Hemberger, M., Koo, B. K., Moffett, A., & Burton, G. J. (2017). Long-term, hormone-responsive organoid cultures of human endometrium in a chemically defined medium. *Nature Cell Biology* 2017 19:5, 19(5), 568–577. <https://doi.org/10.1038/ncb3516>
- Venkata, V. D., Jamaluddin, M. F. B., Goad, J., Drury, H. R., Tadros, M. A., Lim, R., Karakoti, A., O'Sullivan, R., Ius, Y., Jaaback, K., Nahar, P., & Tanwar, P. S. (2022). Development and characterization of human fetal female reproductive tract organoids to understand Müllerian duct anomalies. *Proceedings of the National Academy of Sciences of the United States of America*, 119(30). <https://doi.org/10.1073/pnas.2118054119>
- Vollmuth, N., Schlicker, L., Guo, Y., Hovhannisyan, P., Janaki-Raman, S., Kurmasheva, N., Schmitz, W., Schulze, A., Stelzner, K., Rajeeve, K., & Rudel, T. (2022). c-Myc plays a key role in IFN- γ -induced persistence of *Chlamydia trachomatis*. *ELife*, 11. <https://doi.org/10.7554/eLife.76721>
- Wang, J., Du, H., Ma, L., Feng, M., Li, L., Zhao, X., & Dai, Y. (2023). MitoQ Protects Ovarian Organoids against Oxidative Stress during Oogenesis and Folliculogenesis In Vitro. *International Journal of Molecular Sciences*, 24(2). <https://doi.org/10.3390/ijms24020924>
- Wang, J., Fang, J., Feng, M., Li, L., Ma, L., Zhao, X., & Dai, Y. (2023). Inhibition of EED activity enhances cell survival of female germline stem cell and improves the oocytes production during oogenesis in vitro. *Open Biology*, 13(1). <https://doi.org/10.1098/rsob.220211>
- Wei, Y., Zhang, C., Fan, G., & Meng, L. (2021a). Organoids as Novel Models for Embryo Implantation Study. *Reproductive Sciences*, 28(6), 1637–1643. <https://doi.org/10.1007/s43032-021-00501-w>
- Wei, Y., Zhang, C., Fan, G., & Meng, L. (2021b). Organoids as Novel Models for Embryo Implantation Study. *Reproductive Sciences*, 28(6), 1637–1643. <https://doi.org/10.1007/s43032-021-00501-w>
- Wiwatpanit, T., Murphy, A. R., Lu, Z., Urbanek, M., Burdette, J. E., Woodruff, T. K., & Kim, J. J. (2020). Scaffold-Free Endometrial Organoids Respond to Excess Androgens Associated With Polycystic Ovarian Syndrome. *The Journal of Clinical Endocrinology and Metabolism*, 105(3). <https://doi.org/10.1210/CLINEM/DGZ100>
- Wu, G. M. J., Chen, A. C. H., Yeung, W. S. B., & Lee, Y. L. (2023). Current progress on in vitro differentiation of ovarian follicles from pluripotent stem cells. *Frontiers in Cell and Developmental Biology*, 11, 1166351. <https://doi.org/10.3389/fcell.2023.1166351>
- Xie, Y., Park, E. S., Xiang, D., & Li, Z. (2018). Long-term organoid culture reveals enrichment of organoid-forming epithelial cells in the fimbrial portion of mouse fallopian tube. *Stem Cell Research*, 32, 51–60. <https://doi.org/10.1016/j.scr.2018.08.021>
- Xu, Y., Cai, S., Wang, Q., Cheng, M., Hui, X., Dzakah, E. E., Zhao, B., & Chen, X. (2023). Multi-Lineage Human Endometrial Organoids on Acellular Amniotic Membrane for Endometrium Regeneration. *Cell Transplantation*, 32. <https://doi.org/10.1177/09636897231218408>
- Yang, M., Liu, Q., Chen, Y., Li, J., & He, W. (2024). Generation and Characterization of Rat Uterus Organoids from Rat Endometrial Epithelial Stem Cells. *Journal of Visualized Experiments : JoVE*, 210. <https://doi.org/10.3791/66928>
- Yang, W., Zhang, C., Wu, Y. H., Liu, L. B., Zhen, Z. Da, Fan, D. Y., Song, Z. R., Chang, J. T., Wang, P. G., & An, J. (2023). Mice 3D testicular organoid system as a novel tool to study Zika virus pathogenesis. *Virologica Sinica*, 38(1), 66–74. <https://doi.org/10.1016/j.virs.2022.10.001>

- Yang, Y., Huang, R., Cao, Z., Ma, S., Chen, D., Wang, Z., Feng, Y., Lei, Y., Zhang, Q., & Huang, Y. (2023). In vitro reconstitution of the hormone-responsive testicular organoids from murine primary testicular cells. *Biofabrication*, 15(1). <https://doi.org/10.1088/1758-5090/ac992a>
- Yu, B., McCartney, S., Strenk, S., Valint, D. J., Liu, C., Haggerty, C. L., & Fredricks, D. N. (2024). Vaginal Bacteria Elicit Acute Inflammatory Response in Fallopian Tube Organoids. *Reproductive Sciences*, 31(2), 505–513. <https://doi.org/10.1007/s43032-023-01350-5>
- Yucer, N., Holzapfel, M., Jenkins Vogel, T., Lenaeus, L., Ornelas, L., Laury, A., Sareen, D., Barrett, R., Karlan, B. Y., & Svendsen, C. N. (2017). Directed Differentiation of Human Induced Pluripotent Stem Cells into Fallopian Tube Epithelium. *Scientific Reports*, 7(1). <https://doi.org/10.1038/s41598-017-05519-2>
- Zhang, D., Jin, W., Cui, Y., & He, Z. (2024). Establishment and Characterization of Testis Organoids with Proliferation and Differentiation of Spermatogonial Stem Cells into Spermatocytes and Spermatids. *Cells*, 13(19). <https://doi.org/10.3390/cells13191642>
- Zhang, H., Xu, D., Li, Y., Lan, J., Zhu, Y., Cao, J., Hu, M., Yuan, J., Jin, H., Li, G., & Liu, D. (2022). Organoid Transplantation Can Improve Reproductive Prognosis by Promoting Endometrial Repair in Mice. *International Journal of Biological Sciences*, 18(6), 2627–2638. <https://doi.org/10.7150/ijbs.69410>
- Zhang, T., Zhang, M., Zhang, S., & Wang, S. (2024). Research advances in the construction of stem cell-derived ovarian organoids. *Stem Cell Research & Therapy*, 15(1), 505. <https://doi.org/10.1186/S13287-024-04122-3>
- Zhang, W., Cheng, Y., Zhang, S., Wei, R., & Zou, K. (2023). Application of Matrigel in the 3-dimension culture of female germline stem cells. *Reproductive Biology*, 23(3). <https://doi.org/10.1016/j.repbio.2023.100769>
- Zhang, X., Zhang, L., Li, T., Zhang, Z., Shang, X., Bai, H., Liu, Y., Zong, X., Shang, C., Song, D., Zhang, X., Fan, L., & Liu, Z. (2024). Investigating bacteria-induced inflammatory responses using novel endometrial epithelial gland organoid models. *Frontiers in Reproductive Health*, 6, 1490520. <https://doi.org/10.3389/frph.2024.1490520>
- Zhang, Y., Chen, W., Dong, X., Shang, W., Shao, S., & Zhang, L. (2024). Long-term maintenance of human endometrial epithelial organoids and their stem cell properties. *Reproductive Toxicology*, 123. <https://doi.org/10.1016/j.reprotox.2023.108522>
- Zhang, Y., Zhuang, Y., Zhou, J., Xie, X., Sun, M., Zheng, M., Yuan, K., Zhang, Z., & Zhang, J. (2024). Effect of estradiol after bacterial infection on the Wnt/ β -catenin pathway in bovine endometrium epithelial cells and organoids. *Theriogenology*, 219, 75–85. <https://doi.org/10.1016/j.theriogenology.2024.02.023>
- Zhou, W., Barton, S., Cui, J., Santos, L. L., Yang, G., Stern, C., Kieu, V., Teh, W. T., Ang, C., Lucky, T., Sgroi, J., Ye, L., & Dimitriadis, E. (2022a). Infertile human endometrial organoid apical protein secretions are dysregulated and impair trophoblast progenitor cell adhesion. *Frontiers in Endocrinology*, 13, 1067648. <https://doi.org/10.3389/fendo.2022.1067648>
- Zhou, W., Barton, S., Cui, J., Santos, L. L., Yang, G., Stern, C., Kieu, V., Teh, W. T., Ang, C., Lucky, T., Sgroi, J., Ye, L., & Dimitriadis, E. (2022b). Infertile human endometrial organoid apical protein secretions are dysregulated and impair trophoblast progenitor cell adhesion. *Frontiers in Endocrinology*, 13. <https://doi.org/10.3389/FENDO.2022.1067648>
- Zhu, M., Wang, N., Wang, S., Wang, Y., Yang, X., Fan, J., & Chen, Y. (2023). Effects of Follicular Fluid on Physiological Characteristics and Differentiation of Fallopian Tube Epithelial Cells Implicating for Ovarian Cancer Pathogenesis. *International Journal of Molecular Sciences*, 24(12). <https://doi.org/10.3390/ijms241210154>

Scientific considerations relevant to the 14-day rule

Scope: This topic looks at the characterisation of human embryos in early-stage development (up to 14 days). This includes research studying gastrulation, early organogenesis and neural development, advances in embryo culture systems (including optimisation and extended in vitro cultivation of non-human primates), and the use of stem cell-based embryos models. Papers considering the ethical and/or legal aspects of extending embryo culture beyond 14 days are additionally within scope.

Dimova, T., Alexandrova, M., Vangelov, I., You, Y., & Mor, G. (2024). The modeling of human implantation and early placentation: achievements and perspectives. *Human Reproduction Update*. <https://doi.org/10.1093/humupd/dmae033>

Ismaili M'hamdi, H., Rivron, N. C., & Asscher, E. C. (2024). Going high and low: on pluralism and neutrality in human embryology policy-making. *Journal of Medical Ethics*, 50(12), 846–854. <https://doi.org/10.1136/jme-2022-108515>

Kiya, Y., Watanabe, S., Harada, K., Yui, H., Yashiro, Y., & Muto, K. (2024). Attitudes of patients with IVF/ICSI toward human embryo in vitro culture beyond 14 days. *Regenerative Therapy*, 26, 831–836. <https://doi.org/10.1016/j.reth.2024.09.005>

M'hamdi, H. I., & de Wert, G. (2024). Reconsidering the 14-day rule in human embryo research: Advice from the Dutch Health Council. *Cell Stem Cell*, 31(11), 1560–1562. <https://doi.org/10.1016/j.stem.2024.09.019>

Pennings, G., Dondorp, W., Popovic, M., Chuva de Sousa Lopes, S., & Mertes, H. (2024). Ethical considerations on the moral status of the embryo and embryo-like structures. *Human Reproduction*, 39(11), 2387–2391. <https://doi.org/10.1093/humrep/deae228>

Stem cell-based embryo models

Scope: This topic is limited to research on stem cell derived embryo models and their precursors. It is separate from techniques used to derive embryonic and embryonic-like stem cells, focused on their application to develop models investigating early embryonic development.

Balubaid, A., Alsolami, S., Kiani, N. A., Cabrero, D. G., Li, M., & Tegner, J. (2024). A comparative analysis of blastoid models through single-cell transcriptomics. *IScience*, 27(11). <https://doi.org/10.1016/J.ISCI.2024.111122>

Choi, H. K., & Moon, S. H. (2024). Blastoid: The future of human development in the laboratory. *Cells & Development*, 180. <https://doi.org/10.1016/J.CDEV.2024.203975>

De Santis, R., Rice, E., Croft, G., Yang, M., Rosado-Olivieri, E. A., & Brivanlou, A. H. (2024). The emergence of human gastrulation upon in vitro attachment. *Stem Cell Reports*, 19(1), 41–53. <https://doi.org/10.1016/J.STEMCR.2023.11.005>

Farag, N., Sacharen, C., Avni, L., & Nachman, I. (2024). Coordination between endoderm progression and mouse gastruloid elongation controls endodermal morphotype choice. *Developmental Cell*, 59(17). <https://doi.org/10.1016/J.DEVCEL.2024.05.017>

Fernandez-Rial, C., & Fidalgo, M. (2024). Induction of Transient Morula-Like Cells in Mice Through STAT3 Activation. *Cellular Reprogramming*, 26(1), 8–9. <https://doi.org/10.1089/CELL.2023.0116>

Garge, R. K., Lynch, V., Fields, R., Casadei, S., Best, S., Stone, J., Snyder, M., McGann, C. D., Shendure, J., Starita, L. M., Hamazaki, N., & Schweppe, D. K. (2024). The proteomic landscape and temporal dynamics of mammalian gastruloid development. *BioRxiv: The Preprint Server for Biology*. <https://doi.org/10.1101/2024.09.05.609098>

Guo, M., Wu, J., Chen, C., Wang, X., Gong, A., Guan, W., Karvas, R. M., Wang, K., Min, M., Wang, Y., Theunissen, T. W., Gao, S., & Silva, J. C. R. (2024). Self-renewing human naïve pluripotent stem cells dedifferentiate in 3D culture and form blastoids spontaneously. *Nature Communications*, 15(1). <https://doi.org/10.1038/S41467-024-44969-X>

Hislop, J., Song, Q., Keshavarz F, K., Alavi, A., Schoenberger, R., LeGraw, R., Velazquez, J. J., Mokhtari, T., Taheri, M. N., Rytel, M., Chuva de Sousa Lopes, S. M., Watkins, S., Stolz, D., Kiani, S., Sozen, B., Bar-Joseph, Z., & Ebrahimkhani, M. R. (2024). Modelling post-implantation human development to yolk sac blood emergence. *Nature*, 626(7998), 367–376. <https://doi.org/10.1038/S41586-023-06914-8>

Huang, B., Peng, X., Zhai, X., Hu, J., Chen, J., Yang, S., Huang, Q., Deng, E., Li, H., Barakat, T. S., Chen, J., Pei, D., Fan, X., Chambers, I., & Zhang, M. (2024). Inhibition of HDAC activity directly reprograms murine embryonic stem cells to trophoblast stem cells. *Developmental Cell*, 59(16). <https://doi.org/10.1016/J.DEVCEL.2024.05.009>

- Kılıç, K. D., & Yılmaz, Z. S. (2024). Importance of In Vitro Embryo Model Procedure Standardization. *Journal of Clinical Laboratory Analysis*, 38(13–14). <https://doi.org/10.1002/JCLA.25082>
- Kwon, T. (2024). Role and ethics of cynomolgus monkey (*Macaca fascicularis*) blastoids in primate developmental biology research. *Journal of Medical Primatology*, 53(2). <https://doi.org/10.1111/JMP.12693>
- Li, H., Chang, L., Huang, J., & Silva, J. C. R. (2024a). Protocol for generating mouse morula-like cells resembling 8- to 16-cell stage embryo cells. *STAR Protocols*, 5(2). <https://doi.org/10.1016/J.XPRO.2024.102934>
- Li, H., Chang, L., Huang, J., & Silva, J. C. R. (2024b). Protocol for generating mouse morula-like cells resembling 8- to 16-cell stage embryo cells. *STAR Protocols*, 5(2). <https://doi.org/10.1016/J.XPRO.2024.102934>
- Linneberg-Agerholm, M., Sell, A. C., Redó-Riveiro, A., Perera, M., Proks, M., Knudsen, T. E., Barral, A., Manzanares, M., & Brickman, J. M. (2024). The primitive endoderm supports lineage plasticity to enable regulative development. *Cell*, 187(15), 4010–4029.e16. <https://doi.org/10.1016/J.CELL.2024.05.051>
- Liu, X., & Polo, J. M. (2024). Human blastoid as an in vitro model of human blastocysts. *Current Opinion in Genetics & Development*, 84. <https://doi.org/10.1016/J.GDE.2023.102135>
- Luijckx, D. G., Ak, A., Guo, G., van Blitterswijk, C. A., Giselbrecht, S., & Vrij, E. J. (2024). Monochorionic Twinning in Bioengineered Human Embryo Models. *Advanced Materials (Deerfield Beach, Fla.)*, 36(25). <https://doi.org/10.1002/ADMA.202313306>
- Luo, Y., An, C., Zhong, K., Zhou, P., Li, D., Liu, H., Guo, Q., Wei, W., Pan, H., Min, Z., Li, R., Yu, Y., & Fan, Y. (2024). Exploring the impacts of senescence on implantation and early embryonic development using totipotent cell-derived blastoids. *Journal of Advanced Research*. <https://doi.org/10.1016/J.JARE.2024.02.011>
- Luo, Y. X., & Yu, Y. (2024). Protocol for the Generation of Human EPS-Blastoids Using a Three-Dimensional Two-Step Induction System. *Methods in Molecular Biology (Clifton, N.J.)*, 2767, 27–41. https://doi.org/10.1007/7651_2022_471
- McNamara, H. M., Solley, S. C., Adamson, B., Chan, M. M., & Toettcher, J. E. (2024). Recording morphogen signals reveals mechanisms underlying gastruloid symmetry breaking. *Nature Cell Biology*, 26(11), 1832–1844. <https://doi.org/10.1038/S41556-024-01521-9>
- Pennarossa, G., Arcuri, S., Gandolfi, F., & Brevini, T. A. L. (2024). Generation of Artificial Blastoids Combining miR-200-Mediated Reprogramming and Mechanical Cues. *Cells*, 13(7). <https://doi.org/10.3390/CELLS13070628>
- Rawlings, T. M., Tryfonos, M., Makwana, K., Taylor, D. M., Brosens, J. J., & Lucas, E. S. (2024). Endometrial Assembloids to Model Human Embryo Implantation In Vitro. *Methods in Molecular Biology (Clifton, N.J.)*, 2767, 63–74. https://doi.org/10.1007/7651_2023_495
- Rufo, J., Qiu, C., Han, D., Baxter, N., Daley, G., & Wilson, M. Z. (2024). An explainable map of human gastruloid morphospace reveals gastrulation failure modes and predicts teratogens. *BioRxiv: The Preprint Server for Biology*. <https://doi.org/10.1101/2024.09.20.614192>
- Saadeldin, I. M., Ehab, S., Noreldin, A. E., Swelum, A. A. A., Bang, S., Kim, H., Yoon, K. Y., Lee, S., & Cho, J. (2024). Current strategies using 3D organoids to establish in vitro maternal-embryonic interaction. *Journal of Veterinary Science*, 25(3). <https://doi.org/10.4142/JVS.24004>
- Shibata, S., Endo, S., Nagai, L. A. E., Kobayashi, E. H., Oike, A., Kobayashi, N., Kitamura, A., Hori, T., Nashimoto, Y., Nakato, R., Hamada, H., Kaji, H., Kikutake, C., Suyama, M., Saito, M., Yaegashi, N., Okae, H., & Arima, T. (2024). Modeling embryo-endometrial interface recapitulating human embryo implantation. *Science Advances*, 10(8). <https://doi.org/10.1126/SCIADV.ADI4819>
- Tan, J. P., Liu, X., & Polo, J. M. (2024). Reprogramming fibroblast into human iBlastoids. *Nature Protocols*, 19(8). <https://doi.org/10.1038/S41596-024-00984-2>
- Wang, X., Sun, Y., Shi, H., & Xin, A. (2024). Establishment of an Embryo Implantation Model In Vitro. *Journal of Visualized Experiments: JoVE*, 208. <https://doi.org/10.3791/66873>

Wei, Y., Liu, K., Pinzon-Arteaga, C. A., Logsdon, D., Yu, L., Yuan, Y., & Wu, J. (2024). Generation of Human Blastoids from Naive Pluripotent Stem Cells. *Methods in Molecular Biology* (Clifton, N.J.), 2767, 1–18. https://doi.org/10.1007/7651_2023_485

Testicular tissue transplantation to restore fertility in males

Scope: This topic monitors developments in autologous and xenotransplantation of human testicular tissue (foetal, pre-pubertal and adult) with the intention of restoring fertility potential in males.

Jensen, C. F. S., Mamsen, L. S., Wang, D., Fode, M., Giwercman, A., Jørgensen, N., Ohl, D. A., Fedder, J., Hoffmann, E. R., Yding Andersen, C., & Sønksen, J. (2024). Results from the first autologous grafting of adult human testis tissue: A case report. *Human Reproduction*, 39(2), 303–309. <https://doi.org/10.1093/humrep/dead243>

Kourta, D., Camboni, A., Saussoy, P., Kanbar, M., Poels, J., & Wyns, C. (2024). Evaluating testicular tissue for future autotransplantation: focus on cancer cell contamination and presence of spermatogonia in tissue cryobanked for boys diagnosed with a hematological malignancy. *Human Reproduction*, 39(3), 486–495. <https://doi.org/10.1093/humrep/dead271>

Modanlou, M., Mahdipour, M., & Mobarak, H. (2024). Effectiveness of stem cell therapy for male infertility restoration: A systematic review. *Journal of Investigative Medicine : The Official Publication of the American Federation for Clinical Research*. <https://doi.org/10.1177/10815589241305317>

Sung, Z. Y., Liao, Y. Q., Hou, J. H., Lai, H. H., Weng, S. M., Jao, H. W., Lu, B. J., & Chen, C. H. (2024). Advancements in fertility preservation strategies for pediatric male cancer patients: a review of cryopreservation and transplantation of immature testicular tissue. *Reproductive Biology and Endocrinology*, 22(1). <https://doi.org/10.1186/s12958-024-01219-5>

von Rohden, E., Jensen, C. F. S., Andersen, C. Y., Sønksen, J., Fedder, J., Thorup, J., Ohl, D. A., Fode, M., Hoffmann, E. R., & Mamsen, L. S. (2024). Male fertility restoration: in vivo and in vitro stem cell-based strategies using cryopreserved testis tissue: a scoping review. *Fertility and Sterility*. <https://doi.org/10.1016/j.fertnstert.2024.07.010>